Inhibition of EGFR-STAT3 Signaling with Erlotinib Prevents Carcinogenesis in a Chemically-Induced Mouse Model of Oral Squamous Cell Carcinoma

Rebecca J. Leeman-Neill1, Raja R. Seethala1, Shivendra V. Singh2, Maria L. Freilino3, Joseph S. Bednash3, Sufi M. Thomas3, Mary C. Panahandeh3, William E. Gooding4, Sonali C. Joyce5, Mark W. Lingen6, Daniel B. Neill5, and Jennifer R. Grandis1,2,3

Abstract

Chemoprevention of head and neck squamous cell carcinoma (HNSCC), a disease associated with high mortality rates and frequent occurrence of second primary tumor (SPT), is an important clinical goal. The epidermal growth factor receptor (EGFR)-signal transducer and activator of transcription (STAT)-3 signaling pathway is known to play a key role in HNSCC growth, survival, and prognosis, thereby serving as a potential therapeutic target in the treatment of HNSCC. In the current study, the 4-nitroquinoline-1-oxide (4-NQO)–induced murine model of oral carcinogenesis was utilized to investigate the chemopreventive activities of compounds that target the EGFR-STAT3 signaling pathway. This model mimics the process of oral carcinogenesis in humans. The drugs under investigation included erlotinib, a small molecule inhibitor of the EGFR, and guggulipid, the extract of an Ayurvedic medicinal plant, which contains guggulsterone, a compound known to inhibit STAT3. Dietary administration of guggulipid failed to confer protection against oral carcinogenesis. On the other hand, the mice placed on erlotinib-supplemented diet exhibited a 69% decrease ($P < 0.001$) in incidence of preneoplastic and neoplastic lesions compared with mice on the control diet. Immunostaining of dysplastic lesions demonstrated modest decreases in STAT3 levels, with both drug treatments, that were not statistically significant. The results of the present study provide the basis for exploring the efficacy of erlotinib for prevention of HNSCC in a clinical setting. Cancer Prev Res; 4(2); 230–7. ©2010 AACR.

Introduction

Head and neck squamous cell carcinoma (HNSCC) is a devastating disease associated with a mortality rate of approximately 50% that has not changed for several decades (1). In addition to high rates of recurrence, HNSCC is associated with frequent formation of second primary tumor (SPT), in 3% to 7% per year, among the highest for any malignancy (2). The high rate of SPT has been attributed to the occurrence of “field cancerization,” a term coined by Slaughter and colleagues to define global molecular changes to the upper airway mucosa upon exposure to carcinogens (3). HNSCC, therefore, is a particularly appropriate target for chemoprevention. Chemopreventive agents may serve as appropriate therapy for patients with a premalignant lesion or patients who have had HNSCC and are at high risk for recurrence and/or SPT. If the agent is safe and devoid of major side effects, it may also be considered for use in primary prevention of HNSCC.

On the basis of early observations that tumors of the upper aerodigestive tract and lungs occurred more frequently in cattle who were deficient in vitamin A (4), studies investigating high-dose retinoids as chemopreventive therapy for HNSCC demonstrated efficacy in delaying carcinogenesis in humans but were associated with considerable toxicity (5, 6). Since then, definitive trials involving administration of tolerable doses of retinoids have not consistently demonstrated prevention of HNSCC (7, 8). The need for identification of novel approaches to prevent HNSCC has become apparent. Recent clinical trials demonstrating chemoprevention of HNSCC have employed green tea (9–11) and Bowman-Birk inhibitor derived from soybeans (12). Studies investigating potential roles for COX-2 and epidermal growth factor receptor (EGFR) inhibition in HNSCC chemoprevention are also currently underway (13). Other compounds that have demonstrated efficacy in preventing carcinogenesis in the preclinical model of...
carcinogenesis used in the current study include rapamycin (14) and ABT-510, an inhibitor of angiogenesis (15).

In the current study, we focused on targeting the EGFR-STAT3 signaling pathway. EGFR, one of the ErbB family of receptors, which is overexpressed in over 80% of HNSCC tumors, is a marker of poor prognosis in patients with HNSCC (16). A recent study has found that EGFR gene copy number can be used to predict progression from oral premalignant lesions to oral squamous cell carcinoma (17). Erlotinib (Tarceva) is an EGFR-targeting tyrosine kinase inhibitor (TKI) that has shown promise in clinical trials with HNSCC and is currently in advanced stages of clinical testing for treatment of this disease. The chemopreventive activity of the EGFR inhibitor, gefitinib, has recently been demonstrated in an animal model of lung cancer (18). Similar observations in an animal model of HNSCC would provide further preclinical basis for ongoing clinical trials investigating EGFR-inhibiting agents as chemopreventive therapy in HNSCC (19–21). STAT3 mediates signaling through the EGFR, regulating the expression of genes that control the cell cycle, and apoptosis (22). STAT3 is constitutively active in the majority of HNSCC tumors where it has been found to play an important role in growth and survival in preclinical models of HNSCC as well as in cellular transformation (23). Various agents that have STAT3 inhibitory properties have been studied for their potential use in treatment of HNSCC, though none have been approved for clinical use (22). Guggulipid is a natural product extracted from the Commiphora mukul plant and containing guggulsterone, a compound with a steroid-like structure (24). This compound has been found to have anticancer activity in various preclinical models (25), decreases levels of STAT3 in HNSCC cell lines and tumor xenografts, inhibits tumor growth, and enhances the antitumor activity of EGFR-targeted therapy (26). Furthermore, guggulsterone’s safety in clinical trials (27) and availability in inexpensive clinical formulations, such as guggulipid (Sabinsa Corporation), makes it a plausible candidate for use in chemoprevention.

The myriad challenges and costs associated with clinical trials of chemopreventive agents underscore the importance of preclinical studies to provide a biological rationale for use of a given agent as chemopreventive therapy. Although in vitro studies can be important in identifying molecular alterations involved in carcinogenesis that can be targeted by a specific agent, true preclinical evidence of cancer prevention, however, requires in vivo investigation using a model that recapitulates the process of carcinogenesis in humans. 4-nitroquinoline-1-oxide (4-NQO) is a carcinogen that induces formation of DNA adducts. Oral administration of 4-NQO to mice has been shown to result in the formation of dysplastic lesions and, eventually, neoplasia (28). In addition, the resulting oral squamous cell carcinomas have been found to resemble human HNSCC both histologically and in terms of molecular changes that often characterize human HNSCCs, including changes in levels of EGFR and the cell-cycle regulator p16 (29). This carcinogen has been used to induce HNSCC in rats and mice, through topical application. More recently, optimal tumor formation has been attained through administration in the animals’ drinking water (29).

In the current study, we investigated the chemopreventive properties of both the EGFR-targeting agent, erlotinib, and a natural product with STAT3 inhibitory activity, guggulipid, using the 4-NQO–induced mouse model of oral carcinogenesis. We found that, whereas guggulipid was not effective in inhibiting oral carcinogenesis, erlotinib decreased formation of oral dysplastic lesions.

Materials and Methods

Reagents and animal diets

4-NQO (Sigma Chemical) was dissolved in 100% DMSO (final concentration 50 mg/mL) and kept at −20°C. Guggulipid [solid, 2.65% guggulsterones by high-performance liquid chromatography (HPLC)] was generously provided by Sabinsa Corporation and erlotinib by OSI Pharmaceuticals. Custom erlotinib-supplemented (300 mg/kg) and guggulipid-supplemented diets (28.3 g/kg, equivalent of 750 mg/kg of guggulsterone) were prepared by Harlan Teklad, as an 18% protein rodent diet. Assuming diet consumption of 3 g/mouse/d (based on prior studies; data not shown) and average mouse weight of 22.5 g, mice received approximately 3.77 g/kg of guggulipid (equivalent to 100 mg/kg guggulsterone) or 40 mg/kg of erlotinib per day. Diets were stored in vacuum-sealed bags at 4°C.

Study design and statistics

The study was designed to detect a 25% or greater difference in incidence of preneoplastic and neoplastic lesions in the treated mice versus controls with a power of 0.85. This design required 75 mice per group, relying on the presence of dysplastic or neoplastic lesions in at least 80% of the control group at the end of the experiment. On the basis of prior studies, in which approximately 5% of mice were expected to die from 4-NQO toxicity (data not shown), 80 mice were used per treatment group. In addition, because the kinetics of the 4-NQO model had not been conclusively determined, an adaptive study design was employed. Fifty extra mice were added to the control group so that 10 each could be sacrificed at various time points. If 10 of 10 mice had preneoplastic or neoplastic lesions, there was a probability of 0.89 that at least 80% of the remaining mice in the control group should have lesions as well. The incidences of lesions in the treatment groups were compared using a 2-sided Fisher’s exact test.

In immunohistochemistry (IHC) studies, quantitative scores (described later in the text) were assigned to each histologic section and groups compared using a Wilcoxon test.

Animal treatments

Female CBA/J mice (5–6 weeks; Jackson Laboratories) were treated with either the control diet or one of the drug-containing diets for 2 weeks prior to initiating 4-NQO
Results

Toxicity of combining 4-NQO and erlotinib or guggulipid

To maximize exposure to erlotinib and guggulipid and attempt primary prevention of oral carcinogenesis, mice were treated with drug-supplemented diets for 2 weeks prior to 4-NQO administration. Continued administration of the special diets was intended but precluded by unexpected toxicity occurring with the combination of drug-containing diets and 4-NQO. After 1.5 weeks of 4-NQO treatment combined with the special diets, 28 of 80 mice in the erlotinib group and 5 of 80 mice in the guggulipid group died suddenly (Fig. 2A). The average weight of mice in both groups dropped (Fig. 2B). Deaths did not occur in the groups receiving either 4-NQO water alone (control diet) or the special diets without 4-NQO. The deaths, therefore, resulted from unforeseen toxicity of combining 4-NQO with either guggulipid or erlotinib. Drug-containing diets were promptly discontinued in mice receiving 4-NQO treatments and resumed after completion of the 8 weeks of 4-NQO treatment. Although the death rate slowed dramatically and average weight increased after withdrawal of the drug-containing diets, more mice died over the course of the experiment (Fig. 2A). Necropsy of one of the mice that died after combining 4-NQO with erlotinib indicated hemorrhagic gastroenteritis (data not shown).

Kinetics of lesion development in this model

Examination of mouse tongues at 5 weeks after completion of 4-NQO treatments demonstrated that 9 of 129 mice in the control group and 5 of 75 in the guggulipid group (~7% of each) had oral tumors that were visible on gross examination (Fig. 3A). No tumors were seen in the erlotinib group at this point. Beginning at 5 weeks after 4-NQO treatment, mice were sampled, weekly, from the control group and H&E-stained sections of their tongues examined for the presence of dysplastic and neoplastic lesions (Table 1). By 7 weeks post–4-NQO treatment, 10 of 10 mice had dysplastic or neoplastic lesions. The rationale for...
Toxicity of combining 4-NQO with drug-containing diets. A, percentages of mice that died, without being purposely sacrificed, during the study, are plotted. B, animals were weighed approximately once per week throughout the study.

Figure 3A shows the gross appearance of representative tumors. The histologic appearances of representative lesions are shown in Figure 3B to E.

Erlotinib, but not guggulipid, prevents progression beyond mild oral dysplasia in 4-NQO–treated mice.

Tongues harvested from the mice sacrificed at 8 weeks post-4-NQO treatment were paraffin embedded and sectioned completely onto glass slides. Every tenth slide was H&E stained and examined by an investigator who was blinded to treatment group. The study was designed to compare the incidence of lesions, both preneoplastic and neoplastic versus numbers of normal appearing tongues occurring in each group. Each mouse was classified according to the highest grade lesion found on its tongue. During blinded histologic examination, however, it was clear that the vast majority of mice had at least mild dysplasia. It was decided, at this point, that mild dysplasia, a lesion that is not necessarily preneoplastic in humans, should be considered to be in the same category as normal appearing tongues in this model. Indeed, at the end of the study, it was discovered that the only mice without at least mild dysplasia (i.e., completely normal) were the few (4 per group) treated with drug-containing diets but who never received 4-NQO. Figure 3A shows the gross appearance of representative tumors. The histologic appearances of representative lesions are shown in Figure 3B to E.

Figure 4 shows the incidence of dysplastic and neoplastic lesions in each group. 78.9% of the mice in the control group were found to have lesions that were higher grade than mild dysplasia, whereas only 24.4% of the erlotinib-treated group had such lesions, a 69% decrease overall ($P < 0.001$). 80.6% of the mice in the guggulipid-treated group had lesions that were higher grade than mild dysplasia. This very small increase over the incidence in the control group was not statistically significant.

The incidence of invasive SCC, specifically, did not necessarily fall into the same pattern. SCC was observed on tongues of 14.7% of mice in the control group, 12.19% in the erlotinib group, and 27.5% in the guggulipid group. This is explained, in part, by the very small numbers of mice with SCC (e.g., only 5 SCCs in the erlotinib group), making such a comparison unreliable. In addition, many of the SCCs were noted to be very small, only captured on 1 slide, for example. In general, moderate and severe dysplasia were also detected on tongues with SCC, further supporting the rationale for combining all lesions greater than mild dysplasia into the same category upon analysis.

Immunostaining of selected lesions

As inhibitors of the EGFR-STAT3 signaling axis were tested in this study, select tongues with an approximately even distribution of dysplasia and SCC in each group (total of 30 in the control group, 10 in the erlotinib group, and 20 in the guggulipid group) were stained, by IHC, with an antibody specific for STAT3. Again, because several lesions were very small, some of the lesions seen on H&E staining were not seen on immunostained slides, despite use of immediately adjacent sections in H&E and immunostaining. Numbers of stained SCCs, in the erlotinib group in particular, were very small (n = 3), such that IHC data comparing SCCs were unreliable and showed large variance. In comparing dysplastic lesions, on the other hand, mild decreases, that were not statistically significant, were seen in expression of STAT3 in both erlotinib and guggulipid treatment groups compared with vehicle (Fig. 5). The lack of statistical significance, again, may be due to the low numbers of dysplastic lesions in each group (vehicle: $n = 22$, erlotinib: $n = 7$, guggulipid: $n = 12$). In addition, the stain intensity did not vary greatly between sections, so that all sections, regardless of group, received scores between 70 and 180, despite a possible scoring range of 0 to 300, contributing to small quantitative differences between groups.

In addition to examining STAT3, we also stained these sections for pMAPK and pAKT, proteins also downstream of EGFR. However, no significant decrease in pMAPK or pAKT was seen in the erlotinib-treated group compared with controls, as examined by IHC. Nor did expression of...
these proteins trend significantly over time during carcinogenesis, as observed comparing levels in the tissue from control group mice sampled at 5, 6, and 7 weeks after 4-NQO treatment (data not shown).

**Discussion**

The current study examined guggulipid and erlotinib as potential chemopreventive therapies in HNSCC. Erlotinib has been shown to be an active agent in the treatment of HNSCC (30). Guggulipid has demonstrated anticancer activity in preclinical models of HNSCC (26). The
4-NQO–induced model of oral carcinogenesis provides us with the opportunity to examine the chemopreventive activities of these agents. Erlotinib was found to significantly inhibit progression from mild dysplasia to premalignant and malignant lesions, with rates of these lesions decreasing by 69% compared with the control group. Guggulipid, on the other hand, did not inhibit oral carcinogenesis in this model.

The 4-NQO–induced mouse model of oral SCC has been used to investigate the chemopreventive properties of several agents (14, 15, 28, 29). As 4-NQO induces the formation of DNA adducts and eventually dysplastic and neoplastic lesions that histologically and molecularly resemble those seen in human head and neck carcinogenesis (28, 29), it was thought that this model would be appropriate for use in testing potential chemopreventive agents. Whether or not results seen in this model are translatable, in terms of identifying chemopreventive therapies that will be effective in humans, has yet to be determined in clinical trials. Ongoing studies investigating erlotinib’s chemopreventive activity in humans may contribute information about the translatability of results found in using the 4-NQO model (19, 20).

In the current study, erlotinib and guggulipid were administered in animal diets. This method of administration has been employed in various studies investigating potential chemopreventive agents and has also been used, previously, to deliver guggulipid, showing effects on markers of guggulipid’s cholesterol lowering and antidiabetic activities (31, 32). The observed inhibition of carcinogenesis with the erlotinib-containing diet and the fact that, when administered simultaneously with 4-NQO, both drug-containing diets conferred increased toxicity over the carcinogen alone, confirms that each drug was delivered in the diets. Indeed, compared with oral gavage, which bypasses much of the oral cavity, administering each drug in the diet may have allowed for better exposure to the animals’ oral mucosa. In addition, daily oral gavage over the extended duration of this study was not permitted by the Institutional Animal Care and Use Committee (IACUC), as it would have been impossible to continue the study for the extended duration required.

Figure 4. Incidence of premalignant and malignant lesions in control-, erlotinib-, and guggulipid-treated groups. H&E-stained sections of tongues harvested from mice in each group were examined for the presence of premalignant (moderate and high-grade dysplasia) plus malignant lesions. The group treated with erlotinib demonstrated a decrease in numbers of such lesions ($P < 0.001$).

Figure 5. STAT3 levels in 4-NQO–induced dysplasia. Photomicrographs of the dysplastic lesions with the median immunostaining score for each group, occurring in tongues of animals from the vehicle-treated (A), erlotinib-treated (B), and guggulipid-treated (C) groups stained for STAT3. D, graphical description of immunostaining scores for each group ($P = 0.11$ for vehicle vs. erlotinib and $P = 0.41$ for vehicle vs. guggulipid).
resulted in considerable oral and pharyngeal irritation in the animals, presenting a safety hazard. The dose of erlotinib added to the diet was based on that found to be tolerated, in the long term, in unpublished studies (Dr. Ronald Lubet, NIH, personal communication). The dose of guggulipid chosen was based on that found to have anticancer and STAT3-inhibiting activity in a xenograft model of HNSCC (26) and also based on HPLC data, from Sabinsa Corporation, describing the concentration of guggulsterone in guggulipid. In the current study, administration of drug-containing diets was started prior to the carcinogen to maximize delivery of each potential chemopreventive agent and attempt to determine their maximal chemopreventive activity in vivo. This method does not resemble the time-frame for potential human usage of a chemopreventive therapy. The translational potential of future studies may be enhanced by administering the preventive agent after the initiation of carcinogenesis. For similar reasons, the study was terminated at a time point when most of the animals in the control group had premalignant, rather than malignant lesions, to avoid waiting until most animals in control and treatment groups had carcinoma, potentially obscuring a small chemopreventive effect.

The EGFR-STAT3 signaling axis is known to play a key role in growth and survival of HNSCC (33). Expression of EGFR has been used to predict progression from premalignant lesions to oral SCC (17). Furthermore, levels of EGFR are elevated in the 4-NQO–induced model, similar to human HNSCC (29). This led us to hypothesize that both erlotinib, which inhibits EGFR and guggulipid, containing guggulsterone, which inhibits STAT3, a molecule downstream of EGFR, would prevent oral carcinogenesis in this model.

The study did demonstrate the chemopreventive activity of erlotinib. In this group, 69% fewer mice developed malignant or premalignant lesions compared with vehicle. Only modest decreases, that were not statistically significant, in the molecular target of interest, STAT3, were seen in the control group had premalignant, rather than malignant lesions, to avoid waiting until most animals in control and treatment groups had carcinoma, potentially obscuring a small chemopreventive effect.

The current study justifies targeting the EGFR as a strategy for chemoprevention of HNSCC. Indeed, various clinical studies investigating this potential use for EGFR inhibitors are currently underway (19–21). Erlotinib’s known toxicity presents a challenge in terms of determining appropriate doses for long-term administration in patients who are at risk for HNSCC but do not currently have HNSCC. If erlotinib is found to have chemopreventive activity in humans, it will, however, represent an important therapeutic opportunity for patients who, either due to exposure history, premalignant lesion, or a prior diagnosis of HNSCC, are at high risk for this frequently fatal malignancy for which there is currently no approved preventive therapy.

**Disclosure of Potential Conflicts of Interest**

No potential conflicts of interest were disclosed.

**Grant Support**

This project is supported by grants from the NIH: 1F30 ES015669 to R.J. Leeman-Neill and P50 CA097190 and an American Cancer Society Clinical Research Professorship to J.R. Grandis.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received September 20, 2010; revised November 5, 2010; accepted December 9, 2010; published OnlineFirst December 16, 2010.
References


Inhibition of EGFR-STAT3 Signaling with Erlotinib Prevents Carcinogenesis in a Chemically-Induced Mouse Model of Oral Squamous Cell Carcinoma


Updated version
Access the most recent version of this article at:
doi:10.1158/1940-6207.CAPR-10-0249

Cited articles
This article cites 33 articles, 17 of which you can access for free at:
http://cancerpreventionresearch.aacrjournals.org/content/4/2/230.full.html#ref-list-1

Citing articles
This article has been cited by 8 HighWire-hosted articles. Access the articles at:
/content/4/2/230.full.html#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.