Inhibition of 15-Hydroxyprostaglandin Dehydrogenase by Helicobacter pylori in Human Gastric Carcinogenesis

Yeon-Mi Ryu1, Seung-Jae Myung1,2, Young Soo Park3, Dong-Hoon Yang1,2, Ho June Song1,2, Jin-Yong Jeong1, Sun Mi Lee1, Miyeoun Song1, Do Hoon Kim1,2, Hyo-Jeong Lee1,2, Soo-Kyung Park1,2, Stephen P. Fink4, Sandy D. Markowitz4, Kee Wook Jung1,2, Kyung-Jo Kim1,2, Byong Duk Ye1,2, Jeong-Sik Byeon1,2, Hwoon-Yong Jung1,2, Suk-Kyun Yang1,2, and Jin-Ho Kim1,2

Seoul, Korea; and4Department of Medicine, Case Comprehensive Cancer Asan Institute for Life Sciences; Departments of2Gastroenterology and 3Pathology, University of Ulsan College of Medicine, Asan Medical Center, Olympic-Ro, 43-Gil, Songpa-gu, Seoul 138-736, Korea. Phone: 822-3010-3917; Fax: 822-476-0824; E-mail: sjmyung@amc.seoul.kr

Note: Supplementary data for this article are available at Cancer Prevention Research Online (http://cancerpreventionresearch.aacrjournals.org/).

Introduction

Although the incidence of gastric cancer is declining, it remains the fourth most common cancer and the second leading cause of cancer-related deaths in the world (1). Gastric carcinogenesis is a multifactorial process that involves complex interactions between host and environmental factors. Helicobacter pylori (H. pylori) is one of the most important environmental risk factors for gastric malignancies (2). The chronic inflammation that develops in response to this organism contributes to tumor cell proliferation and metastasis and affects survival (3, 4). Prostaglandins play an important role in the growth and stimulation of the inflammation-associated gastric carcinogenesis that is associated with H. pylori infection (5–7). In particular, the expression of COX-2, a rate-limiting enzyme for prostaglandin biosynthesis, is induced in H. pylori–associated carcinogenesis and results in the induction of the proinflammatory prostaglandin, PGE₂ (8, 9).

PGE₂ levels are regulated not only by its synthesis but also by its degradation (8). The key enzyme responsible for the biologic inactivation of prostaglandins is NAD⁺-dependent 15-hydroxyprostaglandin dehydrogenase (15-PGDH; ref. 10). 15-PGDH inactivates PGE₂ by catalyzing the oxidation of its 15(S)-hydroxyl group, which results in the formation of inactive 15-keto metabolites (10, 11). Therefore, 15-PGDH plays an important role in regulating the local concentration of PGE₂. Studies by our group and others have shown that while 15-PGDH is highly expressed by normal colonic epithelial cells, transcription of 15-PGDH mRNA is lost in most colon cancers (11, 12). Follow-up research showed that 15-PGDH may have a potential tumor suppressor function in lung, breast,
bladder, thyroid, pancreas (13–17), and gastric cancers (18, 19). Previously, we have shown that 15-PGDH expression was lost in 70.1% of malignant human gastric tissues and 83.3% of adenomas but was preserved in normal and metaplastic gastritis (19). In addition, we found that the repression of 15-PGDH in gastric cancer may be due to the activation of the EGFR/ERK1/2 pathway (19). However, it is not known whether 15-PGDH plays any role in H. pylori–associated gastric carcinogenesis. We hypothesized that H. pylori infection may suppress 15-PGDH expression, thereby elevating PGE2 levels, which in turn promote gastric carcinogenesis.

Toll-like receptors (TLR) are surface-exposed pattern recognition receptors. The binding of a microbial antigen to the TLRs activates their interaction with MyD88 (myeloid differentiation primary response protein-88; ref. 20), which triggers intracellular signals that induce the expression of inflammatory cytokines, including NF-kB; this in turn augments antiapoptotic proteins and thereby promotes the invasion and metastasis of cancer (21). In addition, TLR4/MyD88 activation by H. pylori may promote stromal macrophage activation that induces the COX2/PGE2 pathway (22). Accordingly, it is also possible that the TLR4/MyD88 pathway mediates the suppression of 15-PGDH in H. pylori infection–associated gastric carcinogenesis that elevates PGE2 levels.

The present study revealed that H. pylori infection suppresses 15-PGDH transcription, which suggests that the infection decreases the degradation of PGE2, the major procarcinogenic prostaglandin, which in turn promotes gastric carcinogenesis. The suppression of 15-PGDH was mediated by the activation of TLR4/MyD88 and Snail transcriptional regulation by extracellular signal–regulated kinase (ERK)1/2 and EGF receptor (EGFR). In addition, H. pylori–infected gastric specimens exhibited more 15-PGDH DNA promoter methylation than negative specimens, which suggests that epigenetic modulation contributes to the suppression of 15-PGDH transcription. Finally, 15-PGDH expression was restored after H. pylori eradication, which suggests that the inhibition of 15-PGDH transcription is a reversible process. These results indicate that the elevated PGE2 levels in H. pylori–associated gastric carcinogenesis are likely to be the consequence of both reduced catabolism and increased synthesis.

Materials and Methods

Study population

The study population consisted of 40 participants who underwent upper gastrointestinal endoscopy at Asan Medical Center (Seoul, Korea) between June 2011 and November 2011. Their median age was 56 years (range, 21–79 years) and 26 participants were male (65.0%). The exclusion criteria were as follows: (i) consumption of a nonsteroidal anti-inflammatory drug and drugs that affect gastric acid secretion such as antacids, bismuth compounds, H2 receptor antagonists, and proton pump inhibitors in 6 weeks, (ii) previous gastric surgery, (iii) a history of H. pylori infection or eradication therapy, and/or (iv) a known allergy to antibiotics. Of the 40 participants, 32 lacked symptoms and underwent an endoscopy for a routine check-up, whereas the remaining 8 had gastrointestinal symptoms including dyspepsia, acid regurgitation, or epigastric pain (Table 1). Of the 25 patients who were infected with H. pylori, 4 were treated with the standard triple H. pylori eradication therapy for 7 days (proton pump inhibitor 40 mg twice daily, clarithromycin 0.5 g twice daily, and amoxicillin 1 g twice daily). At least 4 weeks later, they underwent follow-up endoscopy and sample collection.

Sample collection and determination of H. pylori infection

At baseline, the subjects underwent endoscopy with biopsies for histology (2 samples from the antrum) and a rapid urease test (1 sample from the antrum; Hp kit; Chongkundang Pharm. Corp.). In addition, 3 more biopsies were taken for DNA, RNA, and protein analysis. The biopsies were frozen in liquid nitrogen immediately and stored at −70°C until processing. Endoscopy (Endoscopy model: CF-H2160AL; Olympus) was conducted by the same physician (S.-J. Myung) after sedation with intravenous midazolam. Histologic examinations were conducted by one experienced specialist gastrointestinal pathologist (Y.S. Park), who used hematoxylin and eosin (H&E) staining for the assessment of gastritis and Gram staining for the detection of H. pylori. At entry, patients were considered H. pylori–positive if the results of 2 tests (histologic analysis and rapid urease test) were positive. Follow-up sample collection and processing after H. pylori eradication was the same as baseline collection. All processes relating to human tissue sampling were conducted under the approval of the Asan Medical Center Institutional Review Board.

Quantitative real-time PCR

Total RNA was extracted by using the RNeasy Kit (Ambion). The concentration and quality of the RNA samples were determined by using a Nanodrop-1000 spectrophotometer, and cDNA was generated by using MultiScribe Reverse Transcriptase (Applied Biosystems), and 40 ng of cDNA was used for real-time PCR assays for 15-PGDH and COX-2. All samples had an optical density (OD) 260/280 ratio greater than 1.8. Quantitative real-time PCR was carried out with Applied Biosystem 7900 Real-time PCR systems. Real-time PCR of 15-PGDH and COX-2 was carried out by using the human 15-PGDH TaqMan Probe/Primer Kit Hs 00168359_m1 and the human COX-2 TaqMan Probe/Primer kit Hs 00153133_m1 (Applied Biosystems). β-Antin was amplified by using the human ACTB TaqMan primer/probe kit Hs 99999903_m1 (Applied Biosystems). The relative gene expression was normalized to mRNA of β-actin and calculated by 2−ΔΔCt method.

Western blot analysis

Human tissue lysates were prepared by homogenization using the TissueLyser II System in cell lysis buffer (Cell Signaling Technology) with 1 mmol/L phenylmethylsulfonyl fluoride (PMSF). The protein content was measured by
the BCA Method using the Bicinchoninic Acid Protein Assay Kit (GenDEPOT) and a SmartSpec 3000 microplate reader (Molecular Devices). Protein samples (30 µg) were separated on 12% PAGE and transferred to Immobilon-P Membrane (Millipore). The primary antibodies were polyclonal rabbit antibodies against 15-PGDH (1:5,000; Novus Biologicals), Snail (1:1,000; Santa Cruz), and glyceraldehyde-3-phosphate dehydrogenase (GAPDH; 1:5,000; Cell Signaling). The secondary antibody was a goat anti-rabbit antibody (1:5,000; Cell Signaling). Detection was conducted by using the ECL Plus Detection System (Amersham Pharmacia) according to the manufacturer’s instructions and then exposing the gel to Kodak CP-BU film (Kodak).

The cells were washed with PBS and lysed with cell lysis buffer with 1 mmol/L PMSF. Western blot analysis was conducted as described above. The primary antibodies were polyclonal anti-15-PGDH (1:5,000; Novus Biologicals); anti-Snail 1 (1:1,000; Santa Cruz); and glyceraldehyde-3-phosphate dehydrogenase (GAPDH; 1:5,000; Cell Signaling). The secondary antibody was a goat anti-rabbit antibody (1:5,000; Cell Signaling). Detection was conducted by using the ECL Plus Detection System (Amersham Pharmacia) according to the manufacturer’s instructions and then exposing the gel to Kodak CP-BU film (Kodak).

Immunohistochemistry

The H. pylori–negative and -positive tissues were subjected to immunohistochemical staining for 15-PGDH by using a monoclonal anti-15-PGDH antibody (provided by S.D. Markowitz; ref. 12). The tissues were fixed in 10% formalin and embedded in paraffin. Four-micrometer-thick sections were deparaffinized with xylene and then gradually rehydrated with a graded series of alcohol solutions. 15-PGDH staining was conducted as described previously (19). In short, antigen retrieval was conducted (60 minutes at 96°C) in 10 mmol/L citrate buffer (pH 6.0). The primary anti-15-PGDH (1:5,000 dilution) antibody was incubated overnight at 4°C. The slides were incubated with Dako Envision System Kit (DAKO) and developed with DAB solution (Dako). Finally, the sections were counterstained with hematoxylin. All slides were reviewed by one gastrointestinal pathologist.

Bisulfite modification and methylation-specific PCR

Genomic DNA was isolated from gastric tissues by using a DNeasy tissue extraction kit (Qiagen) and then subjected to bisulfate modification with an EZ DNA Methylation-Gold kit (Zymo Research). The modified DNA was amplified with primers that were specific for either the unmethylated (U) or methylated (M) sequences of 15-PGDH (18). Methylation and unmethylated human control DNA was purchased from Qiagen. Methylation-specific PCR (MSP) was individually carried out in 25 µL reaction volumes by using the EpiTect MSP Kit (Qiagen). The products were separated on a 2% agarose gel and detected by ethidium bromide staining.

Pyrosequencing analysis

Each primer was designed by using the PSQ assay design program (Biotage). The primer sequences that were used are listed in Supplementary Table S1. The amplification was

Table 1. Characteristics of the H. pylori–positive and -negative study participants

<table>
<thead>
<tr>
<th>Variables</th>
<th>H. pylori-negative (n = 15)</th>
<th>H. pylori-positive (n = 25)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, median, y</td>
<td>52 (21–66)</td>
<td>63 (36–79)</td>
<td>0.002*</td>
</tr>
<tr>
<td>Sex, male, n (%)</td>
<td>9 (60.0%)</td>
<td>17 (68.0%)</td>
<td>0.61</td>
</tr>
<tr>
<td>Current smoking, n (%)</td>
<td>4 (26.7%)</td>
<td>6 (24.0%)</td>
<td>1.00</td>
</tr>
<tr>
<td>Alcohol consumption, n (%)</td>
<td>4 (26.7%)</td>
<td>12 (48.0%)</td>
<td>0.18</td>
</tr>
<tr>
<td>Hypertension, n (%)</td>
<td>4 (26.7%)</td>
<td>11 (44.0%)</td>
<td>0.27</td>
</tr>
<tr>
<td>Diabetes, n (%)</td>
<td>3 (20.0%)</td>
<td>7 (28.0%)</td>
<td>0.72</td>
</tr>
<tr>
<td>BMI, mean ± SD, kg/m²</td>
<td>24.57 ± 2.81</td>
<td>24.50 ± 3.65</td>
<td>0.94</td>
</tr>
<tr>
<td>Cholesterol, mean ± SD, mg/dL</td>
<td>181.33 ± 33.35</td>
<td>171.00 ± 34.17</td>
<td>0.36</td>
</tr>
<tr>
<td>Symptom at endoscopy, n (%)</td>
<td></td>
<td></td>
<td>0.24</td>
</tr>
<tr>
<td>No symptoms</td>
<td>13 (86.7%)</td>
<td>19 (76.0%)</td>
<td></td>
</tr>
<tr>
<td>Dyspepsia</td>
<td>1 (6.7%)</td>
<td>2 (8.0%)</td>
<td></td>
</tr>
<tr>
<td>Acid regurgitation</td>
<td>1 (6.7%)</td>
<td>0 (0.0%)</td>
<td></td>
</tr>
<tr>
<td>Epigastric pain</td>
<td>0 (0.0%)</td>
<td>4 (16.0%)</td>
<td></td>
</tr>
<tr>
<td>Endoscopic diagnosis, n (%)</td>
<td></td>
<td></td>
<td>0.3</td>
</tr>
<tr>
<td>Normal</td>
<td>3 (20.0%)</td>
<td>1 (4.0%)</td>
<td></td>
</tr>
<tr>
<td>Gastritis</td>
<td>11 (73.3%)</td>
<td>20 (80.0%)</td>
<td></td>
</tr>
<tr>
<td>Gastric ulcer</td>
<td>0 (0.0%)</td>
<td>4 (16.0%)</td>
<td></td>
</tr>
<tr>
<td>Duodenal ulcer</td>
<td>0 (0.0%)</td>
<td>0 (0.0%)</td>
<td></td>
</tr>
<tr>
<td>Others</td>
<td>1 (6.7%)</td>
<td>0 (0.0%)</td>
<td></td>
</tr>
</tbody>
</table>

Abbreviation: BMI, body mass index.

*P < 0.01.
carried out initiated at 95°C for 10 minutes, followed by 45 cycles at 95°C for 30 seconds, at 55°C for 30 seconds, at 72°C for 30 seconds, and final extension at 72°C for 5 minutes. The single-stranded DNA template was prepared from biotinylated PCR product by using streptavidin Sepharose HP beads (Amersham Biosciences) as described by the PSQ 96 sample preparation guide. Sequencing was conducted on a PyroMark ID system by using the Pyrogen Gold Reagents Kit (Biotage) according to the manufacturer’s instructions. The percentage of 15-PGDH methylation was determined by calculating the average methylation at 10 CpG sites in pyrosequencing.

Cell culture and H. pylori infection

The human gastric cancer cell line AGS was purchased in the year 2008 from the Korea Cell Line Bank (Seoul, Korea). The AGS cell line was authenticated by Korean Cell Line Bank. The authenticity of this cell line was validated by STR DNA fingerprinting using the AmplFLSTR identifier PCR Amplification Kit (Applied Biosystems). The STR profiles were analyzed using 3730 DNA analyzer (Applied Biosystems) and GeneMapper ID v3.2 (Applied Biosystems). The AGS cell line was last authenticated at the end of this study in the year 2013 by Korean Cell Line Bank. The Korean Cell Line Bank guaranteed the authenticity of cell lines. The AGS cell line was maintained in RPMI-1640 medium supplemented with 10% heat-inactivated FBS, 100 μg/mL penicillin, and 100 μg/mL streptomycin in a humidified atmosphere containing 5% CO2 at 37°C. The cells were seeded onto 60-mm culture dishes at 1 × 10^6 cells/dish in RPMI-1640 medium and cultured for 24 hours before being infected. For AGS cell infections, the H. pylori (ATCC43504, type strain 11637, caAg⁺ vacA⁺) was grown under microaerophilic conditions for 48 hours on Brucella broth agar plates containing 7% sheep blood, counted by using a UV scanning spectrophotometer (Shimazu) and added to the AGS cell monolayers for 4 hours at a multiplicity of infection (MOI) of 20. In another experiment, the cells were pretreated with inhibitors; PD98059 (ERK1/2 inhibitor), SB203580 (p38 inhibitor), SP600125 (JNK inhibitor), pretreated with inhibitors; PD98059 (ERK1/2 inhibitor), SB203580 (p38 inhibitor), SP600125 (JNK inhibitor), AG1478 (EGFR inhibitor), LY294002 [phosphoinositide 3-kinase (PI3K) inhibitor; Calbiochem] for 1 hour before the infection of with H. pylori for 4 hours. For lipopolysaccharide (LPS) treatments, the AGS cells were cultured for 24 hours and subsequently treated with LPS (1 or 10 μg/mL; Sigma) for 12 hours.

siRNA

siRNA for Snail 1 (UUGCAGUUGAAGGCCUUUUGGU), MiYD88 (GGCUAUCGAAAAAGGGUGGC; ref. 23), and negative control siRNA were purchased from Genolution and Invitrogen, respectively. The AGS cells were cultured in 60-mm cell culture dish for 24 hours at 37°C. Before H. pylori infection, the cells were transfected for 48 hours with 50 nmol/L siRNA by using Lipofectamine RNAiMAX (Invitrogen Inc.) as described by the manufacturer’s instructions. The medium was replaced, and the cells were infected with H. pylori.

Statistics

Statistical analysis was conducted by using GraphPad Prism version 5.01 for Window (GraphPad Software). Differences between 2 groups in terms of continuous variables were analyzed by using the Student t test and Mann–Whitney test, whereas differences in terms of categorical variables were evaluated by using the χ² test or Fisher exact test. P < 0.05 was considered to indicate statistically significant differences.

Results

Clinical characteristics of the study population

In total, 40 adult subjects [26 males (60%); median age, 56 years; range, 21–79 years] were recruited. Their baseline demographic and clinical characteristics are described in Table 1. Of the 40 subjects, 25 were positive for H. pylori infection (63%). The H. pylori–positive group was older than the negative group (P = 0.002) but did not differ significantly in terms of sex, smoking status, alcohol consumption, hypertension, diabetes, body mass index or cholesterol level. In addition, the 2 groups did not differ in terms of the symptoms at the time of endoscopy or the endoscopic diagnosis.

15-PGDH expression is downregulated in H. pylori–positive specimens

To investigate the regulation of 15-PGDH expression in the infected gastric specimens, the H. pylori–positive and -negative gastric specimens taken at baseline were first compared in terms of 15-PGDH mRNA expression. As shown in Fig. 1A, the positive specimens had significantly lower 15-PGDH mRNA expression than the negative specimens (P = 0.0004) but did not differ in terms of COX-2 mRNA expression (P = 0.6395; Fig. 1B). The 15-PGDH protein levels in the positive and negative specimens were then determined by Western blot analysis. As shown by Fig. 1C, the H. pylori–positive specimens expressed less 15-PGDH protein than the negative specimens. As shown in Fig. 1D, quantitation with densitometry and normalization relative to the GAPDH levels revealed that the relative mean intensity of 15-PGDH in the positive specimens (0.41; range, 0.00–0.98) was significantly lower than that in the negative specimens (0.75; range, 0.56–1.05; P = 0.0007). To confirm the suppression of 15-PGDH expression in H. pylori–positive tissues, immunohistochemistry with a 15-PGDH–specific antibody was conducted. The H. pylori–negative specimens exhibited strong 15-PGDH expression in epithelial cells located at the upper luminal surface of the gastric proliferative zone. In contrast, the H. pylori–positive specimens showed a significant loss of 15-PGDH expression along with higher numbers of mononuclear inflammatory cells in the lamina propria (Fig. 1E).

15-PGDH downregulation and Snail upregulation in H. pylori–infected AGS cells and their role in regulation of TLR4/ERK pathway

To investigate the mechanism underlying the H. pylori–associated suppression of 15-PGDH expression, AGS cells
which has a high endogenous level of 15-PGDH) were infected with H. pylori (Supplementary Fig. S1A). Western blot analysis revealed that the H. pylori infection significantly decreased 15-PGDH protein levels (Fig. 2A, Supplementary Fig. S1B) and increased the transcription factor, Snail, after 4 hours (Fig. 2A). To determine whether inhibition of 15-PGDH was specific for the H. pylori, we examined whether 15-PGDH expression was affected by LPS treatment. 15-PGDH expressions did not change after LPS treatment in AGS cells (Supplementary Fig. S2). As previous reports have shown that the transcription factor Snail represses the transcription of 15-PGDH (24, 25). We confirmed that inhibition of Snail using siRNA against Snail 1 blocked the H. pylori–mediated downregulation of 15-PGDH expression (Fig. 2B). The present results suggest that H. pylori suppress 15-PGDH protein levels by activating Snail.

Specific chemical inhibitors, namely the ERK1/2 inhibitor PD98059, the EGFR inhibitor AG1478, and the PI3K inhibitor LY294002, were used to elucidate the cell signaling pathway that was involved in the downregulation of 15-PGDH in H. pylori–infected AGS cells. Pretreatment with PD98059, AG1478, and LY294002 partially reversed the stimulatory effect of H. pylori on Snail (Fig. 2C). However, blocking with PD98059 and AG1478, an ERK1/2 and EGFR inhibitor, restored the downregulation of 15-PGDH mediated by H. pylori infection (Fig. 2B). To determine whether ERK1/2 is activated by H. pylori infection, we conducted a Western blot analysis for active, phosphorylated ERK1/2 (p-ERK1/2) and found that ERK1/2 was activated by H. pylori within 1 hour after infection commenced, with maximal activity occurring by 2 hours (Fig. 2D). We also showed that another MAPK pathway signaling p38 and INK were activated by H. pylori (Supplementary Fig. S3A); however, only blocking with PD98059 restored 15-PGDH expressions and suppressed upregulated Snail expressions by H. pylori (Supplementary Fig. S3B). These results indicated that activation of the

![Figure 1](https://example.com/figure1.png)

Figure 1. Expression of 15-PGDH and COX-2 in gastric tissues from H. pylori–positive and -negative subjects. A, expression of 15-PGDH mRNA. The positive specimens expressed significantly less 15-PGDH than the negative specimens. B, expression of COX-2 mRNA. The specimens did not differ in terms of COX expression. HP(+), H. pylori–positive subjects (n = 23); HP(−), H. pylori–negative subjects (n = 15). The values are the means ± SD of 3 independent experiments. ***, statistically significant differences (P < 0.001). C, representative immunoblots showing 15-PGDH expression in infected and noninfected specimens. D, densitometry of 15-PGDH protein expression normalized relative to GAPDH levels. The H. pylori–positive specimens had a significantly lower relative mean 15-PGDH expression intensity (0.44; range, 0.00–0.98) than the H. pylori–negative specimens (0.75; range, 0.56–1.05). HP(−), H. pylori–positive subjects (n = 14); HP(+), H. pylori–positive subjects (n = 14). The values are the means ± SD of 3 independent experiments. ***, statistically significant differences (P < 0.001). E, immunohistochemical staining of 15-PGDH. Compared with the noninfected specimens, the infected specimens had significantly lower 15-PGDH expression in epithelial cells located at the upper luminal surface and higher numbers of mononuclear inflammatory cells in the lamina propria. Original magnification, ×200. Scale bars, 500 mm.
ERK1/2 pathway by *H. pylori* is specific for Snail upregulation and subsequent 15-PGDH suppression by *H. pylori* infection.

TLR4 regulates the response of human gastric epithelial cells to *H. pylori* (26). To test whether it participates in the *H. pylori*–mediated suppression of 15-PGDH, infected AGS cells were subjected to Western blot analysis of TLR4 expression. The expression of MyD88 was also examined because it is an adaptor molecule that participates in TLR signaling (20). *H. pylori* infection significantly induced TLR4 protein expression after 5 minutes of exposure, whereas the expression of MyD88 increased between 5 minutes and 1 hour of exposure (Fig. 2E). In addition, TLR2 expression was increased by *H. pylori* infection (Supplementary Fig. S4).

To investigate the functional role of MyD88 in *H. pylori*–associated 15-PGDH regulation, an siRNA against MyD88 was used to knockdown MyD88 expression. Interestingly, the inhibition of MyD88 blocked the *H. pylori*–mediated downregulation of 15-PGDH expression (Fig. 2F). It also significantly suppressed the *H. pylori*–induced phosphorylation of ERK1/2 (Fig. 2F), which suggests that the 15-PGDH suppression was mediated by TLR4/MyD88 and activated ERK1/2.
Promoter methylation of 15-PGDH DNA in *H. pylori*-positive gastric tissues

To investigate the significance of 15-PGDH promoter methylation in *H. pylori*-infected gastric specimens, the 15-PGDH promoter methylation status of 13 *H. pylori*-negative and 20 positive specimens was examined. In 14 of the 20 *H. pylori*-positive subjects (70%), the 15-PGDH promoter showed increased methylation (Fig. 3A). In contrast, only 5 of the 13 *H. pylori*-negative subjects (38%) showed this (Fig. 3A). To assess the precision with which bisulfate PCR determined the 15-PGDH promoter methylation in gastric specimens, the 20 positive and 13 negative specimens were also subjected to pyrosequencing analysis. The percentage of methylation for 10 CpG sites analyzed ranged from 2.6% to 6.4% methylated in *H. pylori*-positive, whereas only from 1.7% to 4.3% methylated in *H. pylori*-negative subjects. The levels of 15-PGDH promoter methylation were significantly higher in *H. pylori*-positive specimens than in negative (*P* = 0.0111; Fig. 3B). When all samples were pooled and divided into 2 groups that had >4% and ≤4% promoter methylation, respectively, the group with >4% promoter methylation had significantly lower 15-PGDH mRNA levels than the ≤4% group (*P* = 0.0097; Fig. 3C), suggesting that the subtle increase in 15-PGDH methylation seen in *H. pylori*-infected individuals in vivo may play a role in modulating 15-PGDH mRNA levels.

Restoration of 15-PGDH expression after *H. pylori* eradication

To explore whether *H. pylori*-associated suppression of 15-PGDH expression can change if *H. pylori* is eradicated, gastric samples were collected from 4 of the *H. pylori*-positive subjects 8 weeks after triple therapy regimen and the 15-PGDH mRNA expression was evaluated. In all 4 cases, 15-PGDH mRNA expression increased markedly after treatment (Fig. 4A), whereas the expression of COX-2 mRNA decreased in 3 of the 4 cases (Fig. 4B). A follow-up CLO test and histologic examination showed that all 4 patients had become *H. pylori*-negative after treatment.

Discussion

Infection-associated inflammation plays an important role in promoting cancer development (27). Examples of cancers that are linked to infection include bladder cancer (shistosomiasis), hepatocellular carcinoma (hepatitis B and...
reported that EGFR signaling induces Snail, which binds was specific for the transcriptional regulation and this inhibition of 15-PGDH mediated suppression of 15-PGDH was mediated by TLR4-ed. Moreover, an
15-PGDH expression recovered after the infection-
mediated suppression of 15-PGDH was mediated by TLR4-MyD88 activation and involved ERK1/2 or EGFR-Snail transcriptional regulation and this inhibition of 15-PGDH was specific for the H. pylori infection. Previously, it was reported that EGFR signaling induces Snail, which binds conserved E-box elements in the PGDH promoter to repress transcription (25). We elucidated that H. pylori-associated signaling pathway involving TLR4-MyD88 and ERK1/2 is also related to the 15-PGDH suppression that might be related with gastric carcinogenesis. In addition, a modest elevated CpG island methylation of 15-PGDH DNA was found associated with the downregulation of 15-PGDH by H. pylori.

Recent studies including our own, have suggested that 15-PGDH behaves as a tumor suppressor in several tumor types including colon, lung, breast, bladder, thyroid, and pancreas carcinomas (12–17). Previously, we reported that 70.1% of gastric cancer specimens lack 15-PGDH protein expression (19). In addition, exploration of 15-PGDH expression in preneoplastic lesions showed that while 15-PGDH repression was not present in metaplasia, it was markedly reduced in 83.3% of adenomas (19). The present study showed that 15-PGDH was even repressed in H. pylori-positive specimens that only showed mild gastritis. This suggests that H. pylori infection initiates gastric carcinogenesis by elevating PGE2 levels even before the formation of preneoplastic lesion. The elevated expression of COX-2 is also found in more than 70% of gastric cancers, and this is reversed when H. pylori is eradicated (29). In addition, transgenic mice expressing COX-2 and the PGE2-converting enzyme microsomal prostaglandin E synthase-1 (mPGES-1) develop hyperplastic gastric tumors, which suggests that the activation of prostaglandin synthesis plays a major role in H. pylori-associated gastric carcinogenesis (30). The present study also showed that elevated PGE2 levels in gastric carcinogenesis are a consequence of not only increased synthesis but also reduced catabolism.

Murine model studies have indicated that PGE2 suppresses the T-helper 1 immune response in the H. pylori-infected stomach and that inhibition of COX-2 enzymatic activity accelerates the development of gastritis and premalignant gastric lesions; moreover, systemic administration of synthetic PGE2 prevents premalignant changes (7). These observations suggest that PGE2 may suppress infection-associated carcinogenesis, which appears to be inconsistent with the present results. PGE2 also promotes crypt stem cell survival and proliferation (31) and plays a role in the wound-healing process by enhancing the production of VEGF, a key factor in angiogenesis (32). 15-PGDH is also downregulated in inflammatory bowel diseases (33). Thus, in the short term, it seems likely that the reduced degradation of PGE2 in infected gastric tissue contributes to the healing of the inflamed mucosa. However, if the elevation of PGE2 in inflamed mucosa is chronic, it may stimulate mitogenesis and inhibit apoptosis, thereby lowering the threshold for the development of cancer. Recent reports suggest that bacterial infection, in cooperation with PGE2 signaling through the EP4 receptor, induces macrophage recruitment and promotes gastric tumorigenesis by TNF-a and Wnt signaling (6). This supports the notion that chronic activation of PGE2 has procarcinogenic effects. The short-term effect of PGE2 activation on gastric carcinogenesis should be investigated in future studies.

Cancer Prev Res; 6(4) April 2013

Figure 4. 15-PGDH and COX-2 mRNA levels in gastric specimens from 4 subjects before and after H. pylori was eradicated, as measured by quantitative real-time PCR. A, the gastric 15-PGDH mRNA expression increased significantly in all 4 subjects after H. pylori eradication. AT, after treatment; BT, before treatment. B, the gastric COX-2 mRNA expression dropped in 3 of the 4 subjects after eradication.
The 15-PGDH gene has 2 regions that contain clusters of putative transcription factor–binding sites. The transcription factors that have been reported to regulate these regions are Ets, activating protein-1 (AP-1), Snail, Alug, and ZEB1 (10). Previous studies indicate that EGFR activation suppressed 15-PGDH through Snail (25). In addition, blockade of EGFR signaling induces 15-PGDH while reducing Snail in vitro and PGE2-dependent xenograft model (25). EGFR signaling induces Snail, which binds to conserved E-box elements in the 15-PGDH promoter to repress transcription (25). In our study, analysis of the mechanism by which H. pylori regulated 15-PGDH expression in a gastric cancer cell line revealed that infection with H. pylori suppressed 15-PGDH expression and upregulated Snail expression. H. pylori infection also activated the phosphorylation of ERK1/2, and ERK1/2 inhibition clearly prevented the suppression of 15-PGDH and the upregulation of Snail levels by H. pylori. Also, the blockade of EGFR signaling restored H. pylori–mediated down-regulation of 15-PGDH. This suggests that ERK1/2 and EGFR are necessary for the H. pylori–mediated repression of 15-PGDH. This result is consistent with our previous report, which suggests that 15-PGDH is repressed in gastric cancer by the activation of the EGFR/ERK1/2 pathway (19).

The present study also assessed whether the TLR/MyD88 pathway, which is a well-known bacterial recognition system, plays a role in the H. pylori–associated repression of 15-PGDH. It was found that TLR4 and MyD88 were activated by H. pylori, as has been reported previously (20). To elucidate whether MyD88 is needed for H. pylori–induced 15-PGDH repression, an siRNA against MyD88 was used. Interestingly, this siRNA lifted the repression of 15-PGDH and blocked the phosphorylation of ERK1/2. These observations together suggest that H. pylori activates TLR4/MyD88, thereby elevating ERK1/2 phosphorylation, which in turn represses 15-PGDH expression (Fig. 5).

TLR4 signaling via MyD88 activates a signaling cascade that results in the enhanced transcription of COX-2 and the increased production of PGE2 (22, 23). PGE2 activates Wnt and peroxisome proliferator–activated receptor-δ signaling to induce target genes involved in replication and survival; it also triggers cell migration by activating the EGFR pathway (5, 8). The present study identified another pathway that could promote gastric carcinogenesis, namely one where the catalysis of PGE2 by 15-PGDH is suppressed via TLR4/MyD88 signaling. Notably, our analysis of human samples revealed that H. pylori infection clearly downregulated 15-PGDH but did not significantly activate COX-2. It is unclear why COX-2 activation was not evident in the samples, but it is possible that at this stage, of H. pylori infection, PGE2 levels are controlled by the degradation of PGE2 rather than the synthesis of PGE2. It is an intriguing observation that one case had increased COX-2 after H. pylori eradication. It is difficult to explain, however, it might be because the gastric sample obtained from this patient contains large portion of remained lymphocytes resulting in increase of COX-2 expression.

The main strategies of gastric cancer chemoprevention include H. pylori eradication and the use of nonsteroidal anti-inflammatory drugs that mainly target the inhibition of prostaglandin synthesis (34). Previous trials on drugs that inhibit COX-2 activity and that could decrease the risk of colon carcinogenesis showed that these drugs are problematic because they induce unfavorable cardiovascular side effects (35). Our previous data indicated that these side effects can be reduced by using effective biomarkers such as 15-PGDH to select the subjects who will benefit from these chemopreventive drugs (36). In addition, we hypothesize that gastric neoplasia could be prevented by combining H. pylori eradication with compounds that can act in the gastrointestinal tract to re-induce 15-PGDH expression in early neoplastic cells.

We could not recruit enough patients because it was very difficult to get the permission for the sampling in patients before and after H. pylori eradication therapy. However, we believe that our preliminary result provide valuable suggestion that 15-PGDH can be restored after H. pylori eradication. Further study including long-term follow-up measurement will be necessary to prove this hypothesis.

In conclusion, the present study showed that 15-PGDH, a novel tumor suppressor, was significantly downregulated in H. pylori–infected gastric human tissues, which suggests that H. pylori infection may repress the catabolism of prostaglandins, thereby elevating prostaglandin levels, which could promote gastric carcinogenesis. An in vitro study
revealed that this process is mediated by TLR4-MyD88 activation and ERK1/2 or EGFR-Snail transcriptional activation. In addition, H. pylori appeared to suppress 15-PGDH expression by inducing aberrant 15-PGDH promoter methylation. Finally, sequential analysis of 15-PGDH transcription activity after H. pylori eradication revealed that the infection-induced suppression of 15-PGDH expression was reversible. The elucidation of this novel mechanism provides new insights that may improve our understanding of H. pylori–induced gastric carcinogenesis. Further studies into 15-PGDH inducers that could serve as chemopreventive agents of gastric carcinogenesis are warranted, as are studies that determine the usefulness of this novel enzyme as a biomarker of gastric carcinogenesis.

**Disclosure of Potential Conflicts of Interest**

No potential conflicts of interest were disclosed.

**Authors’ Contributions**


Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): Y.-M. Ryu, S.-J. Myung, H.-J. Lee, S.-K. Park, S.P. Fink, K.W. Jung, J.-H. Kim

**References**


Inhibition of 15-Hydroxyprostaglandin Dehydrogenase by Helicobacter pylori in Human Gastric Carcinogenesis

Yeon-Mi Ryu, Seung-Jae Myung, Young Soo Park, et al.


Updated version
Access the most recent version of this article at:
doi:10.1158/1940-6207.CAPR-12-0389

Supplementary Material
Access the most recent supplemental material at:
http://cancerpreventionresearch.aacrjournals.org/content/suppl/2013/02/19/1940-6207.CAPR-12-0389.DC1

Cited articles
This article cites 36 articles, 13 of which you can access for free at:
http://cancerpreventionresearch.aacrjournals.org/content/6/4/349.full.html#ref-list-1

Citing articles
This article has been cited by 3 HighWire-hosted articles. Access the articles at:
/content/6/4/349.full.html#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.