

# Crown-like Structures in Breast Adipose Tissue from Normal Weight Women: Important Impact

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## Abstract

Crown-like structures (CLS), composed of macrophages surrounding dead or dying adipocytes, are a histologic hallmark of the proinflammatory process by which adipose tissue contributes to the increased risk and worse prognosis of breast cancer in obese, postmenopausal patients. In this issue of *Cancer Prevention Research*, Iyengar and colleagues report the intriguing finding that CLS can be identified in a significant proportion of normal-BMI women undergoing mastectomy for breast cancer risk reduction or therapy. This surprising observation suggests that some normal weight women may

have similar mechanisms driving initiation and/or progression of breast cancer as those contributing to the increased incidence and worse prognosis of breast cancer in obese postmenopausal women. The possibility of a common mechanism in both lean and obese women provides added impetus to more fully define this process and evaluate its important implications for prevention and screening strategies as well as therapeutic interventions. *Cancer Prev Res*; 10(4); 1–3. ©2017 AACR.

See related article by Iyengar et al., p. 235–43

Articles by Iyengar et al. published in this issue *Cancer Prev Res* (1) as well as in 2015 (2) provide important insight into the potential mechanism(s) by which adipose tissue promotes cancer, especially in the breast, in both obese and some normal weight women. Many malignancies, including postmenopausal breast cancer, are epidemiologically and etiologically associated with obesity, which has been linked to all phases of carcinogenesis, including initiation, promotion, and progression (3, 4). The linkage between obesity and cancer has been attributed to multiple interacting factors, including changes in physical structure, altered adipokine secretion, increased secretion of growth factors, shifts in microbiome composition, dysregulation of hormone balance, and increased local and circulating inflammatory factors (4). Adipose tissue has long been recognized as an energy storage depot that accumulates or releases triglycerides, respectively, in response to energy excess or needs (5). It also is an active metabolic organ, communicating and regulating many physiologic and metabolic processes (appetite, hunger, satiety) through synthesis and secretion of a host of adipokines, signaling protein molecules synthesized primarily in adipocytes (5, 6). Among the multitude of these adipokines is leptin, whose synthesis is upregulated in obesity (6). Under physiologic conditions, leptin serves an anorexigenic function. It also functions to stimulate proinflammatory pathways in monocytes and macrophages and has been shown to promote tumor growth in mammary tumor systems (7). In contrast, synthesis and secretion of adiponectin, an anti-inflammatory and proapoptotic adipokine, is often downregulated in obesity (6). Adipose tissue serves also as the source for

localized and circulating factors characterizing a state of chronic low-grade inflammation associated with obesity (5, 6, 8). This metabolic inflammation (metaflammation) has been attributed to areas of adipose tissue where infiltrating bone marrow–derived macrophages (5, 9), showing a proinflammatory phenotype, cluster around the rim of enlarged, hypertrophied, dead and dying adipocytes, to form crown-like structures (CLS; refs. 8, 10–12). CLSs show greater frequency in adipose tissue from obese compared with lean individuals. They have been identified in most adipose deposits and show greater frequency in visceral compared with subcutaneous adipose tissue. These adipose tissue macrophages (ATM) in CLS phagocytose cellular debris, lipid droplets, and release triglycerides and free fatty acids (13, 14), products that can lead to insulin resistance and hyperinsulinemia (11–13, 15). Proinflammatory ATMs generate reactive oxygen species and reactive nitrogen species (5), both of which are potential mutagens. CLSs are associated with NF- $\kappa$ B activation and secretion of multiple inflammatory factors, including TNF $\alpha$ , IL1 $\beta$ , IL6, and COX-2–derived prostaglandin E2 (5, 6, 16). A variety of additional important immune abnormalities occur in inflamed adipose tissue (5). Adipose tissue is responsible for synthesis of aromatase, the rate-limiting enzyme that converts androgens to estrogens, and aromatase expression and enzymatic activity are increased in inflamed breast white adipose tissue (WAT; ref. 17). Importantly, an increase in the estrone:androstenedione ratio was found in breast tissue from postmenopausal breast cancer patients that contained CLS (18). Interestingly, inflamed adipose tissue has recently been shown to release cell-free DNA capable of stimulating insulin resistance (19). In addition, adipose tissue has been shown to be a source of circulating exosomal miRNA with a capacity for regulating glucose tolerance (20). These recent studies indicate that it will be important to investigate the role of circulating nucleic acids in contributing to the carcinogenic impact of inflamed WAT.

The Dannenberg laboratory has demonstrated the presence of CLS in WAT of the breast (CLS-B) in women with and without breast cancer, as well as in the noninvolved breast in women with

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breast cancer (16). CLS number and density increased with body mass index (BMI) and increased adipocyte size. CLS-B were more prevalent in post- compared with premenopausal women, and within each group, this difference was proportional to BMI. Thus, the highest percentage of patients positive for CLS-B occurred in postmenopausal women with BMI greater than 25 kg/m<sup>2</sup> (2, 21). They showed that the presence of CLS-B was associated with features of the metabolic syndrome. The presence of CLS-B was also associated with insulin resistance and changes in levels of circulating triglycerides, and increased proinflammatory factors, including leptin, hs-CRP, and IL6. These studies provide strong support for the premise that both obesity and postmenopausal status contribute to the development of CLS-B. As CLS-B occur both in the absence and presence of breast cancer and in the uninvolved breast in patients with cancer, these studies collectively suggest that WAT inflammation manifested as CLS-B may be causally linked to the increased incidence of hormone receptor-positive postmenopausal breast cancer in obese women. Interestingly, it has recently been reported that breast WAT from Black women, who have worse prognosis with breast cancer, have higher density of CLS-B than their non-Black counterparts (22). The probability that WAT inflammation helps to explain the obesity-inflammation-carcinogenesis axis is further supported by observations showing the occurrence of CLS in adipose tissue associated with other malignancies, including oral squamous cell carcinoma of the tongue (23), endometrial cancer (24), and in the report of liver macrophages, identified as Kupffer cells, forming CLS with lipid-laden hepatocytes in NAFLD, a disorder now considered to be a precursor for hepatocellular cancer (14, 25).

In this report by Iyengar and colleagues, in which CLS-B were found in women with normal BMI, there were, nonetheless, hypertrophied adipocytes, elevated levels of aromatase, increased circulating levels of proinflammatory factors, including hsCRP and leptin as well as elevated insulin and dyslipidemia, the latter contributing to a diagnosis of metabolic syndrome (1). These patients constitute a group of individuals with characteristics of metabolic syndrome with normal BMI now recognized as the metabolic equivalent of obesity. In addition to the likelihood that these normal-BMI women are at increased risk for breast cancer, occult WAT inflammation, a systemic process, is likely to place them at elevated risk for diabetes mellitus and cardiovascular disease. The practical implication of this observation is that these women deserve special attention regarding screening and risk reduction strategies. Successful

strategies to noninvasively identify these women, if coupled with interventions that attenuate WAT inflammation, could lead to a multitude of clinical benefits, including reducing the risk of breast cancer. Moreover, the findings of Iyengar and colleagues raise the question of what stimulates the carcinogenic process. Is it a consequence of obesity or do the causes of obesity independently promote the carcinogenic process? In response to this intriguing question, it is interesting to note that multiple models have now been reported wherein transplanted and/or spontaneous tumors, including breast cancer, are promoted by high-fat diet in the absence of obesity (26–30).

In a different tumor system, APC<sup>Min/+</sup> mice, we have recently shown that a high-fat coconut oil diet promotes obesity, inflammation, and aggressive tumor growth in some mouse strains, whereas in other strains, the same high-fat diet promotes inflammation and tumor growth independent of obesity (30). We showed, moreover, that an isocaloric high-fat, corn oil diet promoted inflammation and tumor growth in the absence of obesity, whereas in contrast, a high-fat olive oil diet promoted obesity but not inflammation or tumor growth. Thus, depending on the quality and quantity of dietary fat and its interaction with genetic background, it is clear that all combinations of obesity, inflammation, and carcinogenesis can be stimulated. Given these findings, it would be of considerable interest to determine whether the type and/or amount of dietary fat impacts the incidence of breast WAT inflammation reported by Iyengar and colleagues in this issue of *Cancer Prevention Research* (1). In addition, the human counterpart of our study may explain differential beneficial versus carcinogenic effects of the Mediterranean versus the Western diet relative to obesity, inflammation, and cancer (Di Daniele and colleagues 2017; ref. 31). The important issue that these studies raise regarding primary and secondary cancer prevention is whether it is more important to control the quality and/or quantity of dietary fat. The public health implications are significant.

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No potential conflicts of interest were disclosed.

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### References

- Iyengar NM, Brown KA, Zhou XK, Gucalp A, Subbaramaiah K, Giri DD, et al. Metabolic obesity, adipose inflammation and elevated breast aromatase in women with normal body mass index. *Cancer Prev Res* 2017;10:235–43.
- Iyengar NM, Morris PG, Zhou XK, Gucalp A, Giri D, Harbus MD, et al. Menopause is a determinant of breast adipose inflammation. *Cancer Prev Res* 2015;8:349–58.
- Calle EE, Kaaks R. Overweight, obesity and cancer: epidemiological evidence and proposed mechanisms. *Nature Rev Cancer* 2004;4: 579–91.
- Berger NA. Obesity and cancer pathogenesis. *Ann NY Acad Sci* 2014;1311: 57–76.
- Hotamisligil GS. Inflammation, metaflammation and immunometabolic disorders. *Nature* 2017;542:177–185.
- Ouchi N, Parker JL, Lugus JJ, Walsh K. Adipokines in inflammation and metabolic disease. *Nat Rev Immunol* 2011;11:85–97.
- Zheng Q, Dunlap SM, Zhu J, Downs-Kelly E, Rich J, Hursting SD, et al. Leptin deficiency suppresses MMTV-Wnt-1 mammary tumor growth in obese mice and abrogates tumor initiating cell survival. *Endocr Relat Cancer* 2011;18:491–503.
- Kang YE, Kim JM, Joung KH, Lee JH, You BR, Choi MJ, et al. The roles of adipokines, proinflammatory cytokines, and adipose tissue macrophages in obesity-associated insulin resistance in modest obesity and early metabolic dysfunction. *PLoS One* 2016;11:e0154003.
- Lumeng CN, DelProposto JB, Westcott DJ, Saltiel AR. Phenotypic switching of adipose tissue macrophages with obesity is generated by spatiotemporal differences in macrophage subtypes. *Diabetes* 2008; 57:3239–46.

10. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AWJr. Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest* 2003;112:1796–808.
11. Xu H, Barnes GT, Yang Q, Tan G, Yang D, Chou CJ, et al. Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J Clin Invest* 2003;112:1821–30.
12. Murano I, Barbatelli G, Parisani V, Latini C, Muzzonigro G, Castellucci M, et al. Dead adipocytes, detected as crown-like structures, are prevalent in visceral fat depots of genetically obese mice. *J Lipid Res* 2008;49:1562–8.
13. Glass CK, Olefsky JM. Inflammation and lipid signaling in the etiology of insulin resistance. *Cell Metab* 2012;15:635–45.
14. Jung UJ, Choi MS. Obesity and its metabolic complications: the role of adipokines and the relationship between obesity, inflammation, insulin resistance, dyslipidemia and nonalcoholic fatty liver disease. *Int J Mol Sci* 2014;15:6184–223.
15. Suganami T, Nishida J, Ogawa Y. A paracrine loop between adipocytes and macrophages aggravates inflammatory changes: role of free fatty acids and tumor necrosis factor alpha. *Arterioscler Thromb Vasc Biol* 2005;25:2062–8.
16. Iyengar NM, Gucalp A, Dannenberg AJ, Hudis CA. Obesity and Cancer Mechanisms: Tumor Microenvironment and Inflammation. *J Clin Oncol* 2016;34:4270–4276.
17. Morris PG, Hudis CA, Giri D, Morrow M, Falcone DJ, Zhou XK, et al. Inflammation and increased aromatase expression occur in the breast tissue of obese women with breast cancer. *Cancer Prev Res* 2011;4:1021–9.
18. Mullooly M, Yang HP, Falk RT, Nyante SJ, Cora R, Pfeiffer RM, et al. Relationship between crown-like structures and sex-steroid hormones in breast adipose tissue and serum among postmenopausal breast cancer patients. *Breast Cancer Res* 2017;19:8.
19. Nishimoto S, Fukuda D, Higashikuni Y, Tanaka K, Hirata Y, Murata C, et al. Obesity-induced DNA released from adipocytes stimulates chronic adipose tissue inflammation and insulin resistance. *Sci Adv* 2016;2:e1501332.
20. Thomou T, Mori MA, Dreyfuss JM, Konishi M, Sakaguchi M, Wolfgram C, et al. Adipose-derived circulating miRNAs regulate gene expression in other tissues. *Nature* 2017;542:450–5.
21. Iyengar NM, Zhou XK, Gucalp A, Morris PG, Howe LR, Giri DD, et al. Systemic correlates of white adipose tissue inflammation in early-stage breast cancer. *Clin Cancer Res* 2016;22:2283–9.
22. Koru-Sengul T, Santander AM, Miao F, Sanchez LG, Jorda M, Gluck S, et al. Breast cancers from black women exhibit higher numbers of immunosuppressive macrophages with proliferative activity and of crown-like structures associated with lower survival compared to non-black Latinas and Caucasians. *Breast Cancer Res Treat* 2016;158:113–26.
23. Iyengar NM, Ghossein RA, Morris LG, Zhou XK, Kochhar A, Morris PG, et al. White adipose tissue inflammation and cancer-specific survival in patients with squamous cell carcinoma of the oral tongue. *Cancer* 2016;122:3794–802.
24. Berstein LM, Iyevleva AG, Mukhina MS, Vasilyev DA, Poroshina TE. Features of omental adipose tissue in endometrial cancer patients with 'standard' or 'metabolically healthy' obesity: associations with tumor process characteristics. *Springerplus* 2016;5:1900.
25. Itoh M, Kato H, Suganami T, Konuma K, Marumoto Y, Terai S, et al. Hepatic crown-like structure: a unique histological feature in non-alcoholic steatohepatitis in mice and humans. *PLoS One* 2013;8:e82163.
26. Kim EJ, Choi MR, Park H, Kim M, Hong JE, Lee JY, et al. Dietary fat increases solid tumor growth and metastasis of 4T1 murine mammary carcinoma cells and mortality in obesity-resistant BALB/c mice. *Breast Cancer Res* 2011;13:R78.
27. Tang FY, Pai MH, Chiang EP. Consumption of high-fat diet induces tumor progression and epithelial-mesenchymal transition of colorectal cancer in a mouse xenograft model. *J Nutr Biochem* 2012;23:1302–13.
28. Park H, Kim M, Kwon GT, Lim DY, Yu R, Sung MK, et al. A high-fat diet increases angiogenesis, solid tumor growth, and lung metastasis of CT26 colon cancer cells in obesity-resistant BALB/c mice. *Mol Carcinog* 2012;51:869–80.
29. Lamas B, Nachat-Kappes R, Goncalves-Mendes N, Mishellany F, Rossary A, Vasson MP, et al. Dietary fat without body weight gain increases in vivo MCF-7 human breast cancer cell growth and decreases natural killer cell cytotoxicity. *Mol Carcinog* 2015;54:58–71.
30. Doerner SK, Reis ES, Leung ES, Ko JS, Heaney JD, Berger NA, et al. High-fat diet-induced complement activation mediates intestinal inflammation and neoplasia, independent of obesity. *Mol Cancer Res* 2016;14:953–65.
31. Di Daniele N, Noce A, Vidiri MF, Moriconi E, Marrone G, Annicchiarico-Petruzzelli M, et al. Impact of Mediterranean diet on metabolic syndrome, cancer and longevity. *Oncotarget* 2017;8:8947–79.

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