Isoflavone Pharmacokinetics and Metabolism after Consumption of a Standardized Soy and Soy–Almond Bread in Men with Asymptomatic Prostate Cancer

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Abstract

Epidemiologic associations suggest that populations consuming substantial amounts of dietary soy exhibit a lower risk of prostate cancer. A 20-week randomized, phase II, crossover trial was conducted in 32 men with asymptomatic prostate cancer. The crossover involved 8 weeks each of soy bread (SB) and soy–almond bread (SAB). The primary objective was to investigate isoflavone bioavailability and metabolite profile. Secondary objectives include safety, compliance, and assessment of biomarkers linked to prostate carcinogenesis. Two distinct SBs were formulated to deliver approximately 60 mg aglycone equivalents of isoflavones per day. The isoflavones were present as aglycones (~78% as aglycones) in the SAB whereas in the standard SB predominantly as glucosides (18% total isoflavones as aglycones). Compliance to SB (97% ± 4%) and SAB (92% ± 18%) was excellent; toxicity was rare and limited to grade 1 gastrointestinal complaints. Pharmacokinetic studies between SB and SAB showed modest differences. Peak serum concentration time (tmax) was significantly faster with SAB meal compared with SB in some isoflavonoids, and AUC0–24h of dihydrodaidzein and O-desmethylangolensin was significantly greater after an SB meal. An exploratory cluster analysis was used to identify four isoflavone-metabolizing phenotypes. Insulin-like growth factor–binding protein increased significantly by 41% (P = 0.024) with soy intervention. Findings from this study provide the necessary framework to study isoflavone-metabolizing phenotypes as a strategy for identification of individuals that might benefit or show resistance to cancer preventive strategies using dietary soy. A standardized SB used for future large-scale randomized clinical trials to affect human prostate carcinogenesis is feasible. Cancer Prev Res; 8(11); 1045–54. ©2015 AACR.

Introduction

The impact of soy consumption on health has been frequently evaluated in both preclinical and human studies, yielding provocative results (1). One area of particular interest for applying a soy intervention is in the setting of prostate cancer. Initial epidemiologic observations suggested that populations consuming substantial amounts of dietary soy have a lower risk of prostate cancer (2). It is thought that the impact of soy on health or disease processes may be due to the nature of the soy protein fraction, or the unique array of non-nutrients referred to collectively as bioactive phytochemicals. Phytochemicals of interest include isoflavones, protease inhibitors, inositol hexaphosphate (phytic acid), lignans, phytosterols, and saponins (3). Isoflavones have been studied and demonstrate antiprostate cancer activity in vitro (4) and in rodent studies of carcinogenesis (5) and tumorigenesis (6). Potential mechanisms of action for soy isoflavones suggest multiple targets in prostate cancer cells that affect growth control and sensitivity to apoptosis. For example, the inhibition of IGFI-associated tyrosine protein kinase activity (4) or processes related to testosterone and androgen receptor activity (7), two of the most critical hormones involved in prostate development, function, and cancer. The tumor–host interaction may also be targeted via modulating angiogenesis (8), inflammation, or immune surveillance (9).

Epidemiologic and laboratory studies have stimulated human intervention studies with soy components to affect various biomarkers or physiologic processes relevant to prostate cancer. Several small studies are beginning to provide insight into bioactivity (10, 11). However, the largest human intervention study completed thus far, a study of biochemical failure in 177 post-prostatectomy patients, with randomization to a soy protein powdered drink (41 mg as aglycone equivalents of isoflavones per day for ~2 years) or similar product derived from the milk protein casein...
showed no impact on risk of biochemical (PSA) progression (12). We propose that soy will more likely affect earlier events in prostate carcinogenesis and that metabolism of soy, perhaps via host genetics and/or the microbiome may be a key cofactor.

Soy isoflavone absorption is thought to occur by passive absorption of the aglycones in the small intestine. However, debate continues regarding the bioavailability of isoflavone aglycones compared with their glucosides (13, 14, 15–17). Studies suggest that isoflavones administered in their glucoside forms (predominant in native soybeans) have greater (14), lesser (13, 16), or no difference in absorption as compared with their aglycone forms (15). Thus, understanding isoflavone bioavailability and postprandial pharmacokinetics (PK) remains an important question in soy literature.

Adding to the complexity in elucidating biologic impact of soy isoflavones, numerous metabolites of soy isoflavones can be formed in the human gut depending on the commensal bacteria in the large intestine (18, 19). Several factors influence the bacterial ecology of an individual’s colon, including: genetics (20), gender (21, 22, 23), biologic entrapment of isoflavones (24), diet composition (25), food matrix (21, 23, 26), and daidzein-degrading phenotype (25). The ability of an individual to produce certain isoflavone metabolites may be important to derive the health benefits from soy consumption (14). The metabolite equol has been of interest because it has selective estrogen receptor modulating activity in vitro (14). Similarly ODMA, another metabolite of daidzein, as well as equol, suppress prostate cancer growth in cell culture (27). Because there are several isoflavone metabolites in urine and plasma, a multifaceted biologic effect is possible. However, few studies have attempted to characterize isoflavone metabolism phenotypes by simultaneously analyzing several metabolites. These profiles could be used to identify individuals most likely to derive benefit from soy consumption.

This study characterizes the absorption and bioavailability of isoflavones from novel, chemically standardized soy (SB) and soy–almond bread (SAB) products. We propose that this is a critical step in defining an optimal and relevant delivery system for the evaluation of soy in human studies. We also hypothesize that bread with greater quantities of bioavailable isoflavones may improve isoflavone absorption and distribution. This may be particularly true for the majority (~60%) of the Western population who appear less capable of metabolizing daidzein conjugates to equol (14, 18). In the present study, we tested bread products in men previously treated for clinically localized prostate cancer but are now experiencing asymptomatic biochemical (PSA failure) recurrence, with no evidence of disease on standard staging studies. This study with standardized food products will also define possibly relevant patterns of absorption and metabolism within a potential target population for future longer-term intervention studies.

Materials and Methods

Preparation and isoflavone quantification of standardized SB and SAB

The standardization and production of soy breads (SB) used for this study are detailed in Supplementary Methods S1.

Participants

Study participants were enrolled from the Genitourinary Oncology Clinics at The Ohio State University Medical Center (Columbus, OH). Men had histologically documented prostate adenocarcinoma with a minimum of two increases in PSA after primary therapy or during active surveillance. Men were asymptomatic with no changes in hormone therapy planned for 5 months. Men were excluded if they had an active malignancy (other than prostate cancer) requiring medical therapy; abnormal kidney or liver function; active digestive illnesses or malabsorptive disorders, metabolic disorders requiring special diet modifications; endocrine disorders requiring hormone administration; known allergy to almonds, wheat, or soy; did not wish to discontinue dietary supplement use, or recent antibiotic use (<6 months before enrollment). All participants were regularly followed by a Medical Oncologist and would be suspended from participation if disease progression required more aggressive medical care. The study protocol was approved by The Ohio State University Comprehensive Cancer Center’s Clinical Scientific Review Committee and the Biomedical Sciences Cancer Institutional Review Board (NCT:01682941). Written and verbal informed consent was obtained from each enrolled participant.

Study design

This study was a 20-week, phase II, randomized, crossover design. Study visits were conducted at the Center for Clinical and Translational Science—Clinical Research Center. Each arm of the study was preceded with a 2-week washout (legume-free diet) and intervention commenced with a 24-hour PK study, which was then followed by an 8-week intervention with one of the two study breads. After completion of the first arm, participants then had crossed over to the alternate bread (Fig. 1A). A 2-week washout period is typical of feeding trials with soy isoflavone intervention (23). For each of the two PK studies (collected on days 0 and 70), fasted participants arrived at the research center, had an indwelling venous catheter placed and consumed two slices of study bread. Blood was collected immediately before (0 hours) and then at 2, 3, 4, 5, 6, 8, 10, and 12 hours after consumption of the test meal (Fig. 1B). Men were sent home after 12 hours and returned 24 hours after the test meal for a blood draw. The test meal consisted of either two slices of SB (167 ± 2 g) or SAB (167 ± 1 g) with water. The two slices of SB provided 404 kcal, whereas the SAB provided 420 kcal (Supplementary Table S2). Participants received legume-free meals while in the research unit and they were instructed to follow a legume-free diet for the duration of the study. During each 8-week intervention period, men were instructed to supplement their diet with 2 slices of study bread and document consumption in a journal. Frozen loaves of bread were distributed at each study visit with thawing instructions. Condiments were permitted on the bread, but heating was discouraged. Additional blood draws and 24-hour urine collections were completed at days 0, 28, 56, 70, 98, and 126. Men completed a 3-day diet record during each intervention period. All men were also provided a standardized multivitamin/mineral supplement (One Daily Multiple Plus Minerals, CVS Caremark, Woonsocket, RI) and asked to discontinue all other nutritional supplements for the duration of the study.

Chemicals

Anhydrous sodium acetate, and diethyl ether (Et2O) as well as HPLC or MS-grade acetonitrile, formic acid (≥98% purity), methanol, and water were purchased from Thermo Fisher Scientific. Lyophilized glucuronidase/sulfatase from Helix pomatia, DMSO, and equol were obtained from Sigma-Aldrich. Daidzein,
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Figure 1.
A, schematic of a 20-week randomized crossover design with two standardized SBs as dietary interventions. Gray boxes represent visits with oncologist. B, design of postprandial PK studies conducted on days 0 and 70. Details of the specific hours when blood was collected are represented with ▲ and controlled meals were served at the indicated hours.

genistein, glycine, daidzin, genistin, glycinin, acetyl daidzin, malonyl genistin, and acetyl genistin were from LC Laboratories division (PKC Pharmaceuticals, Inc.) and malonyl daidzin, and malonyl glycinin from Wako Chemicals USA, Inc. Isoflavone metabolites (dihydrodaidzein, dihydrogenistein, ODMA, and 6-OH-ODMA) were from Plantech.

Isoflavonoid quantification from blood
A 12-hour fasting blood was collected (K3EDTA, Vacutainer; Becton-Dickson), centrifuged (Sorvall Legend RT Kendro Laboratory Products) at 1,000 × g at 4°C for 10 minutes and plasma collected. Plasma was aliquoted and stored at −80°C until analysis. Extraction of isoflavones from plasma is detailed by Zuniga and colleagues (28) and a detailed description of the UPLC-MS/MS conditions is provided in Supplementary Methods S1.

Urine preparation for isoflavonoid analysis
Urinary collection (pre-weighed with 0.5 g/L boric acid) collected for more than 20 hours was considered complete. Urine volume was determined using mass and urine specific gravity. Aliquots of urine were stored at −80°C until HPLC analysis. Extraction of isoflavonoids in urine was adapted as described previously by Zuniga and colleagues (28).

Quantification of isoflavonoids in urine
HPLC instrumentation for urine analyses was the same as described for soy food analysis (29). Symmetry C8 column (4.6 × 75 mm, 3.5 μm; Waters Associated) affixed to a Symmetry C8 guard column (3.0 × 20 mm, 3.5 μm) was used for separation of isoflavonoids extracted from urine. The binary mobile phase was 0.1% aqueous formic acid (solvent A) and 0.1% formic acid in 90% methanol and 10% acetonitrile (solvent B) with a flow rate of 1.3 mL/min. The linear gradient began at 95:5 (solvent A: solvent B), increased to 65:35 in 3 minutes, 42:58 in 12 minutes, 10:90 at 14 minutes, and then returning to 95:5 at 18 minutes, for a total run time of 18 minutes. The interassay and intraassay variability in urine as well as plasma was less than 15%. Urine concentrations less than 46 nmol/L (12.4 ng/mL) to 56 nmol/L (14.2 ng/mL) for the parent (daidzein, genistein, and glycine) isoflavonoids and less than 45 nmol/L (12.3 ng/mL) to 60 nmol/L (15.6 ng/mL) for intermediate (DHG and DHD) isoflavonoids were considered below the level of quantification. Moreover, urine concentrations less than 125 nmol/L (30 ng/mL) for equol, 85 nmol/L (21.9 ng/mL) for O-DMA, and 137 nmol/L (39.5 ng/mL) for 6’OH-ODMA were considered below the level of quantification.

Laboratory and clinical measures of toxicity
Side effects were recorded and scored on the National Cancer Institute Common Toxicity Criteria for Adverse Events (CTCAE v. 4.0; ref. 30) and serum was collected for PSA analysis at all clinic visits. Enrolled men were seen (days 56 and 126) by their attending oncologist for routine assessment of clinically detectable disease and blood (complete blood count with differential, electrolytes, and liver function tests) was collected to evaluate toxicity. All clinical blood samples and PSA were analyzed by OSU’s Wexner Medical Center clinical laboratories. Because soy protein has been associated with altering lipid metabolism, fasting serum lipids (total cholesterol, LDL, HDL, and triglycerides), and endocrine biomarkers (human growth hormone, IGF1, IGFBP-3, and leptin) were analyzed by the CRC core laboratories using either chemiluminescence or radioimmunoassay methods.
with a Dimension Xpand Clinical Chemistry analyzer (Siemens Medical Diagnostics). The Intraassay%CV for lipids and biomarkers were <2.31% and the interassay%CV was <2.35%.

Calculations and statistical analysis
Isoflavonoid absorption in plasma during the 24 hours PK study was assessed using maximum concentration (C\text{max}), time to maximum concentration (T\text{max}), and area under the curve (AUC\text{0 to 24 hours}) using trapezoidal approximation.

Statistical analysis was conducted using SPSS version 19.0 software (IBM) and Stata v10.1 (StataCorp LP). Descriptive analysis was used to determine mean ± SD for food samples and mean ± SE for clinical data, laboratory data, dietary intake of SBs, and isoflavonoids in biologic samples. An independent t test determined significant differences in isoflavone composition between SB and SAB. For all analyses, P values ≤ 0.05 were considered significant. A repeated measures ANOVA was used to assess differences due to treatment, period, and time in plasma and urine isoflavonoids. If significant effects (P ≤ 0.05) were found, then Tukey post hoc analysis was performed. An exploratory cluster analysis was performed using a hierarchical agglomerative technique using Euclidean distances to identify clusters of daidzein-metabolizing phenotypes. Because of variation in the total amount of daidzein metabolites across patients, proportions were used in the clustering to identify relative contributions of the constituents. In addition, the intraclass correlation coefficient was used to evaluate the consistency of the daidzein metabolites over time.

Results
Standardized SB and SAB have respectively high glycoside and high aglycone forms of isoflavones
The isoflavone composition in the starting soy mix used for both breads was virtually the same (P > 0.80; Fig. 2A). As predicted, because of the presence of glucosidase activity from almonds, the bread making process transformed the chemical composition of the isoflavones to be significantly different between SB and SAB yet the total isoflavone content was preserved (Fig. 2B). Isoflavones in one slice of SB was 34.9 ± 6.0 mg AE of isoflavones (18% aglycones) and SAB was 34.7 ± 4.8 mg AE isoflavones (78% aglycones). The isoflavone composition and content was found to remain be stable through 2 years of storage (−25°C).
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Participant adherence

Of the 32 men (52–84-years-old) enrolled in this study, 25 completed all study activities, detailed demographics in Supplementary Table S3. Three men withdrew from the study due to undisclosed personal reasons, 4 men required more aggressive prostate cancer therapy, and 1 man was diagnosed with a second malignancy requiring therapy. Of the 7 men who withdrew from the study, 4 completed all visits through day 70. Adherence to legume-restricted diet and interventions was excellent (SB = 97 ± 4% and SAB = 92 ± 19%; Supplementary Table S3). Isoflavonoid excretion in 24-hour urine collections were used to verify adherence to the legume-free diet and SB interventions. A significant increase (P < 0.001) in total isoflavonoid excretion was observed in urine on days 28 (24.83 ± 2.00 mg/24 hours), 56 (23.01 ± 2.01 mg/24 hours), 98 (23.88 ± 1.53 mg/24 hours), and 246 (22.49 ± 1.35 mg/24 hours) of the intervention compared with the wash-out days (day 0 = 0.12 ± 0.03 mg/24 hours and day 70 = 0.13 ± 0.05 mg/24 hours) confirming that dietary compliance was excellent and SBs were effective vehicles for isoflavone delivery. Likewise, the total isoflavone concentration in urine among the intervention visits did not significantly differ (P = 0.337). Daidzein (65% of total, 16.9 mg total daidzein isoflavonoids/25.8 mg dietary daidzeins) and genistein (19% of total, 7.0 mg total genistein isoflavonoids/36.3 mg dietary genisteins) was present in urine of all men.

Isoflavone PK after SB and SAB meals show modest differences independent of bread type, parent isoflavonoids (daidzein, genistein, and glycitein) were first detected in plasma at 2 hours after test meals (Supplementary Fig. S4). Isoflavone metabolites, dihydrodaidzein and dihydrogenistein, were observed in plasma starting at 3 hours, and microbial metabolites of daidzein, ODMA and equol, were observed in plasma starting at 4 hours after the test meal (Supplementary Fig. S4). The time to reach peak serum concentration (T_{max}) of daidzein, dihydrodaidzein, genistein, and 6’hydroxy-O-desmethylangolesin (6’OH-ODMA) was significantly more rapid (P < 0.05) following the meal with SAB compared with the SB meal (Supplementary Fig. S4). Moreover, peak absorption (C_{max}) of genistein (P = 0.025) was significantly higher after a SB meal compared with SB or SAB (Supplementary Table S3). The diarrhea resolved when men abstained from consuming SBs after symptoms resolved. Although bloating returned for some men, mild gastrointestinal symptoms improved by the second treatment period. No toxicity greater than grade 1 was observed. Over the course of this study, a significant decrease in BMI and weight (P < 0.001) was observed. When stratified by bread intervention, weight loss predominately occurred during the SAB intervention period (2.5 ± 0.3 kg) compared with the SB (0.5 ± 0.7 kg; P = 0.011) intervention (Supplementary Table S3).

Chemistry, liver enzymes, and complete blood count were not affected by dietary intervention. PSA was monitored during this 20-week intervention (Table 1). Although not a primary outcome in this small study, we observed a 3-fold prolongation in PSADT during the soy intervention period (Table 1) leading to a leveling of PSADT (Supplementary Fig. S5). Cholesterol (total, HDL, and LDL) and triglycerides were not significantly affected over the entire 20 weeks study (Table 1). However, among men who were hypercholesterolemic at enrollment and who were not on cholesterol-lowering agents (n = 7), a significant decrease (P = 0.026) in total cholesterol (day 0, 229 ± 10 mg/dL and day 126, 207 ± 7 mg/dL) and LDL cholesterol (day 0, 150 ± 9 mg/dL and day 126, 132 ± 9 mg/dL) was observed between days 0 and 126. A significant increase in insulin-like growth factor–binding protein 3 (IGFBP3) was observed in our cohort of men, and a significant decrease in proinflammatory cytokines, which is detailed in a parallel publication (31).

SB and SAB intervention was safe with minimal side effects

Using standard National Cancer Institute CTCAE version 4.0, mild (grade 1) gastrointestinal side effects were the most common adverse event in this study. Ten men (40%) complained of bloating and 4 men (16%) complained of diarrhea while consuming SB or SAB (Supplementary Table S3). The diarrhea resolved when men abstained from consuming SBs after symptoms resolved. Although bloating returned for some men, mild gastrointestinal symptoms improved by the second treatment period. No toxicity greater than grade 1 was observed. Over the course of this study, a significant decrease in BMI and weight (P < 0.001) was observed. When stratified by bread intervention, weight loss predominately occurred during the SAB intervention period (2.5 ± 0.3 kg) compared with the SB (0.5 ± 0.7 kg; P = 0.011) intervention (Supplementary Table S3).

Cluster analysis used to explore individual variability and isoflavone-metabolizing phenotypes

An exploratory cluster analysis was used to characterize isoflavone-metabolizing phenotypes in patients. From this analysis using daidzein metabolites from 24 hours urine collections, four clusters emerged consistently over four different time points. Relative to the other three clusters, cluster 1 (n = 7) as demonstrating the highest excretion of parent (daidzein and genistein) and intermediate (DHD and DHG) isoflavonoids in their urine (Fig. 3). ODMA production in cluster 1 was modest and inconsistent and excreted ODMA in relatively low levels (0.40 ± 0.16 mg/24 h). Cluster 2 (n = 12) showed the greatest concentration of ODMA in the urine (3.97 ± 0.35 mg/24 h), but very limited amounts of equol (0.31 ± 0.13 mg/24 h) among the equol producers. Men in cluster 3 (n = 3) had similar proportions of equol (5.46 ± 0.47 mg/24 h) and ODMA (4.48 ± 0.78 mg/24 h). Cluster 4 (n = 3) also had the greatest excretion of equol (8.84 ± 0.98 mg/24 h) and very low levels of ODMA (0.12 ± 0.08 mg/24 h).

Isoflavonoids from plasma were stratified using the same four clusters generated from the urine isoflavonoid profiles. The plasma isoflavonoid profiles generated from 24 hours AUC showed a similar pattern to that profiled from 24 hours urine (Fig. 4). Those in cluster 1 show the highest plasma daidzein (18.42 ± 4.84 μmol/24 h·L) and genistein (31.34 ± 6.49 μmol/24 h·L) AUC. The ODMA producers in cluster 1 reached peak concentrations of ODMA at 12.5 ± 2.0 hours, whereas the other clusters reached peak concentrations of ODMA in approximately 7 hours. Cluster 2 demonstrates the greatest AUC for ODMA (3.98 ± 0.94 μmol/24 h·L) overall, and lowest AUC of equol (0.26 ± 0.13 μmol/24 h·L) compared with those producing equol but categorized in other clusters. Men in cluster 3 demonstrate a similar metabolic profile as cluster 2. Yet, the men in cluster 3 compared with those in cluster 2 show a higher DHD AUC (3.91 ± 1.43 μmol/24 h·L), a lower ODMA AUC (2.56 ± 0.94 μmol/24 h·L), and produced equol more consistently and with greater AUC (0.58 ± 0.14 μmol/24 h·L). The plasma isoflavonoid PK revealed that men in cluster 4 had the fastest rate of absorption and production of metabolites with differences ranging between 1 and 10 hours. Cluster 4 reached peak equol concentrations in 9 hours, whereas clusters 2 and 3 began to peak at 24 hours, thus contributing to cluster 4 having the greatest AUC (4.04 ± 1.28 μmol/24 h·L) for equol.
Figure 3.
Proportion of daidzein isoflavonoids from 24 hours urine collections (mg/24 hours) taken during four time points over 20 weeks (n = 25), left. Cluster analysis determined four distinct clusters and their respective isoflavonoid profile, right. White bars indicate urine samples not collected. Significant differences (P < 0.05) between metabolizing phenotype clusters were determined using ANOVA, reported using an asterisks. ODMA, O-desmethylangolensin; DHD, dihydrodaidzein; DHG, dihydrogenistein.
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Discussion

Future studies testing relevant hypotheses regarding the biologic impact of soy in the human diet will be most informative if fully characterized, convenient, and tasteful food products are used that provide phytochemical exposures similar to that observed in many Asian populations. To this end, we successfully created two SBs containing a diverse array of bioactive phytochemicals at concentrations typically observed in Asian diets. We also demonstrate that food processing techniques can be used to rationally design foods with altered isoflavone chemistry to favor absorption and subsequently, may enhance their bioactivity (32). The two breads tested in this study have complementary compositions of isoflavones, one rich in aglycone forms (SAB) and the other rich in glucoside forms (SB) of isoflavones (29). Most critically, this study demonstrates that characterizing an investigational food product, both chemically and with in vivo PK and safety data, should be a key step toward defining a relevant agent for human intervention trials for cancer, other disease outcomes, and health.

Our study carefully documents the impact of food matrix on isoflavone metabolism, a concept previously addressed in a few studies using liquid and solid forms of soy foods (21, 26, 23). The PK measures ($T_{\text{max}}$, $C_{\text{max}}$, and AUC) found in our study with SB were similar to those reported by Cassidy and colleagues (21) after a textured vegetable protein (TVP) meal. In that study, the $T_{\text{max}}$ of daidzein and genistein from soy milk was significantly faster compared with TVP and tempeh yet no significant differences in daidzein and genistein $T_{\text{max}}$ and $C_{\text{max}}$ observed between the two solid matrices (TVP and tempeh). In our previous study and in Franke and colleagues (26), isoflavone metabolites in urine were significantly greater with either SB (23) or soy nuts compared with a soy-based beverage intervention. Furthermore, aglycone composition is reported to affect isoflavone PK. Aglycone composition among soy foods varies widely, ranging from 12% in soy bars (26) to 50% in tempeh (21) and among the few studies that have
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Table 1. Changes in laboratory measures during the 20-week studya

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Day 0</th>
<th>Day 126</th>
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<tbody>
<tr>
<td>PSA (ng/mL)</td>
<td>9.36 ± 2.46</td>
<td>15.93 ± 3.45</td>
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<td>PSA velocity (ng/mL/mo)b</td>
<td>1.64 ± 0.42</td>
<td>2.12 ± 0.84</td>
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<td>PSADT (mo)c</td>
<td>13.5 ± 3.4</td>
<td>27.6 ± 15.5</td>
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<tr>
<td>Lipids Overall</td>
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<tr>
<td>Total cholesterol (mg/dL)</td>
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<td>169 ± 7</td>
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<tr>
<td>HDL (mg/dL)</td>
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<td>50 ± 4</td>
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<tr>
<td>LDL (mg/dL)</td>
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<td>103 ± 6</td>
</tr>
<tr>
<td>Triglycerides (mg/dL)</td>
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<td>139 ± 12</td>
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<tr>
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<td>cholesterol (mg/dL)</td>
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</tr>
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<td>LDL (mg/dL)</td>
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<tr>
<td>hGH (ng/mL)</td>
<td>1.16 ± 0.29</td>
<td>1.02 ± 0.31</td>
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<tr>
<td>IGf (mg/mL)</td>
<td>121.4 ± 6.5</td>
<td>125.2 ± 7.2</td>
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<tr>
<td>IGFBP-3 (µg/mL)</td>
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<td>3.51 ± 0.87</td>
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<td>IGF1/IGFBP-3 ratio</td>
<td>35.2 ± 0.9</td>
<td>35.5 ± 0.8</td>
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<tr>
<td>Leptin (ng/mL)</td>
<td>12.96 ± 2.1</td>
<td>13.51 ± 2.0</td>
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</table>

Abbreviations: PSADT, PSA-doubling time; HDL, high-density lipoprotein; LDL, low-density lipoprotein; hGH, human growth hormone; Igf, insulin-like growth hormone; Igfbp-3, insulin-like growth hormone-binding protein.

aMean ± SD (n = 25, men completed study).
bDay 0 (PSA values collected before but not including day 0); day 126 (PSA from enrollment to and including day 126).
cSignificance (P ≤ 0.05) was determined using ANOVA.

evidence that agarlycones are readily absorbed in the small intestines. Thus, intestinal beta glucosidase and lactase phlorizin hydrolase activity appear to be proficient in liberating isoflavone aglycones for absorption. Standardized breads used in our clinical trial can in the future be tailored to favor absorption of isoflavone aglycones or metabolites to enhance health outcomes.

A number of soy isoflavone metabolites can be formed by the intestinal microbiota, a variable that is only beginning to be understood. Different proportions of isoflavone metabolites constitute the metabolic phenotype and most likely affect downstream biologic responses. Daidzein can be metabolized in sequence to DHD to equol or ODMA (36). Similarly, genisteen has been reported to be metabolized into dihydrogenistein and 6’OH-ODMA (19, 36). Previous studies suggest that isoflavone metabolites such as equol may have greater antioxidant activity (37) and may be more estrogenic (38) than their parent forms, but the unique role(s) of specific isoflavones, isoflavone patterns, or metabolites in prostate carcinogenesis is very limited and warrants further investigation. A strength of the present study, unlike earlier studies of isoflavone absorption, is the inclusion of a comprehensive investigation of plasma and urine isoflavone metabolites (DHD, DHG, ODMA, equol, and 6’OH-ODMA) over a 24-hour period.

Early studies have characterized and defined daidzein-metabolizing phenotypes either by those who produce equol (14) or those who produce ODMA (19) or by their rate of isoflavonoid degradation (25). Moreover, previous studies (39) have identified bacteria responsible for O- DMA production is distinct from those who produce equol (40, 41). In our cohort of men, cluster analysis was used to elucidate similarities and differences among individuals who produced daidzein and genistein metabolites. Using this approach, we discovered that among equol producers (cluster 3 and 4) there are individuals who produce relatively high levels of ODMA and those who produce low amounts. Moreover, we observed a cluster (cluster 1) that did not produce terminal metabolites of daidzein (equol or ODMA) yet they had the highest level of genistein in plasma and urine. In addition, over the 20-week investigation, we observed a high intraclass correlation (equol 91%, ODMA 83%, and daidzein 89%) in these isoflavone-metabolizing phenotypes, which align with previous studies suggesting a high concordance (~80%) over time in Americans (42) and Japanese (43). Distinguishing these isoflavonoid-metabolizing clusters may help to explain interindividual differences and assist in identifying those individuals that might be more or less responsive to soy intervention, which has important implications for designing future cancer prevention trials.

Of the many purported benefits of soy, the most well-established benefit is lowering blood cholesterol, which is mostly attributed to soy protein, and which forms the basis for the approved FDA claim (44). Although no overall changes in blood lipids were observed in our cohort, a subset with untreated hypercholesterolemia at enrollment had a significant decrease in total cholesterol and LDL cholesterol. Our earlier studies with a similar SB demonstrated significant decrease in LDL and triglycerides in men with hypercholesterolemia (23). Platz and colleagues (45) reported a potential relationship between lower cholesterol or use of statins and inverse lower risk of prostate cancer-related mortality. The significant decrease in lipid parameters in response to soy and the associated prolonged PSADT in this same cohort was an incidental finding and a causal relationship would need extensive additional investigation.
Meta-analyses suggest that high circulating levels of IGF is associated with increased risk of prostate cancer (46) and predictive of advanced disease (47) However, dietary effects on IGF have been mixed. Earlier studies with rat prostate cancer lines treated with genistein demonstrated a decrease in IGF whereas dietary intervention trials with soy isolavones in men with prostate cancer had shown no significant change in IGF (48) or increase in IGF (49). Although in our cohort there was no significant increase in IGF, there was a significant increase in insulin-binding protein 3 (IGFBP 3) with soy intervention. IGFBP 3 has been shown to inhibit the proliferation of prostate cancer cell lines independent of IGF by behaving as a potent inducer of apoptosis (50). Moreover, there was a concomitant decrease in the ratio of IGF to IGFBP-3 in 64% (16/25) of our patients.

Conclusion

In conclusion, food technology can be used to produce fully characterized food products to assess dietary hypotheses in cancer. We produced two SB products that can be easily incorporated into the diet of men consuming a typical Western diet on a daily basis. We have achieved a dose of soy phytochemicals in two slices of bread per day that will mimic typical soy intake in Asian populations. This study documents the adherence, safety, and bioavailability of isolavones from two fully characterized SB products; data that are essential to enhance our understanding of potential mechanisms where SBs may affect biologic processes. Evaluation of plasma isolavonoids and isolavonoids in urine suggests that differences between the aglycone and glycoside composition in the SBs had a modest impact on the overall rate of isolavone absorption. This study is the first to provide a comprehensive approach to characterizing isolavonoid phenotypes using both plasma PK and urine steady states. We have identified four isolavone metabolite–producing phenotypes. Strategies to identify metabolic phenotypes of soy isolavone intervention are critical to help decipher heterogeneity in biologic responses among individuals in clinical studies. The tremendous collaborative effort invested in standardizing SBs for clinical studies have led four isolavone proles with de and optimized palatability that will serve as an excellent source of soy isolavones for future large-scale cancer prevention trials.

Disclosure of Potential Conflicts of Interest

Y. Vodovoz has ownership interest (including patents) in U.S. patent 7,592,028. No potential conflicts of interest were disclosed by the other authors.

Disclaimer

Any opinions, findings, and conclusions expressed in this article are those of the authors and do not necessarily reflect those of the Pelotonia Fellowship Program or of the National Center for Advancing Translational Sciences.

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