Lack of ABCG2 shortens latency of BRCA1-deficient mammary tumors and this is not affected by genistein or resveratrol

Serge A.L. Zander¹, Ariena Kersbergen¹, Wendy Sol¹, Maaike Gonggrijp¹, Koen van de Wetering¹, Jos Jonkers², Piet Borst¹, Sven Rottenberg¹

Authors' affiliations:
Divisions of ¹Molecular Oncology and ²Molecular Pathology, The Netherlands Cancer Institute (Antoni van Leeuwenhoek Hospital), Plesmanlaan 121, 1066 CX Amsterdam, The Netherlands

Running title:
ABCG2 loss shortens latency of BRCA1-deficient tumors

Keywords: resveratrol, genistein, tumor latency, mouse model, ABCG2

Grant support:
This work was supported by grants from the Dutch Cancer Society 2006-3566 (PB, SR, JJ), 2009-4303 (SR, JJ, PB), the European Union FP6 Integrated Project 037665-CHEMORES (PB, SR), CTMM Breast Care (JJ, SR) and ZonMw 40-00812-98-07-028 (KvdW, PB). SR is supported by The Netherlands Organisation for Scientific Research (NWO VIDI-91711302).

Corresponding author:
Sven Rottenberg, Division of Molecular Oncology, The Netherlands Cancer Institute (Antoni van Leeuwenhoek Hospital), Plesmanlaan 121, 1066 CX Amsterdam, The Netherlands. Phone: +31 20 512 2082; Fax: +31 20 669 1383; E-mail: s.rottenberg@nki.nl

Disclosure of Potential Conflicts of Interest:
No potential conflicts of interest were disclosed.

word count: 3984
total number of figures and/or tables: 3
Abstract

In addition to their role in drug resistance, the ATP binding cassette (ABC) transporters ABCG2 and ABCB1 have been suggested to protect cells from a broad range of substances that may foster tumorigenesis. Phytoestrogens or their metabolites are substrates of these transporters and the influence of these compounds on breast cancer development is controversial. Estrogen-like properties might accelerate tumorigenesis on the one hand, while their proposed health-protective properties might antagonize tumorigenesis on the other. To address this issue, we used a newer generation mouse model of BRCA1-mutated breast cancer and examined tumor latency in K14cre,Brca1F/F,p53F/F; Abcb1a/b−/−,K14cre,Brca1F/F,p53F/F or Abcg2−/−,K14cre,Brca1F/F,p53F/F animals, fed with genistein- or resveratrol-supplemented diets. Ovariectomized K14cre,Brca1F/F,p53F/F animals were included to evaluate whether any estrogen-mimicking effects can restore mammary tumor development in the absence of endogenous estrogens. Compared with the ABC transporter proficient model, ABCG2-deficient animals showed a reduced median tumor latency of 17.5 days (P<0.001), whereas no significant difference was observed for ABCB1-deficient animals. Neither genistein nor resveratrol altered this latency reduction in Abcg2−/−,K14cre,Brca1F/F,p53F/F animals. Ovariectomy resulted in nearly complete loss of mammary tumor development, which was not restored by genistein or resveratrol. Our results show that ABCG2 contributes to the protection of genetically instable epithelial cells against carcinogenesis. Diets containing high levels of genistein or resveratrol had no effect on mammary tumorigenesis, whether mice were lacking ABCG2 or not. Since genistein and resveratrol only delayed skin tumor development of ovariectomized animals, we conclude that these phytoestrogens are no effective modulators of mammary tumor development in our mouse model.
Several ATP binding cassette (ABC) transporters act as cellular efflux pumps of drugs and cause multidrug resistance (1). In addition to this classical role, ABC transporters might have other functions in tumor biology (2). For instance, loss of ABC transporters has been claimed to decrease, as well as increase carcinogenesis. Mochida et al. (3) reported in heterozygous \( Apc^{Min/+} \) mice, prone to develop intestinal malignancies, that loss of P-glycoprotein (P-gp/MDR1/ABCB1) decreased tumor formation. They disrupted the \( Abcb1a \) gene and found decreased intestinal polyp and tumor incidence compared with \( Abcb1a \) wildtype mice, suggesting that P-gp-mediated protection from xenotoxins allows epithelial cells with strong driver mutations to survive and progress into malignant tumors. This protective effect of P-gp may be specific for mice, as tumor-initiating cells of human colorectal tumors do not appear to be protected by ABCB1 (4). In contrast, Gupta et al. (5) found loss of ABCG2 protein and mRNA expression in human colorectal and cervical cancers and they hypothesized that increased exposure to carcinogenic genotoxins in premalignant cells might enhance tumor evolution through stimulated mutagenesis. Fletcher et al. (2) also discuss active transport-independent functions of ABC transporters in apoptosis and tumor cell proliferation, but no mechanisms for these functions have been established. Taken together, these studies illustrate that we still do not fully understand what roles ABCB1 and ABCG2 play in protecting normal cells from carcinogenic xenobiotics.

Some members of the phytoestrogen family are ABCG2 substrates, and it has been suggested that these compounds competitively inhibit ABC transporter-mediated drug transport (6-8). In addition, phytoestrogens have received attention because epidemiological studies attributed the lower incidence of breast cancer in Asian countries to their traditional diets, consisting of phytoestrogen-rich soy food (9). A major phytoestrogen present in soy is the isoflavone genistein. Another phytoestrogen with putative health-promoting effects is the stilbenoid resveratrol, present in red grapes (10). Numerous studies impart preventive effects of resveratrol on cancer development, including breast cancer. To explain the benefit of phytoestrogen consumption various mechanisms have been suggested. These include inhibition of oncogenic signaling pathways, topoisomerasers, cyclooxygenases, angiogenesis, proliferation, or endogenous estradiol production; SIRT1 activation; scavenging of oxygen radicals; and anti-apoptotic effects (10-12). Consequently, phytoestrogens have become popular as dietary supplements. Given their claimed chemopreventive features, phytoestrogens may in particular appear attractive to women with an increased risk of developing breast cancer, such as carriers of \( BRCA1 \) or \( BRCA2 \) mutations.
The polyphenolic phytoestrogens genistein and resveratrol share structural characteristics with 17β-estradiol and therefore interact with the mammalian estrogen receptors, although with lower binding affinity than estradiol itself (13, 14). Such estrogenic effects of phytoestrogens could alleviate discomforts of menopausal women and supplementing the diet of these women with phytoestrogens might appear an attractive alternative to estrogen-replacement therapy, which promotes breast tumorigenesis (15). Like estrogen-replacement therapy, however, substantial estrogen-mimicking effects of phytoestrogens might also increase the risk of carcinogenesis, especially in women who have a breast cancer predisposition. Despite several years of research, the disagreement on the positive versus negative effects of phytoestrogen-rich diets on breast cancer development remains (16, 17). This question has been addressed using rodent models of chemically induced or xenografted tumors, but the data reported are ambiguous and did not resolve the controversy (reviewed in (12, 18)).

To tackle these issues from a different angle, we investigated the effect of genistein or resveratrol in a newer generation mouse model. $K14\text{cre};Brca1^{F/F};p53^{F/F}$ mice lose the tumor suppressor function of BRCA1 and p53 stochastically in mammary (and skin) epithelial cells early during development. BRCA1 dysfunction results in genomic instability and drives additional mutagenic events that result in spontaneous tumors when the mice are about 7 months old (19). The mammary tumors highly resemble BRCA1-associated breast cancer in humans and, like their human counterparts, these mouse tumors are “triple-negative”, lacking expression of estrogen, progesterone or HER2 growth factor receptors. Despite this absence of hormone receptor expression, tumorigenesis is nevertheless estrogen-dependent: when young $K14\text{cre};Brca1^{F/F};p53^{F/F}$ mice are ovariectomized, mammary tumor development is abolished, but can be restored when estradiol is re-introduced (van de Ven et al., submitted for publication). This conditional mouse model, therefore, represents a suitable system to investigate whether any estrogenic effects of genistein- or resveratrol-rich diets affect mammary tumor formation. In particular, we studied the ability of phytoestrogens to restore mammary tumorigenesis in ovariectomized animals. The exposure of premalignant mammary epithelial cells to genistein or resveratrol might also be influenced by the ABC transporters ABCB1 or ABCG2. It has been shown that ABCG2-deficient animals have increased resveratrol or genistein plasma levels compared with wild-type mice (20-22). Hence, to increase the exposure of mammary epithelial cells to genistein or resveratrol, we also investigated mammary tumor formation in $Abcb1a/b^{-/-};K14\text{cre};Brca1^{F/F};p53^{F/F}$ and $Abcg2^{-/-};K14\text{cre};Brca1^{F/F};p53^{F/F}$ females.
Materials and methods

Mice and special diets

To study the effect of ABC transporters on phytoestrogen disposition, the K14cre;Brca1F/F;p53F/F mouse model (19) was crossed with Abcg2-/- (23) and Abcb1a/b-/- (24) mice on the same FVB/N genetic background to generate animals that develop spontaneous mammary tumors lacking functional ABCG2 or ABCB1A and ABCB1B. The Abcg2-/-;K14cre;Brca1F/F;p53F/F model was generated by backcrossing FVB.129P2-Abcg2tm1AhsN7, FVB-Tg(KRT14-cre)8Brn, FVB.129P2-Trp53tm1Bm, or FVB.129P2-Brca1tm1Bm mice on FVB/N animals for at least 8 generations (the first 5 generations using marker-assisted breeding) and eventually crossing these animals to generate the FVB.Cg-Abcg2tm1Ahs Trp53tm1Bm Brca1tm1Bm Tg(KRT14-cre)8Brn/A compound mice. The Abcg2Δ3-6/Δ3-6 genotype was confirmed by PCR with specific primers, as described previously (25). The Abcb1a/b-/-;K14cre;Brca1F/F;p53F/F model was generated in a similar fashion and described previously (26).

Weened pups were genotyped by tail DNA PCR and littermates were fed either phytoestrogen-free, genistein- or resveratrol-supplemented AIN-93G diets (27) from 6 weeks age onwards. Subsequently, animals were monitored three times per week and sacrificed once a mammary tumor of 10 mm in diameter or a skin tumor of 5 mm in diameter was identified. Other reasons for sacrifice included discomfort due to weight loss (> 20%), dyspnoe, or apathy. In addition to registration of age at sacrifice and tumor types, tumor and other tissue samples were collected during necropsy as previously described (25). To study tumor development in the absence of endogenous estrogens, 6 week old K14cre;Brca1F/F;p53F/F females were ovariectomized and fed one of the 3 special diets until sacrifice. All experimental procedures were approved by the Animal Ethics Committee of the Netherlands Cancer Institute.

The genistein- and resveratrol-supplemented AIN-93G diets were formulated by mixing concentrated phytoestrogen stocks (Wuxi Gorunjie Technology Co., Ltd) to a final concentration of 300 mg per kg pelleted food (SDS Special Diet Services), packed into 5kg bags and stored at -20°C until fed to the animals. To ensure that nutritional requirements of the mice were met continuously, each food batch was used for maximally 6 months after thawing.
The latency in mammary tumor development is reduced in Abcg2−/− animals, but not in Abcb1a/b−/− animals

For experiments on topotecan resistance (25), we introduced Abcg2 null alleles (23) into the K14cre;Brca1F/F;p53F/F mouse model (19) on a mixed 129/Ola and FVB/N genetic background. These ABCG2-deficient females (Fig. 1A, red line) developed spontaneous mammary tumors with a 33 day shorter median latency (180 days, \( P < 0.001 \)) than ABCG2-proficient ones (Fig. 1A, black line, 213 days). To eliminate artefacts caused by the mixed genetic background, we back-crossed the K14cre;Brca1F/F;p53F/F and Abcg2−/−;K14cre;Brca1F/F;p53F/F models to the FVB/N background. In addition, we generated K14cre;Brca1F/F;p53F/F animals deficient for the Abcb1a and Abcb1b genes, which encode the mouse P-glycoprotein homologues (24). On the FVB/N background we found that the median tumor latency of the K14cre;Brca1F/F;p53F/F females shortened from 213 to 201 days. The median mammary tumor latency was still 17.5 days shorter (\( P < 0.001 \)) in ABCG2-deficient K14cre;Brca1F/F;p53F/F FVB/N animals (Fig. 1B, red line, 183 days) than in transporter wildtype littermates (Fig. 1B, black line, 200.5 days). This was not the case in the ABCB1-deficient K14cre;Brca1F/F;p53F/F FVB/N animals (Fig. 1B, green line, 194 days), however (\( P = 0.770 \)). In addition to mammary tumors, K14 promoter-driven Cre transgene expression also results in skin tumors, mainly squamous cell carcinomas and some hair follicle tumors (19). Compared with the base-line tumor model (Fig. 1C, top panel), no significantly altered tumor type distributions were observed in the transporter deficient animals (Fig. 1C, middle and bottom panels). The FVB/N background or the introduction of Abcb1a/b or Abcg2 null alleles did not alter the distribution of morphologic phenotypes that were described previously (19).

Since ABCG2 transports estrogen-sulfates (28), ABCG2 ablation may stimulate tumorigenesis indirectly through elevating endogenous estrogen levels in the mammary epithelium. A more detailed analysis of overall survival per tumor type revealed, however, that both mammary and skin tumor latency were reduced in the Abcg2−/− animals, indicating that the transporter effect was not mammary gland-specific (Fig. 1D).

Supplementing the diet with genistein or resveratrol does not restore mammary tumor development in ovariectomized K14cre;Brca1F/F;p53F/F females

To rigorously put any cancer-preventing effects of genistein or resveratrol to the test, we determined whether the shortened mammary tumor latency, observed in our ABCG2-deficient K14cre;Brca1F/F;p53F/F females, was reversed by supplementing these compounds to the diet. To exclude the possibility that the estrogen-like
structure of these phytoestrogens promotes mammary tumorigenesis, we first fed ovariectomized

*K14cre;Brca1^{f/f};p53^{f/f}* females either a phytoestrogen-free diet (AIN-93G) or a diet containing 300 mg genistein
or resveratrol per kg food (22, 29) (Fig. 2A+B), starting at the age of 6 weeks. It has previously been shown that
this dosage level yields plasma concentrations in mice which are also attainable in humans, not toxic when fed
continuously, and which resulted in measurable effects on health and survival (30). Van de Ven et al. (submitted
for publication) showed that ovariectomy does not result in mammary gland regression of *K14cre;Brca1^{f/f};p53^{f/f}*
mice. Instead, the frequency of pre-neoplastic lesions, such as mammary duct dilatation, hyperplasia or
carcinoma in situ, is clearly reduced. Eventually, ovariectomy results in near complete loss of mammary tumor
development and a shift to skin tumor development, which we also observed in this study (compare Fig. 1C and
Fig. 2B). The relevance of estrogen on the tumor spectrum was demonstrated by the full restoration of mammary
tumor formation when 5 or 15 week-old ovariectomized females received subcutaneous pellets that released
17β-estradiol at physiological plasma levels (van de Ven et al., submitted for publication). In contrast to the
results with 17β-estradiol, we did not observe restoration of mammary tumor development by genistein or
resveratrol, nor a reduction in overall survival (Fig. 2A+B). Compared with the non-ovariectomized
*K14cre;Brca1^{f/f};p53^{f/f}* females (black lines, Fig. 1B and Fig. 2A), there was a significant (*P*=0.001, Log-rank
test) reduction in overall survival of the ovariectomized *K14cre;Brca1^{f/f};p53^{f/f}* females on phytoestrogen-free
diet (blue line, Fig. 2A). Both genistein and resveratrol reversed this ovariectomy effect (respectively *P*=0.004
and 0.012, Log-rank test). However, ovariectomized animals mainly developed skin tumors and if only the rate of
these tumors was analysed (Supplementary Fig. S1), the resveratrol effect was no longer significant, while the
genistein effect still was (respectively *P*=0.115 and 0.037, Log-rank test). This suggested that there may be a
modest effect of genistein on preventing skin tumor formation in our model. In contrast, resveratrol may reduce
the incidence of other pathological findings. Regarding tumor type distributions (Fig. 2B), there was no effect of
the phytoestrogens on mammary tumor incidence. However, there was a modest decrease in the incidence of
degenerative disease or infection (genistein and resveratrol) and in the incidence of tumors, which did not
originate from epithelial skin or mammary gland cells (resveratrol), such as thymoma or salivary gland
adenocarcinoma (other tumors). When the relative incidences of these two death causes were compared
between diet groups, the differences appeared to be appreciable, but they did not reach statistical significance
(respectively *P*=0.10 and *P*=0.11, Fisher's exact test), presumably due to the low numbers.
Resveratrol and genistein do not change the tumor latency reduction in *Abcg2*<sup>−/−</sup>;*K14cre;Brca1<sup>F/F</sup>;*p53<sup>F/F</sup>* females

Since we found a mild benefit of resveratrol on the overall survival of ovariectomized animals, we tested whether the shortened latency of mammary tumors in the *Abcg2*<sup>−/−</sup>;*K14cre;Brca1<sup>F/F</sup>;*p53<sup>F/F</sup>* model (FVB/N) could be reversed by genistein or resveratrol. We have previously shown that lack of ABCG2 results in increased plasma levels of resveratrol and its metabolites in FVB/N mice (22). Hence, any antagonizing effects of this phytoestrogen on mammary tumor development should even be more pronounced in these ABCG2-deficient *K14cre;Brca1<sup>F/F</sup>;*p53<sup>F/F</sup>* mice. However, neither the supplementation with genistein (Fig. 3A, red line) nor with resveratrol (Fig. 3A, green line) increased the overall survival in ABCG2-deficient animals compared with their control diet litter mates (Fig. 3A, blue line). The tumor type distributions were not significantly influenced by the diet either (data not shown). This lack of dietary effects was also observed in the ABCG2-proficient animals (Fig. 3B), showing that diets with high concentrations of genistein or resveratrol do not retard mammary tumorigenesis in our mouse model.
Discussion

We show here that ablation of the ABC transporter ABCG2 (BCRP), but not ABCB1 (P-gp/MDR1), shortens mammary tumor latency in the \textit{K14\textasciitilde{}cre;Brca1\textasciitilde{}F/F;\textasciitilde{}p53\textasciitilde{}F/F} mouse model for hereditary breast cancer. This effect was not influenced by genistein or resveratrol, supplemented at the high dose of 300 mg/kg in the diet.

Following Cre-mediated deletion of floxed \textit{Brca1} and \textit{p53} alleles, mammary epithelial cells become genomically unstable in our conditional mouse model and accumulate additional (epi)genetic alterations. This stochastic process requires a long latency period for cells to form clonal outgrowths that become full-blown mammary tumors (19). Although tumors from different individual animals show identical histomorphology, tumor-specific signatures are found at the molecular level (19, 25, 31). What role xenobiotics play in this tumor formation process is not clear. \textit{Abcg2\textasciitilde{}} mice are not tumor-prone. However, lack of ABCG2 reduces the time required for mammary tumor development caused by loss of BRCA1 and p53. ABCG2 actively eliminates endogenous metabolites or dietary xenobiotics from the body, and increased systemic exposure to a specific compound in \textit{Abcg2\textasciitilde{}} animals may enhance the mutagenesis of BRCA1;p53-deficient epithelial cells. This compound might be an ABCG2-specific substrate, since ablation of ABCB1 did not alter latency. We do not know the compound(s) and mechanism that cause the observed effects. An unbiased, mass spectrometry-based approach (32) might be able to detect the relevant metabolite differences between the plasma or tumor tissues derived from \textit{Abcg2\textasciitilde{}} versus \textit{Abcg2\textasciitilde{}+\textasciitilde{}} animals. Moreover, it may be interesting to investigate in future experiments whether a shorter tumor latency is also found in response to ABCG2-specific inhibitors.

Our data suggest that functional ABCG2 polymorphisms might influence tumorigenesis in women with a familiar breast cancer predisposition. Of the ABCG2 single nucleotide polymorphisms (SNPs) identified, only a few influence drug transport efficiency and therefore phamacokinetics-related side effects in patients (33). The common SNP rs2231142 (421 C>A), which encodes a Q141K loss of function mutation, causes at least 10\% of all gout cases in the Caucasian American population of the US (34). To our knowledge, no reports have yet been published linking ABCG2 loss-of-function SNPs to accelerated tumor progression in human BRCA1 or BRCA2 mutation carriers. Our mouse data suggest that ABCG2 dysfunction might increase the already high tumor incidence in the human population even further. Whether this effect is mammary gland-specific or also relevant to other cancer predispositions warrants further investigation.

We did not find an effect of genistein or resveratrol on mammary tumor development in our model. In other models the exposure of female Sprague-Dawley rats to 500 ppm genistein for 2 years significantly increased the risk of mammary adenocarcinoma development, but decreased the number of benign mammary fibroadenomas.
Overall, the evidence of carcinogenic activity of genistein in female rats was determined as being "equivocal". It was also found in rats that perinatal exposure to genistein renders animals more resistant to DMBA-induced mammary tumorigenesis (36, 37), whereas no effect was seen when genistein was given to adult animals (38). These differential effects appear to be caused by genistein-induced alterations of mammary gland development (39). In nude mice, physiologically attainable concentrations of genistein stimulate the growth of ER-positive mammary tumor xenotransplants, such as MCF-7 cells (40). Ju et al. (41) also reported that genistein stimulates tumor outgrowth of mice that have low estradiol levels due to ovariectomy, and they conclude that "products containing genistein may not be safe for postmenopausal women with estrogen-dependent breast cancer". It is possible that this effect of genistein is specific for already existing ER-positive tumor cells. The BRCA1; p53-deficient tumors derived from our model are ER-negative, like their human counterparts (19). Nevertheless, their development is clearly hormone-dependent. As shown in Fig. 2B ovariectomy results in nearly complete absence of mammary tumor development, which can be reversed by 17β-estradiol but not progesterone application (van de Ven et al., submitted for publication). Hence, mammary tumor formation in our model is ER-dependent. In BRCA1 mutation carriers, ovariectomy is also an effective prophylactic strategy to prevent tumorigenesis (42, 43). Our model may be useful to investigate whether aromatase inhibitors or selective estrogen receptor modulators have a similar effect as ovariectomy. The precise mechanisms responsible for this preventive effect are still unclear. It has been suggested that BRCA1-associated cancers arise from luminal progenitor cells which are still ER-positive (44). Another hypothesis is that paracrine RANKL signaling through ER-positive luminal mammary cells drives mammary stem cell expansion, which gives rise to tumors if additional genetic alterations during progression are acquired (45). Similarly, paracrine signaling through stromal cells has been shown to be critical during normal mammary gland development (46) and proposed as a mechanism for tissue-specificity of tumor development in BRCA1 patients (47). While BRCA1-deficient cells usually undergo apoptosis, survival in the mammary gland may be promoted through supportive signals from an estrogen-responsive stroma (48). Our data show that even high concentrations of genistein and resveratrol in the diet do not have the same effect as 17β-estradiol. Most likely the binding affinity of these compounds to the estrogen receptor is not sufficient to trigger any mammary tumor development (14, 49, 50).

In addition to a shift from mammary to skin tumor development, we observed that ovariectomy of K14cre; Brca1F/F; p53F/F animals resulted in a significant (P=0.001) reduction of overall survival (Fig. 2A). We do not know why. There were some ovariectomized animals on the phytoestrogen-free diet that died because of degenerative or infectious causes, whereas no such cases were present in the non-ovariectomized animals. It is
possible that the lack of endogenous estrogens increased the background pathology in our mouse model.

Whether the animals on the pytoestrogen-supplemented diets experienced a health benefit in addition to the modest delay in skin tumor development warrants further investigation.

In summary, we found that in a genetically engineered mouse model for BRCA1-associated breast cancer ABCG2, but not ABCB1, ablation significantly reduced mammary tumor latency. This reduction was not affected by dietary supplementation with genistein or resveratrol.
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Figure 1. Overall survival, cause of death and tumor latency of ABC transporter-proficient or -deficient K14cre;Brca1F/F;p53F/F females, fed with a phytoestrogen-free control diet. A, Kaplan-Meier (K-M) curves indicating overall survival (%) of K14cre;Brca1F/F;p53F/F and Abcg2-/-;K14cre;Brca1F/F;p53F/F females on a mixed Ola/129 – FVB/N background. Compared with the base-line model (black line, N=45), median mammary tumor latency was significantly shorter (P<0.001) in the ABCG2-deficient animals (red line, N=36). B, K-M curves indicating overall survival (%) of K14cre;Brca1F/F;p53F/F, Abcg2-/-;K14cre;Brca1F/F;p53F/F and Abcb1a/b-/-;K14cre;Brca1F/F;p53F/F females on a pure FVB/N background. Compared with the base-line model (black line, N=30), median mammary tumor latency was significantly shorter (P<0.001) in the ABCG2-deficient animals (red line, N=33), but not in the ABCB1-deficient animals (green line, N=32, P=0.770). C, Pie charts indicating tumor type distributions of K14cre;Brca1F/F;p53F/F, Abcg2-/-;K14cre;Brca1F/F;p53F/F and Abcb1a/b-/-;K14cre;Brca1F/F;p53F/F females on a pure FVB/N background. For each mouse strain the number (percentage) of animals killed due to skin tumors (dark blue), mammary tumors (brown), other tumors (yellow) or alternative pathology (light blue) is indicated. D, Boxplots of median mammary (MT) and skin tumor (ST) latencies in K14cre;Brca1F/F;p53F/F females (wildtype), Abcg2-/-;K14cre;Brca1F/F;p53F/F (ABCG2) or Abcb1a/b-/-;K14cre;Brca1F/F;p53F/F (ABCB1) animals. P values in A and B were calculated using the Log-rank test, whereas P values in D were calculated using the Wilcoxon signed rank test.

Figure 2. Overall survival and cause of death of ovariectomized K14cre;Brca1F/F;p53F/F females, fed with three special diets. A, K-M curves indicating overall survival (%). As a reference the K-M curve of non-ovariectomized K14cre;Brca1F/F;p53F/F females (black line, N=30) of Fig. 1B was added, which was significantly (P=0.001) different from the K-M curve of the ovariectomized animals on phytoestrogen-free control diet (blue line, N=32). Compared with these latter animals, median tumor latency was delayed in animals on genistein- (red line, N=34, P=0.012) and resveratrol- (green line, N=34, P=0.004) supplemented (300 mg/kg) test diets. B, Pie charts indicating tumor type distributions. For each diet the number (percentage) of animals killed due to skin tumors (dark blue), mammary tumors (brown), other tumors (yellow) or alternative pathology (light blue) is indicated. When death causes were compared between diet groups, no statistical significant differences could be detected. Even the categories degenerative/infection or other tumors, for which there seemed to be an appreciable effect in the pie charts, no significance was reached (respectively P=0.10 and P=0.11, Fisher’s exact test). The K-M curve P values were calculated using the Log-rank test.
Figure 3. Overall survival of non-ovariectomized Abcg2<sup>+/+</sup>;K14cre;Brca1<sup>F/F</sup>;p53<sup>F/F</sup> and Abcg2<sup>−/−</sup>;K14cre;Brca1<sup>F/F</sup>;p53<sup>F/F</sup> females, fed with three special diets. A, K-M curves indicating overall survival (% of Abcg2<sup>+/+</sup>;K14cre;Brca1<sup>F/F</sup>;p53<sup>F/F</sup> females on phytoestrogen-free control diet (blue line, N=33) or genistein- (red line, N=31) and resveratrol- (green line, N=34) supplemented (300 mg/kg) test diets. B, K-M curves indicating overall survival (% of Abcg2<sup>−/−</sup>;K14cre;Brca1<sup>F/F</sup>;p53<sup>F/F</sup> females on phytoestrogen-free control diet (blue line, N=30) or genistein- (red line, N=31) and resveratrol- (green line, N=26) supplemented (300 mg/kg) test diets.
Figure 1

A. 

Ola/129 – FVB/N

overall survival (%)

K14cre;Brca1F/F;p53F/F (N=45)
Abcg2–/-;K14cre;Brca1F/F;p53F/F (N=36)

B.

FVB/N

overall survival (%)

K14cre;Brca1F/F;p53F/F (N=30)
Abcg2–/-;K14cre;Brca1F/F;p53F/F (N=33)
Abcb1a/b–/-;K14cre;Brca1F/F;p53F/F (N=32)

C. cause of death:

- skin tumors
- mammary tumors
- other tumors
- degenerative/infection

K14cre;Brca1F/F;p53F/F
1 (3%) 0 (0%) 7 (23%)
22 (72%)

Abcg2–/-;K14cre;Brca1F/F;p53F/F
4 (12%) 1 (3%) 8 (24%)
20 (61%)

Abcb1a/b–/-;K14cre;Brca1F/F;p53F/F
3 (9%) 1 (3%) 7 (22%)
21 (66%)

D.

P = 0.6141 (NS)

P = 0.0283 (*)

P = 0.0479 (*)

P = 0.7266 (NS)

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**Figure 2**

(A) Kaplan-Meier cumulative survival curves for three treatment groups. There were no significant differences in survival between genistein and control diets (log-rank test, p=0.94); however, the resveratrol diet was associated with significantly better survival compared to the control diet (log-rank test, p=0.002).

(B) Pie charts showing the cause of death for each treatment group. In the control diet group, 5 rats (16%) died from degenerative/infection causes, 3 from skin tumors (9%), and 1 from mammary tumors (3%). In the genistein diet group, 3 rats (9%) died from skin tumors, 1 from mammary tumors, and 1 from other tumors (3%). In the resveratrol diet group, 0 rats died from skin tumors, 1 from mammary tumors, 1 from other tumors, and 1 from degenerative/infection causes (3%).
Figure 3

A

control diet (N=33)  genistein diet (N=31)  resveratrol diet (N=34)

overall survival (%)

0  50  100  150  200  250  300  350
days

Abcg2\textsuperscript{−/−};K14cre;Brca1\textsuperscript{F/F};p53\textsuperscript{F/F}

B

control diet (N=30)  genistein diet (N=31)  resveratrol diet (N=26)

overall survival (%)

0  50  100  150  200  250  300  350
days

Abcg2\textsuperscript{+/+};K14cre;Brca1\textsuperscript{F/F};p53\textsuperscript{F/F}

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