## New Perspectives of Curcumin in Cancer Prevention

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### Abstract
Numerous natural compounds have been extensively investigated for their potential for cancer prevention over the decades. Curcumin, from Curcuma longa, is a highly promising natural compound that can be potentially used for chemoprevention of multiple cancers. Curcumin modulates multiple molecular pathways involved in the lengthy carcinogenesis process to exert its chemopreventive effects through several mechanisms: promoting apoptosis, inhibiting survival signals, scavenging reactive oxidative species (ROS), and reducing the inflammatory cancer microenvironment. Curcumin fulfills the characteristics for an ideal chemopreventive agent with its low toxicity, affordability, and easy accessibility. Nonetheless, the clinical application of curcumin is currently compromised by its poor bioavailability. Here, we review the potential of curcumin in cancer prevention, its molecular targets, and mechanisms of action. Finally, we suggest specific recommendations to improve its efficacy and bioavailability for clinical applications.

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### Introduction
Cancer is a major health problem that can debilitate and destroy human lives. One out of every 4 deaths in the United States is caused by cancer. More than $124.6 billion was spent in direct medical costs for 13.7 million cancer survivors and 1.5 million newly diagnosed cancer patients in the United States in 2010. Increasing human life expectancy will inevitably raise cancer prevalence and the related costs. Consequently, the development of effective cancer prevention strategies is increasingly important. Histologically, the development of cancer involves multiple steps, which occur over several years after the initial carcinogen exposure from normal to hyperplasia, mild, moderate, and severe dysplasia, and carcinoma in situ, before finally progressing to invasive cancer (1). Throughout this long, multi-step developmental course, there is a wide scope of possible preventive approaches that can delay or prevent the development of cancer. Different cancer prevention strategies such as behavioral modification, vaccines, surgical manipulation, and chemoprevention have evolved with tremendous research efforts (2). Many investigations have proven that healthy lifestyles involving balanced diets, regular exercise, smoking cessation, alcohol reduction, weight control, and stress management are beneficial for decreasing cancer risk and can never be overemphasized (3–7). One particular milestone in cancer prevention was the approval by the U.S. Food and Drug Administration (FDA) of the human papilloma virus (HPV) cervical cancer vaccine in 2009 as a result of positive randomized controlled clinical trials.

The term chemoprevention was first coined in 1976 by M. B. Sporn, who defined it as a preventive modality in which natural or synthetic agents can be employed to slow, stop, reverse, or prevent the development of cancer. Since then, researchers have investigated numerous agents for this purpose with few successes. The first important translational study of a potentially chemopreventive agent was conducted with 13-cis retinoic acid (13-cRA), which resulted in successful size reduction of the premalignant lesion oral leukoplasia, albeit with some notable toxicities (8). In an attempt to reduce the toxicity, this study was followed by another trial using high-dose isotretinoin induction and maintenance with isotretinoin or beta-carotene, which suggested that isotretinoin is significantly more effective than beta-carotene against leukoplasia (9). Another follow-up study using low-dose isotretinoin and a large cohort of patients resulted in a negative outcome (10). In contrast, the field of breast cancer chemoprevention research gained considerable momentum after positive large-scale clinical trials of tamoxifen, a selective estrogen receptor modulator (SERM), led to its FDA approval (11). However, not all cancer types have successful chemoprevention stories. In colorectal cancer, despite positive secondary clinical trials of sulindac, celecoxib, and aspirin, primary prevention using cyclooxygenase-2 (COX-2) inhibitors was shown to have no benefit in the general population and the study was terminated early because of cardiovascular toxicity (12–14). Another disappointment was the recently conducted selenium and vitamin E cancer prevention trial (SELECT), which gave negative results in patients with lung and prostate cancers (15). After several large negative clinical trials were reported, the focus of the new era in chemoprevention has shifted toward molecularly targeted agents and less toxic natural compounds.
In chemoprevention, the safety of the participants is the first priority and should be considered of the utmost importance because essentially healthy people will receive the chemopreventive treatment for a long period of time. Moreover, the toxicity of the agents could impact patient accrual in larger scale studies in real clinical practice. To this end, unlike synthetic compounds, the safety of natural compounds present in fruits, vegetables, and spices are well established through their long-term consumption in human history (16). Therefore, taking natural compounds for cancer prevention can be a well-justified and effective strategy for people with increased risk for cancer development – such as those with premalignant lesions of intraepithelial neoplasia. Among many such natural compounds, curcumin has drawn special attention for its chemoprevention potential because of its safety, multi-targeted anticancer effects, and easy accessibility (16). The following sections will discuss different aspects of curcumin as a chemopreventive agent, including its safety, efficacy, and mechanism of action.

Curcumin in Chemoprevention

Since 1987, the National Cancer Institute (NCI) has tested more than 1,000 different potential agents for chemopreventive activity, of which only about 40 promising agents were moved to clinical trials (17). Curcumin, present in the Indian spice “haldi”, is 1 such agent that is currently under clinical investigation for cancer chemoprevention. Three polyphenols (Fig. 1) were isolated from Curcuma longa, of which curcumin (bis-α,β-unsaturated β-diketone) is the most abundant, potent and extensively investigated (16). Curcumin has been used empirically as a remedy for many illnesses in different cultures. It is only in the last few decades that curcumin’s effects against cancer and cancer therapy-related complications have emerged, through much investigation. The first clinical report of the anticancer properties of curcumin was from Kuttan and coworkers, who used a 1% curcumin ointment on skin cancerous lesions with a reduction in smell in 90% of patients (18); 10% patients experienced a reduction in pain and lesion size. In an experimental model of mammary cancer induced by 7,12-dimethylbenz-[a]-anthracene (DMBA) in female rats, the initiation of DMBA-induced mammary adenocarcinoma was significantly decreased by intraperitoneal infusion of curcumin 4 days before DMBA administration (19). In a study of esophageal cancer prevention in curcumin-fed F344 rats, the chemopreventive activity of curcumin was observed not only in the initiation phase but also in post-initiation phases (20). In addition, in a familial adenomatous polyposis (FAP)-simulated study in which the APC gene of C57Bl/6 Min/+ mice was mutated to result in the development of numerous adenomas by 15 weeks of age, an oral curcumin diet prevented adenoma development in the intestinal tract, suggesting the chemopreventive effect of curcumin in colorectal cancer with APC mutation (21). Moreover, in a rat model of N-nitrosodiethylamine and phenobarbital-induced hepatic cancer, curcumin reduced lipid peroxidation and salvaged hepatic glutathione antioxidant defense, which eventually may have contributed to hepatic cancer prevention (22). Several studies of cancer prevention at different stages have shown the multi-targeted anticancer and chemopreventive effects of curcumin and have suggested it as a very favorable agent for chemoprevention.

Mechanisms of Anticancer Effects

According to their mode of action, chemopreventive agents are classified into different subgroups: antiproliferatives, antioxidants, or carcinogen-blocking agents. Curcumin belongs to all 3 subgroups, given its multiple mechanisms of action. The anticancer effects of curcumin mainly result from multiple biochemical mechanisms that are involved in the regulation of programmed cell death and survival signals. The curcumin targets that are involved in signaling pathways include transcription factors, growth factors, inflammatory cytokines, receptors, and enzymes (Fig. 2). In different types of cancers, curcumin exhibits anticancer actions through a combination of different mechanisms, including survival signal reduction, proapoptotic promotion, anti-inflammatory actions, and reactive oxygen stress (ROS) scavenging to different degrees. The effects of curcumin on these signaling pathways are expected to be more complicated in the real setting, and the mechanisms of curcumin’s chemopreventive, chemosensitizing, and radiosensitizing effects are more vigorously being studied now.

Survival signals – nuclear factor-κB

The survival signals in cancer cells are upregulated to support proliferation and survival against anticancer treatment. The central role players in this process are nuclear
factor-kB (NF-kB), Akt, and their downstream cascades that can lead to the upregulation of antiapoptotic Bcl-2 proteins. Curcumin can modulate these signals by inhibiting the NF-kB pathways at multiple levels (23, 24). Curcumin significantly inhibited the growth of squamous cell carcinoma of head and neck (SCCHN) xenograft tumors in nude mice. Inhibition of nuclear and cytoplasmic IκBβ kinase (IKKβ) in the xenograft tumors decreased NF-kB activity (25). In addition, curcumin was shown to enhance chemosensitivity in 5-fluorouracil and cisplatin-treated esophageal adenocarcinoma as well as in paclitaxel-treated breast cancer cells by inhibiting compensatorily upregulated NF-kB (26). Likewise in a colon cancer cell line during radiotherapy, curcumin blocked NF-kB and reduced radioresistance (27).

Apoptotic signals— intrinsic and extrinsic

Curcumin induces programmed cell death (apoptosis) in many cancer cell types. Both intrinsic and extrinsic apoptotic pathways are activated by curcumin. In the intrinsic pathway, various cell stresses – irreversible DNA damage, defective cell cycle, or loss of growth factors – can generate death signals and ultimately pass them down to mitochondria. Then, depending on the balance of Bcl-2 family members, the destiny of the cell is driven into apoptosis. Curcumin upregulates the p53 modulator of apoptosis (PUMA) and Noxa, which, in turn, activates the proapoptotic multidomain Bcl-2 family members Bax, Bim, and Bak and downregulates Bcl-2 and Bcl-xl. Loss of balance between pro- and antiapoptotic Bcl-2 proteins causes calcium influx into mitochondria and decrease in mitochondrial outer membrane permeability (MOMP) that allows cytochrome C and Smac release into the cytoplasm, eventually leading to the activation of a cascade of caspases and formation of the apoptosome, causing apoptosis (28).

In the extrinsic pathway, death signals are initiated from the exterior environment of the cells via Fas, tumor necrosis factor (TNF), and death receptors (DR) 3–6. When the signal is received, conformational change in the receptors allows Fas-associated death domain (FADD) binding and recruits the death-induced signaling complex (DISC),

Figure 2. Molecular targets of curcumin. C, Curcumin; CIAP, cleavage inhibitor of apoptosis; FADD, Fas-associated protein with death domain; FLIP, FLICE-like inhibitory protein; DISC, death-inducing signaling complex; MOMP, mitochondrial outer membrane permeabilization; PKC, protein kinase C; PLC, phospholipase C; XIAP, X-linked inhibitor of apoptosis protein; VEGF, vascular endothelial growth factor; FGF, fibroblast growth factor; PDGF, platelet-derived growth factor; EGF, epidermal growth factor.
which activates the formation of initiator caspases 8 and 10. Curcumin was shown to upregulate extrinsic apoptosis pathway signals via the Fas pathway. In TNF-related-apoptosis-inducing ligand (TRAIL)-resistant cell lines, curcumin also enhanced apoptosis by upregulating the expression of DR 4 and 5 (29). After DISC recruitment, activation of the initiator caspases is regulated by Flice-like inhibitory protein (FLIP) and curcumin was shown to downregulate c-FLIP in natural killer/T-cell lymphoma (30). Afterwards, the initiator caspase cleaves Bid and the truncated Bid (tBid) provides cross-talk between the intrinsic and extrinsic pathways by delivering death signals from initiator caspases directly to the mitochondrial pathway. In SKOV3 and OVCA429 ovarian carcinoma cells, curcumin showed induction of both intrinsic and extrinsic apoptosis by cleavage of pro-caspase 3, 8, 9, and cytochrome C release followed by tBid formation (31).

p53 plays a major role in tumor development and treatment; however, more than 50% of all cancers have p53 mutations. p53 proofreads DNA and recognizes uncorrectable mutations, at which point it arrests the cell cycle and steers the cell toward programmed cell death. Curcumin was shown to upregulate p53 expression followed by an increase in p21 (WAF-1/CDK-1), resulting in cell-cycle arrest at G0/G1 and/or G2/M phases. This is eventually followed by the upregulation of Bax expression, which induces apoptosis (32). On the other hand, curcumin has also shown its p53-independent anticancer effect as an inhibitor of the proteasome pathway by inhibiting ubiquitin isopeptidase (33). In a prostate cancer cell line, curcumin downregulated MDM2, the ubiquitous ligase of p53, and displayed enhanced anticancer effect via PI3K/mTOR/ETS2 pathways in PC3 xenografts in nude mice receiving gemcitabine and radiation therapy (34). In p53 mutant or knockout ovarian cancer cell lines, curcumin induced p53-independent apoptosis which involved p38 mitogen-activated protein kinase (MAPK) activation and inhibited Akt, resulting in decreased expression of Bcl-2 and survivin (31). Taken together, cancers with both deleted/mutant and wild-type p53 can benefit from curcumin treatment to achieve an anticancer effect.

Trophic signals—growth factors and cytokines

Different kinds of trophic factors including growth factors and cytokines can contribute to growth signals in cancer cells. Curcumin inhibits epidermal growth factor receptor (EGFR) kinase phosphorylation and strongly degrades Her2/neu protein, which ultimately inhibits cancer growth (35). In SCCHN, curcumin targets both EGFR and VEGF to inhibit cell growth (36). Therefore, the multi-targeted activity of curcumin may be potentially more effective. In an estrogen receptor-negative breast cancer cell line, curcumin inhibited angiogenesis factors such as VEGF and basic fibroblast growth factor (b-FGF) at the transcriptional level (37). Curcumin was also shown to inhibit expression of proinflammatory cytokines such as interleukin-1β (IL-1β) and IL-6 and exhibited growth inhibitory effects through inhibition of the NF-κB and MAPK pathways (38). In a breast cancer cell line, curcumin was shown to inhibit phosphorylation of Akt within the MAPK/PI3K pathway, which led to proapoptosis (39).

Roles of reactive oxidative stress

ROS has opposing effects on cancers: it can be an insult causing DNA mutations in carcinogenesis, and it can also drive mitochondrial apoptosis. Minimizing DNA insult by scavenging ROS is important for the prevention of cancer, while generating ROS to drive mitochondrial apoptosis is more important when treating malignancies. In terms of ROS scavenging, curcumin was shown to induce phase II metabolizing enzymes in male mice-glutathione-S-transferase (GST) and quinine reductase, which can neutralize ROS derived from chemical carcinogens (40). In addition, curcumin was shown to induce another important ROS scavenging enzyme – hemeoxygenase-1, the redox-sensitive inducible enzyme, via nuclear factor 2-related factor (Nrf-2) regulation (41). Curcumin is a ROS scavenging enzyme inducer but, on the other hand, it also uses ROS to kill cancer cells. ROS generated by curcumin in human renal Caki cell downregulated Bcl-xl and inhibitors of apoptosis proteins (IAP), thereby inducing apoptosis (42). In cervical cancer cell lines, curcumin-generated ROS activated extracellular signal-regulated kinase (ERK), which modified radiosensitivity (43). Despite the paradoxical roles of curcumin in scavenging and generating ROS, the overall effect of curcumin is an anticancer activity.

Microenvironments—inflammation

Regarding the cancer microenvironment, the anticancer effect of curcumin is also described as being antagonistic to leaky vessels and loss of adhesion, which are closely related to cancer development and invasiveness. The relationship between the proinflammatory enzymes COX-2 and lipoxynase (LOX) and the possible development of colorectal, lung, and breast cancers has been investigated (44). In colorectal cancer, development of the premalignant lesion aberrant crypt foci (ACF) was shown to be related to upregulated COX-2 level via inducible nitric oxide synthase (iNOS). As a nonspecific iNOS inhibitor, curcumin significantly inhibited colonic ACF formation in F344 rats (45). Moreover, curcumin downregulates CXCL-1 and -2 via NF-κB inhibition and, accordingly, downregulates the metasis-promoting gene CXCR4 in a breast cancer cell line (46). In the normal Wnt pathway, β-catenin participates in the regulation of cell-to-cell adhesion integrity; however, in certain cancers, aberrant β-catenin accumulation promotes survival through an upregulated Akt pathway. In colon cancer cells, curcumin promoted caspase-3-mediated cleavage of β-catenin and decreased the level of the oncogene c-Myc (47). In addition, the invasiveness/metastasis of cancers was shown to be related to matrix metalloproteinase-9 (MMP-9) secretion, and treatment of an invasive hepatocellular cancer cell line with curcumin resulted in diminished invasiveness due to inhibition of MMP-9 secretion by curcumin (48).
Cancer stem cells and miRNA

Cancer stem cells (CSC) are a rare population of cells within the tumor having cell-renewal properties and are thought to be responsible for tumor initiation and treatment failures. The cancer stem-cell concept has important implications for cancer therapy and targeting CSCs is a relatively new strategy that can decrease cancer recurrence and relapse and treatment failure. Several studies have suggested that curcumin and its analogs can also target CSCs. In prostate cancer cells under hypoxic conditions, the curcumin analog CDF decreased CSC markers such as Nanog, Oct4, and EZH2 as well as miR-21, which contributed to deregulation of CSC function through the effects of CDF on the hypoxic pathway via HIF-1α (49). In colon cancer cells, STAT3 overexpression was found in ALDH (+)/CD133+ CSCs. The curcumin analog GO-Y030 inhibited the expression of STAT3 expression and suppressed CSC growth in colon cancer cells (50). Furthermore, in the rat glioma cell line C6, curcumin was shown to decrease the side population which is known to be associated with stem cell populations (51).

In addition, miRNAs play essential roles in tumorigenesis and anticancer drug development because of their ability to target both tumor suppressor and oncogenes. Curcumin and its analogs also target miRN, which contributes to their chemopreventive potential. By controlling epigenetic gene expression via EZH2-miR regulation, CDF increased the levels of tumor-suppressive miRN that are mostly absent in pancreatic cancer cells including let-7a, b, c, d, miR-26a, miR-101, miR-146a, and miR-200b, c and resulted in decreased pancreatic cancer cell survival and aggressiveness (49). Curcumin and its cyclohexanone and piperidine analogs inhibited growth of multiple colon cancer cells by targeting Sp transcription factors (52). Induction of the Sp repressors ZBTB10 and ZBTB4 and downregulation of miR-27a, miR-20a, and miR-17–5p by these compounds are important for inhibiting Sp transcription factors. In addition, miR-203, which regulates the Src–Akt axis, is a target of curcumin in bladder cancers (53).

Curcumin and host factors: Immunomodulation and metabolism

Because of poor bioavailability, it is practically impossible to reach the in vitro effective dose of curcumin in vivo. Nonetheless, curcumin is effective in vitro in inhibiting tumor growth and modulating biomarkers, suggesting that the host factors such as the host immune system and metabolic systems have an effect on its activities. Lack of functional T-cells or T-cell derived cytokines such as interferon-γ promotes spontaneous as well as carcinogen-induced tumorigenesis. CD8(+) cytotoxic T lymphocytes (CTL) are involved in antigen-specific tumor destruction and CD4(+) T cells are essential for helping this CD8(+) T-cell-dependent tumor eradication. Curcumin prevented loss of T cells, expanded T-cell populations, and reversed the type 2 immune bias and attenuated the tumor-induced inhibition of T-cell proliferation in tumor-bearing hosts (54). Moreover, curcumin inhibited the production of immunosuppressive cytokines such as TGF-B and IL-10 in these hosts. Another study suggested that increased CD8+ T cells enhance the production of INF-γ by curcumin (55). Another host effect is on the metabolism of curcumin, which involves 2 routes: 1 route transforms curcumin to hexahydrocurcumin through successive reductions (probably through its intermediates dihydrocurcumin and tetrahydro-curcumin), while the other route involves rapid molecular modification by conjugation to glucuronide, sulfate, and glucuronide–sulfate forms (56). Although the main curcumin metabolites remain controversial, both in vivo and in vitro cell-free studies suggest that hydrocurcumin are more potent antioxidants than the parent curcumin in scavenging free radicals, reducing lipid peroxidation, and in enzyme activation (of superoxide dismutase, catalase, GSH peroxidase, and GST; refs. 57, 58). These antioxidant effects were shown to be critical for the chemopreventive potential of curcumin. Thus, curcumin displayed efficacy in vivo probably due to the presence of these host effects.

Clinical Trials of Curcumin Use in Cancer

Many positive preclinical cell line and animal model studies have brought curcumin to clinical trials to test its safety and efficacy as a chemopreventive agent. Several clinical trials have already been completed, the results of which are summarized in Table 1 (59–68). In phase I trials, curcumin was tested for its toxicity and tolerability, and was found to be highly tolerable at doses up to 12 g/d with no curcumin-related toxicities (60, 61). However, due to its poor bioavailability, curcumin was not detectable in blood when administered at doses up to 8 g, and was detected at very low levels following 10- and 12-g doses with a peak concentration at 1 to 2 hours. Histologic improvements of the lesions were observed in most of the treated patients (60). Radiologically stable disease was also shown in 5 out of 15 colorectal cancer patients who were refractory to chemotherapy in a second study (61). In a colorectal cancer trial, curcumin was shown to modulate biomarkers such as GST activity, deoxyguanosine adduct M(1)G, and PGE2 (prostaglandin E2; refs. 61–63). A decrease in lymphocytic GST activity of 59% resulted after administration of 440 mg of Curcuma extract (61). The levels of M(1) decreased from 4.8 ± 2.9 adducts per 107 nucleotides to 2.0 ± 1.8 adducts per 107 nucleotides after curcumin administration (63). Oral administration of 3.6 g curcumin significantly (P < 0.05) decreased inducible PEG2 production in blood 1 hour after curcumin administration as compared with the predosing level (62). The same study also showed the poor bioavailability and systemic distribution of curcumin. After encapsulated curcumin was administered in different amounts ranging from 0.45 to 3.6 g for 4 months, its biodistribution was examined by biopsy, which showed that malignant mucosal tissue had a higher concentration of curcumin while only a negligible amount was found in tissues outside the mucosa (61). This result may also be very beneficial for colorectal malignancy because any possible toxicity outside of the area of interest can be minimized. In 1 study where 5


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Table 1. Completed clinical trials using curcumin

<table>
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<th>Type</th>
<th>Materials and methods</th>
<th>Results and conclusions</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Phase I</td>
<td>Patients: 10; 2,000 mg/d + piperine 20 mg/kg</td>
<td>Piperine, a known inhibitor of hepatic and intestinal glucuronidation enhanced serum concentration, extent of absorption, and bioavailability.</td>
<td>Shoba et al. 1998 (59)</td>
</tr>
<tr>
<td>Safety trial</td>
<td>Much higher concentration with piperine at 0.25 to 1 hour post drug ($P &lt; 0.01$ at 0.25 and 0.5 hour; $P &lt; 0.001$ at 1 hour).</td>
<td></td>
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<tr>
<td>Phase I</td>
<td>Patients: 25; Oral 500–12,000 mg/d for 90 days</td>
<td>Oral curcumin is not toxic to humans up to 8,000 mg/d for 3 months.</td>
<td>Cheng et al. 2001 (60)</td>
</tr>
<tr>
<td>Safety trial</td>
<td>Bx done after treatment</td>
<td>Histologic improvement of precancerous lesions were observed in bladder cancer, oral leukoplaikia, intestinal metaplasia of the stomach, CIN, and Bowen disease</td>
<td></td>
</tr>
<tr>
<td>Phase I</td>
<td>Patients: 15, Oral curcumin extract of 440–2200 mg/d for 120 days.</td>
<td>Safe administration of curcumin extract at doses up to 2.2 g/day, equivalent to 180 mg of curcumin.</td>
<td>Sharma et al. 2001 (61)</td>
</tr>
<tr>
<td>Colon cancer</td>
<td>Activity of GST and levels of M1G were measured.</td>
<td>Curcumin has low oral bioavailability in humans and may undergo intestinal metabolism.</td>
<td></td>
</tr>
<tr>
<td>Phase I</td>
<td>Patients: 15, Oral 450–3,600 mg/d for 120 days.</td>
<td></td>
<td>Sharma et al. 2004 (62)</td>
</tr>
<tr>
<td>Colorectal cancer</td>
<td>Dose-escalation study.</td>
<td></td>
<td></td>
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<tr>
<td>Phase I</td>
<td>Patients: 12, Oral 450–3,600 mg/d for 7 days.</td>
<td>M1G levels were 2.5-fold higher in malignant tissue as compared with normal tissue ($P &lt; 0.05$ by ANOVA).</td>
<td>Garceo et al. 2005 (63)</td>
</tr>
<tr>
<td>Colorectal cancer</td>
<td>Bx samples of normal and malignant colorectal tissue, at diagnosis and at 6–7 hours after the last dose of curcumin.</td>
<td>The concentrations in normal and malignant colorectal tissue of patients receiving 3,600 mg of curcumin were $12.7 \pm 5.7$ and $7.7 \pm 1.8$ nmol/g, respectively. The daily dose of 3.6 g curcumin achieves pharmacologic efficacy in the colorectum with negligible distribution of curcumin outside the gut.</td>
<td></td>
</tr>
<tr>
<td>Phase I</td>
<td>Patients: 24, Oral 500–12,000 mg/day. Dose-escalation study for MTD and safety</td>
<td>Seven of 24 subjects (30%) experienced only minimal toxicity.</td>
<td>Lao et al. 2006 (64)</td>
</tr>
<tr>
<td>Safety trial</td>
<td>Systemic bioavailability of curcumin or its metabolites may not be essential for CRC chemoprevention because CRC can still benefit from curcumin.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Phase I</td>
<td>Patients: 14, Docetaxel (100 mg/m²) 1 hour i.v. every 3 weeks on day 1 × 6 cycles + Oral 500 mg/d for 7 consecutive days and escalated the dose until toxicity.</td>
<td>MTD at 8,000 mg/d</td>
<td>Bayet-Robert et al. 2010 (65)</td>
</tr>
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</table>
FAP patients who had undergone a prior colectomy were treated with the combination of curcumin and quercetin for 6 months, the size and number of adenomas were reduced significantly, supporting the use of curcumin for FAP colorectal cancer prevention (66). Administration of 4 g curcumin per day for 30 days significantly inhibited (40%) the number of ACF although the 2 g/d dose was found to be ineffective (68). Several patients with advanced pancreatic cancers also responded to 8 g/d of curcumin treatment (69). These promising results are very convincing but need to be validated with further larger-scale randomized trials. Table 2 shows ongoing clinical studies with curcumin.

### Biomarkers in Curcumin Chemoprevention Trials

Biomarkers can be very useful in identifying high-risk subjects for intervention, monitoring the effects of treatment, predicting outcome, and selecting patients who may benefit most from a given intervention. A well-validated biomarker may also serve as a surrogate endpoint to replace the current use of size reduction or histologic improvement of the precancerous lesion as the sole measure of success of chemoprevention; the use of a surrogate marker would potentially provide a more accurate assessment of outcome and would resolve the current difficulty in patient accrual due to the requirement for biopsy in chemoprevention clinical trials.

Although biomarkers for chemoprevention by curcumin have been extensively studied in cell culture and rodent models, only a few clinical studies have focused on biomarker modulation and attempted to correlate these with outcomes. In a recent pilot study, IKK\(\beta\) kinase activity and the levels of proinflammatory cytokine IL-8 in the saliva of SCCHN patients were measured and the results suggested that IKK\(\beta\) kinase activity could be used to detect the effect of curcumin treatment in SCCHN (70). In a double-blind randomized trial, curcumin was found to significantly decrease the levels of serum calcitonin gene-related peptide (CGRP) as compared with placebo (71). There were also significant decreases in serum IL-8 and high-sensitivity C-reactive protein (hs-CRP) in both the curcumin and placebo groups, with a higher magnitude in the curcumin group. In a placebo-controlled study, oral administration of curcumin significantly reduced erythrocyte malonaldehyde (MDA) and increased GSH levels in patients with tropical pancreatitis (72). Curcumin was also found to significantly decrease the serum levels of markers of oxidative damage (MDA and 8-hydroxydeoxyguanosine) and increase those of antioxidants (vitamins C and E) in patients with oral leukoplakia, oral submucous fibrosis, or lichen planus.

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**Table 1. Completed clinical trials using curcumin (Cont’d)**

<table>
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<th>Type</th>
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<th>Results and conclusions</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Open-label</td>
<td>VEGF, and tumor markers measured</td>
<td>8/14 patients had measurable lesions, with 5 PR and 3 SD.</td>
<td></td>
</tr>
<tr>
<td>Advanced metastatic breast cancer</td>
<td></td>
<td>Some biological and clinical responses were observed in most patients.</td>
<td></td>
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<tr>
<td></td>
<td>The recommended dose of curcumin was 8,000 mg/d for 7 consecutive days every 3 weeks in combination with a standard dose of docetaxel.</td>
<td></td>
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<tr>
<td>Phase II</td>
<td>Patients: 62, 1% ointment, several months for &quot;External cancerous lesion&quot;</td>
<td>The first clinical study.</td>
<td>Kuttan et al. 1987 (18)</td>
</tr>
<tr>
<td>Efficacy trial</td>
<td></td>
<td>Reduction in smell in 90% patients, reduction of itching in all cases, dry lesions in 70% patients, and reduction in lesion size and pain in 10% patients.</td>
<td></td>
</tr>
<tr>
<td>Skin lesion</td>
<td>Patients: 5, Oral curcumin 480g + quercetin 20 mg t.i.d. for 180 days.</td>
<td>Decrease in the number of polyps was seen in 60.4%.</td>
<td>Cruz-Correa et al. 2006 (66)</td>
</tr>
<tr>
<td>FAP</td>
<td>Polyps size and number assessed</td>
<td>Decrease in the size of polyps was 50.9% in FAP patients RCT in the future are necessary</td>
<td></td>
</tr>
<tr>
<td>Cohort study</td>
<td>Patients: 24</td>
<td>Zyflamend, a novel herbal anti-inflammatory mixture, as a potential chemopreventive agent in a phase I trial for patients diagnosed with PIN.</td>
<td>Rafailov et al. 2007 (67)</td>
</tr>
<tr>
<td>PIN</td>
<td>Patients: 44</td>
<td>40% reduction in ACF numbers with 4-g dose</td>
<td>Carroll et al. (68)</td>
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Table 2. Ongoing clinical trials with curcumin

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<th>Cancer type</th>
<th>Identifier number</th>
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<tbody>
<tr>
<td>Phase 1</td>
<td>Phase I Study of Surface-Controlled Water Soluble Curcumin (THERACURMIN CR-011L) in Patients With Advanced Malignancies</td>
<td>Advanced malignancies</td>
<td>NCT 01201694</td>
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<tr>
<td></td>
<td>Non-randomized</td>
<td></td>
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<tr>
<td>Phase 1</td>
<td>Phase I Pharmacokinetic Trial of Curcuminoids Administered in a Capsule Formulation</td>
<td>Colon cancer</td>
<td>NCT 00027495</td>
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<tr>
<td></td>
<td>Non-randomized</td>
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<tr>
<td>Phase 1</td>
<td>Phase I Clinical Trial Investigating the Ability of Plant Exosomes to Deliver Curcumin to Normal and Malignant Colon Tissue</td>
<td>Colon cancer</td>
<td>NCT 01294072</td>
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<td>Randomized controlled trial</td>
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<td>Phase 1</td>
<td>Crossover, Multiple Dose Pharmacokinetics of Two Curcumin Formulations in Healthy Volunteers</td>
<td>Healthy volunteers</td>
<td>NCT 01330810</td>
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<td>Phase 1</td>
<td>Curcumin Chemoprevention of Colorectal Neoplasia (Curcumin biomarker)</td>
<td>Colorectal cancer</td>
<td>NCT 01333917</td>
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<td>Non-randomized</td>
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<tr>
<td>Phase 1</td>
<td>Pilot Study of Curcumin, Vorinostat, and Sorafenib in Patients With Advanced Solid Tumors</td>
<td>Advanced solid tumor</td>
<td>NCT 01608139</td>
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<td>Phase 2</td>
<td>Phase II Double Blind Placebo-Controlled Trial of Curcuminoids’ Effect on Cellular Proliferation, Apoptosis and COX-2 Expression in the Colorectal Mucosa of Subjects With Recently Resected Sporadic Adenomatous Polyps</td>
<td>Colorectal cancer</td>
<td>NCT 00118989</td>
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<td>Phase 2</td>
<td>Phase II Trial of Curcumin in Cutaneous T-cell Lymphoma Patients</td>
<td>Cutaneous T-cell lymphoma</td>
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<tr>
<td>Phase 2</td>
<td>Phase II Trial of Curcumin in Patients With Advanced Pancreatic Cancer</td>
<td>Advanced pancreatic cancer</td>
<td>NCT 00094445</td>
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<td>Non-randomized</td>
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along with a significant decrease in pain and lesion size (73). Another clinical study suggested that cytokines and NF-κB pathway markers are important targets for curcumin chemoprevention (69). Other enzymes including COX-2 and hepatic GST nucleotidase have also been suggested for use in monitoring the effect of curcumin in chemoprevention studies (74). In addition, in a recent phase Ila clinical trial of curcumin chemoprevention in colorectal neoplasia, although the levels of PGE2 and 5-HETE did not significantly correlate with curcumin treatment, the amount of the premalignant lesion ACF was decreased (68); however, other studies showed marked modulation of PGE2 (62).

To clarify these ambiguous results, many more clinical studies testing different surrogate biomarkers in larger patient numbers must be carried out to overcome the limitations to the study of surrogate monitoring biomarkers. On the basis of these previous results, however, biomarkers of oxidative stress, NF-κB pathway markers, and cytokine levels in serum and tissues appear to be promising markers that new studies should be designed to measure.

**Hurdles: Pharmacokinetics and Pharmacodynamics**

Phase I/II clinical trials have clearly shown that curcumin exhibits poor bioavailability in humans, reaching approximately 1% after oral administration, a major barrier for its use in the clinic. The major factors contributing to the low plasma and tissue levels of curcumin appear to be its poor absorption due to insolubility in water, rapid systemic elimination in the bile and urine due to extensive enterohepatic recirculation, and fast metabolism (56). In fact, 40% of orally administered curcumin is excreted unchanged in the feces. To circumvent the bioavailability problem, numerous approaches have been considered, including structural modification or modification of the delivery system such as adding adjuvant, liposomal curcumin, curcumin nanoparticles, and phospholipid complex.

**Curcumin analogs**

Studies suggest that the β-diketone moiety is responsible for the instability and weak pharmacokinetic profile of curcumin. Modifications of the structure of natural curcumin significantly improved solubility, stability, and bioavailability. James Snyder’s group at the Emory University has synthesized a series of curcumin analogs by modifying the diketone moiety and the side chains of the benzene rings. Many of these compounds showed increased water-solubility and improved pharmacokinetic properties including tissue distribution and terminal elimination half-life (75). Analog HO3867 also showed tremendous improvement in cellular uptake and tissue distribution compared with its natural counterpart (76). Gagliardi and colleagues (77) synthesized more than 40 curcumin analogs and studied the bioavailability of some of these compounds in mice. One particular compound with a valine substitution at the phenyl ring showed more than 50-fold greater bioavailability compared with natural curcumin. A Japanese group also synthesized 86 different analogs of curcumin and determined their IC50 against 16 cancer cell lines. Many of these analogs, namely GO-Y078, 079, 030, 097, and 098, were at least 10-fold more potent than natural curcumin. This set of compounds is also more soluble in water, suggesting that they might show better bioavailability (78). Another synthetic curcumin known as dimethoxycurcumin exhibited significantly higher stability in vivo and against microsomal metabolism (79). In attempts to overcome the poor bioavailability of curcumin and to increase its tumor-specificity, many more innovative analogs have also been studied (Table 3; refs. 80–93).

**Curcumin nanoparticles**

Delivery of drugs via their formulation as nanoparticles is an emerging platform for an efficient approach to improve pharmacokinetic properties such as solubility and stability and, thus, bioavailability of poorly bioavailable drugs. This approach has been extensively used for curcumin with success in preclinical studies. Formulation of curcumin by encapsulation in polymeric micelles, liposomes, polymeric nanoparticles, lipid-based nanoparticles, and hydrogels makes the formulation watersoluble (56). Many of these formulations also showed improved bioavailability and pharmacokinetic properties in vivo. Encapsulation of
Table 3. Curcumin analogs and their benefits

<table>
<thead>
<tr>
<th>Analog</th>
<th>Study conclusions</th>
<th>Benefits and aims</th>
<th>References</th>
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<tbody>
<tr>
<td>EF24</td>
<td>In ovarian cancer cells, VEGF was dose-dependently reduced with EF24, showing ~ 8-fold greater potency than curcumin (P &lt; .05). Synergism with cisplatin. Novel strategy curcumin analog EF24 with a p38 inhibitor for lung cancer</td>
<td>Enhanced potency</td>
<td>Tan and colleagues (79)</td>
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<td>In MDA-MB231 and PC3, EF-24 inhibits HIF-1 and genuinely disrupts the microtubule cytoskeleton unlike curcumin</td>
<td>Mechanism</td>
<td>Thomas and colleagues (80)</td>
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<td>EF24 shows anticancer potency 10 times higher than curcumin, against lung, breast, ovarian, and cervical cancer cells by blocking the nuclear translocation of NF-kB</td>
<td>Enhanced potency</td>
<td>Thomas and colleagues (81)</td>
</tr>
<tr>
<td>EF31</td>
<td>EF31 has greater potency in NF-kB activity inhibition compared with curcumin and another analog EF24 and its action mechanism is based on its anti-inflammatory and antisurvival activities.</td>
<td>Enhanced potency</td>
<td>Olivera and colleagues (83)</td>
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<td>BDMCA</td>
<td>Chemopreventive effect through prevention of circulatory oxidative stress is not by methoxy group but by the terminal phenolic moieties or the central 7-carbon chain</td>
<td>Mechanism, structure, and roles</td>
<td>Devasena and colleagues (84)</td>
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<td>BDMCA is antioxidant and lipid peroxidation and antioxidant status could be used as markers for colon cancer chemoprevention using BDMCA</td>
<td>Mechanism and biomarker</td>
<td>Devasena and colleagues (85)</td>
</tr>
<tr>
<td>CDF</td>
<td>Combination of CDF and conventional 5-FU + oxaliplatin could be an strategy for preventing the emergence of chemoresistant colon cancer cells</td>
<td>Overcoming resistance</td>
<td>Kanwar and colleagues (86)</td>
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<td>CDF had better retention and bioavailability and the concentration of CDF in the pancreas tissue was 10-fold higher compared with curcumin</td>
<td>Improved bioavailability</td>
<td>Padhye and colleagues (87)</td>
</tr>
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<td>FLLL32</td>
<td>FLLL32 has biochemically superior properties and more specifically targets STAT3, a transcription factor</td>
<td>Enhanced specificity</td>
<td>Fossey and colleagues (88)</td>
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<tr>
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<td>FLLL32 reduced expression of STAT3-target genes</td>
<td>Enhanced specificity</td>
<td>Bill and colleagues (89)</td>
</tr>
<tr>
<td>GO-Y030</td>
<td>GO-Y030 has 30-fold higher potency in suppressing tumor cell growth compared with curcumin by inhibition of IKKβ</td>
<td>Enhanced potency</td>
<td>Sato and colleagues (90)</td>
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<td>Improved chemopreventive effect with GO-Y030 compared with curcumin (191 days). Diminished polyp incidence in Apc(S80D/−) mice fed GO-Y030.</td>
<td>Enhanced prevention</td>
<td>Shibata and colleagues (91)</td>
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curcumin in poly(lactic-co-glycolic acid) (PLGA) and PLGA–polyethylene glycol (PEG–PLGA–PEG) blend nanoparticles by a single-emulsion solvent-evaporation technique increased its mean half-life to approximately 4 and 6 hours, respectively, \( C_{\text{max}} \) by 2.9- and 7.4-fold, and bioavailability by 15.6- and 55.4-fold, respectively (94). Encapsulation of curcumin in poly(butyl) cyanoacrylate (PBCA) nanoparticles led to a 52-fold increase in elimination half-life and 2-fold increase in AUC (95). Curcumin encapsulated in MePEO-b-PCL micelles also showed similar improvements in pharmacokinetic parameters (96). Formulation of curcumin in solid-lipid nanoparticles also tremendously increased its bioavailability (more than 80-fold higher concentration in blood; ref. 97).

**Curcumin conjugates**

Conjugation of curcumin with polymers or other lipophilic compounds is another widely used approach to improve the water-solubility and stability of curcumin. Conjugation of curcumin with hyaluronic acid or polyvinylpyrrolidone forms water-soluble micelles with improved stability at physiologic pH and cytotoxic activities (98, 99). Polymerization of curcumin using diacid also produced a water-soluble curcumin polymer with improved anticancer activity (100). Complexation of curcumin with phosphatidylcholine also significantly (3–20 fold) improved its pharmacokinetic parameters, including bioavailability in animal models (101).

**Adding adjuvant**

One of the major reasons for the poor bioavailability of curcumin is its rapid glucuronidation. Protection of curcumin from such metabolic conversion using an adjuvant was found to be successful in improving its bioavailability. Piperine is an inhibitor of intestinal and hepatic glucuronidation. Concomitant administration of curcumin with piperine increased the bioavailability of curcumin by 2000% in human volunteers and 154% in rats (59).

**Future Possibilities**

High-risk individuals and cancer survivors alike may benefit from chemoprevention, not only because primary cancer chemoprevention is beneficial for high-risk groups but also because of the devastating nature of the disease course when patients experience SPT or recurrence. As curcumin is a non-prescription dietary derivative that has multiple targets at different levels in multiple pathways, it has great potential in the prevention of cancer and SPT. When its systemic bioavailability is increased through the development of different analogs and formulations, the promise of curcumin in chemoprevention may be feasible in many cancer types, not necessarily limited only to gastrointestinal cancers. A number of new analogs and formulations have already been developed with higher systemic bioavailability and potency. More standardized clinical trials for bioavailability and randomized control trials for efficacy should validate the potential of these newer agents and formulations. First, specific trials can improve the application of curcumin through changing the route of administration, achieve targeted delivery straight to the lesion sites by increasing tumor-specific affinity, and develop different analogs that can bypass or minimize the first-pass metabolism occurring in the gastrointestinal mucosa and liver. Second, to minimize its metabolism before reaching the targeted site, different preparations of curcumin may improve its delivery to the target and, therefore, increase its bioavailability. Third, formulating curcumin using nanoparticles and microparticles, which are among the most innovative modalities that can maximize delivery to a target tissue and increase sensitivity and specificity, may enhance its therapeutic index.

Defining the optimal precancerous candidates and surrogate endpoints to properly assess chemopreventive response is mandatory in chemoprevention research. Although we expect the network of signaling pathways to be considerably more complicated than we currently understand, further studies will better dissect the molecular effects of curcumin in different cancers. Specifically, microarray or recently developed RNASeq studies may be particularly valuable in defining unknown positive and negative signaling loops, and may represent a new field of future research directed at understanding the critical factors necessary for chemoprevention. In the future, targeting specific patient populations with certain biomarkers, so-called tailored chemoprevention, is necessary. Defining critical biomarkers
will help to better design a personalized plan for tailored chemoprevention. Progress in personal genome-based risk assessment and profiling of individual patients may also help to identify the patient population best suited to curcumin chemoprevention in the future.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Authors' Contributions
Conception and design: W. Park, A.R.M. Ruhul Amin, Z.G. Chen, D.M. Shin
Development of methodology: W. Park
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): W. Park
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): W. Park, A.R.M. Ruhul Amin

References

Writing, review, and/or revision of the manuscript: W. Park, A.R.M. Ruhul Amin, Z.G. Chen, D.M. Shin

Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): D. M. Shin

Study supervision: Z.G. Chen

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Curcumin in Cancer Prevention


Correction: New Perspectives of Curcumin in Cancer Prevention

In this review article (Cancer Prev Res 2013;6:387–400), which was published in the May 2013 issue of Cancer Prevention Research (1), the text within Table 1, a list of completed clinical trials using curcumin, contained a typographical error pertaining to curcumin dosage. Specifically, Table 1 referred to a phase II clinical trial conducted by Cruz-Correa and colleagues, with the following description: "Patients: 5, Oral curcumin 480 g + quercetin 20 mg t.i.d. for 180 days." The text incorrectly reported curcumin dosage in grams rather than milligrams. With this correction, the affected portion of Table 1 now accurately reads, "Patients: 5, Oral curcumin 480 mg + quercetin 20 mg t.i.d. for 180 days."

The online version of the article has been corrected and therefore no longer matches the print version. The authors regret this error.

Reference

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