

## Kava, a Tonic for Relieving the Irrational Development of Natural Preventive Agents

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The last two decades have witnessed explosive growth in the study of natural and other cancer chemopreventive agents (1, 2). Extensive preclinical data have been generated for natural agents, and many (such as green tea, curcumin, phenyl isothiocyanate, indole-3-carbinol, silibinin, lycopene, genistein, selenium, and vitamins A, E, and D) are currently in different phases of clinical testing (3). The definitive clinical prevention trials of natural agents completed thus far have been largely negative, suggesting that detailed mechanistic and efficacy studies are necessary to supplement the epidemiologic data before clinically testing novel natural compounds (3). Therefore, investigators increasingly are studying the mechanisms of cancer chemopreventive agents (natural or synthetic, including molecular targeted) to substantiate their potential efficacy.

A substantial body of work showing a broad spectrum of natural-agent mechanisms has raised important issues. For example, what is a relevant, achievable dose *in vivo* for targeting relevant pathways or targets (versus the potentially high, unachievable doses studied *in vitro*)? Which of the multiple mechanisms are potential causes of toxicity? A multiplicity of mechanisms certainly could enhance natural agent effects, but it is important also to try to identify specific relevant or predominant mechanism(s) that are critical for preventive activity in specific carcinogenic systems. Besides giving scientific credibility, mechanistic insight will facilitate clinical development by elucidating key pathway(s) and target(s) to be monitored in dose-finding early-phase clinical trials (4). It also helps in selecting patient populations based on appropriate prevention settings and potential sensitivity to the preventive and/or toxic effects of the agent. Understanding relevant mechanisms also helps in developing more specifically targeted analogues with potentially less toxicity, greater preventive activity, and less variability in formulation, which is important for assuring the desired effects of an intervention.

The substantial data reported thus far on mechanisms of chemopreventive agents are just the tip of an iceberg of mostly unknown mechanisms involving about 20,000 protein-coding human genes and the epigenetic machinery that contributes to the regulation of gene expression. Most advances in understanding the mechanisms of natural agent actions have been

confined, until recently, to cell culture studies. Now, however, various animal models (e.g., knockout, knock-in, or transgenic mice) are frequently used to establish the *in vivo* mechanistic aspects of natural agents. Furthermore, the use of “omic” approaches (genomic, proteomic, metabolomic, etc.) in chemoprevention has helped in speedily measuring altered expression of thousands of genes in response to phytochemicals (plant-derived chemical compounds) and promises to further crystallize natural-agent mechanisms.

Silibinin, a constituent of milk thistle, can help in illustrating current mechanistic study of natural agent mechanisms. Milk thistle extracts have been used for centuries as a medicament for hepatobiliary diseases and during the last few decades in clinical testing for treating acute mushroom poisoning, hepatic cirrhosis, and acute viral hepatitis (5, 6). Milk thistle extract, also labeled silymarin or silibinin, is now marketed as a nutritional supplement to promote healthy liver function (5). In the 1970s, we reported the first evidence of the cancer preventive activity of silymarin/silibinin in a series of *in vivo* studies using mouse skin cancer models (7, 8). Studies of the last 15 years in different *in vitro* and *in vivo* models have established the mechanistic details of silibinin cancer preventive effects in various epithelial cancers, including prostate, lung, bladder, colorectal, kidney, oral, skin, renal, breast, ovarian, and tongue cancers (6, 7, 9, 10). These mechanistic studies showed that silibinin treatment inhibits unchecked cellular proliferation in cancer cells by targeting the cell cycle through modulation of various cell cycle regulators (11). This activity includes strongly inhibiting constitutive as well as growth factor-induced receptor tyrosine signaling and inhibiting androgen receptor and signal transducer and activator of transcription signaling (9). The growth of cancer cells is almost always accompanied by the loss of apoptotic response, and silibinin treatment has been shown to induce apoptosis in many cancer cell lines and in tumor tissues via modulation of expression of various Bcl2 and inhibitor of apoptosis family members through inhibition of nuclear factor  $\kappa$ B and with or without the activation of various caspases (7, 9). Silibinin treatment has been shown to target the expression of various proangiogenic factors (e.g., vascular endothelial growth factor, basic fibroblast growth factor, and inducible nitric oxide synthase), thereby affording strong antiangiogenic efficacy (7, 9). Overall, these studies suggest pleiotropic mechanisms for silibinin anticancer activity; more recent studies, however, showed that inhibition of epidermal growth factor receptor activation is necessary and sufficient for the anticancer effects of silibinin (12). This places silibinin in the class of receptor tyrosine kinase inhibitors, which have undergone extensive clinical testing for cancer control (13).

Another good example of a mechanistically evaluated natural agent with chemopreventive potential is the phytochemical deguelin. Deguelin has several relevant mechanisms

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including inhibition of Akt, a very prominent target for molecular-targeted drug development, *in vitro* and *in vivo* and blocking tobacco-induced lung carcinogenesis in A/J mice (14). *In vitro* study of deguelin in premalignant human bronchial epithelial cells seems to be the first work to illustrate the importance of Akt targeting in lung chemoprevention (14). In other words, mechanistic study of deguelin was used for target identification and stimulated tremendous interest in developing specific inhibitors of Akt and the phosphatidylinositol 3-kinase/Akt pathway for lung cancer prevention and therapy. More recently, deguelin has been shown to inhibit heat shock protein-90 function leading to degradation of its various client proteins including Akt and hypoxia-inducible factor 1 $\alpha$  (15). The caveat in developing deguelin as a cancer preventive drug, however, is that it can inhibit NADH:ubiquinone oxidoreductase activity (16), which could cause neuronal or other toxicity. Therefore, investigators are now developing deguelin analogues with greater specificity for Akt and thus greater potency in lung carcinogenesis and lesser potential toxicity. These analogues are being patented and thus have enhanced potential for developmental funding support from federal and industry sources. Mechanisms have been reported not only for silibinin and deguelin but also for several other well-studied natural agents including genistein, curcumin, apigenin, indole-3-carbinol, green tea, lycopene, grape seed extract, inositol hexaphosphate (or phytic acid), garlic, and cruciferous constituents.

All of the foregoing evidence supports a role for phytochemicals as cancer chemopreventive agents and warrants more vigorous work to identify, and preclinical testing of, novel natural agents with chemopreventive activity. Such preclinical work is reflected in the kava studies by Johnson et al. (17) and Tang et al. (18) reported in this issue of the journal. Discussed in detail below, these studies portray a rational sequence of natural agent development for prevention, with a proof of the preventive potential of kava extract *in vivo* in mice (17) and a more detailed mechanistic analysis of a specific kava constituent both *in vitro* and in a mouse xenograft model (18). Such work is a necessary precursor to clinical development of kava or any other natural agent, notwithstanding seemingly compelling epidemiologic and general biological evidence of its preventive potential.

Kava (*Piper methysticum*) is an ancient crop of the western Pacific islands, where it has been used as a medicine, social drink, and sacred plant in religious ceremonies (19). The traditional kava drink is prepared from the plant roots, and its consumption causes a mildly talkative and sociable behavior, clear thinking, and anxiolytic and muscle-relaxing effects (20). Kava extract consists mainly of two classes of compounds: kavalactones and flavokawainins (FK), or chalcones. Kavalactones are the major constituents (3–20%) of kava extract and mainly include methysticin, dihydromethysticin, 7,8-dihydrokawain, kawain, desmethoxyyangonin, yangonin, and 7,8-epoxyyangonin (20). Chalcones include FKA, FKB, and FKC and constitute less than 1% of total kava extract (20). Kava (20) attracted global (and mechanistic) attention in the 1990s as an herbal supplement for reducing anxiety, stress, and insomnia (21). Strong epidemiologic evidence suggests that kava-drinking populations have an unusually low cancer incidence despite high rates of smoking (22, 23). The age-standardized cancer incidences for Fiji, Va-

nuatu, and Western Samoa, the three countries with the highest consumption of kava drink, were reported to be one third or one fourth of the cancer incidences in non-kava-drinking countries (23). Furthermore, the cancer incidences in these populations were lower in men compared with women, despite much higher smoking rates among men, who also consume more kava (22–24). This collection of evidence suggests that the traditional herb kava would be useful in the prevention and/or treatment of smoking-related diseases such as lung and bladder cancers. Several previous studies also have examined kava mechanisms in various preclinical carcinogenesis systems (20, 25, 26).

As published in this issue of the journal, Johnson et al. (17) showed for the first time the *in vivo* cancer chemopreventive potential of kava against chemical carcinogen-induced lung tumorigenesis. Kava extract (10 mg/g mixed in food) significantly reduced 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone plus benzo(a)pyrene-induced lung tumor multiplicity in A/J mice. The kava regimen significantly reduced lung tumor multiplicity whether given during carcinogen treatment only, after carcinogen treatment only, or both during and after. These results suggest that kava might inhibit events of initiation as well as promotion associated with chemical carcinogenesis. Kava also reduced the proliferation marker proliferating cell nuclear antigen, increased markers of apoptosis [caspase-3 and poly(ADP-ribose) polymerase cleavage], and inhibited activation of nuclear factor  $\kappa$ B (17). Of note, this study also showed that kava extract given in food for 30 weeks at a dose of 10 mg/g does not cause hepatotoxicity, which was measured in terms of liver weight, liver pathology, and markers of liver damage (alanine aminotransferase, aspartate aminotransferase, and  $\gamma$ -glutamyltransferase).

Also in this issue of the journal, Tang et al. (18) published a report on kava and bladder carcinogenesis, which strongly complements the promising results of Johnson et al. in lung carcinogenesis. The bladder study follows up on work reported by the same group (26) showing that kava extract and FKA, FKB, and FKC strongly induced apoptosis in human bladder cancer cells via an increase in the active form of Bax protein and a decrease in the expression of X-linked inhibitor of apoptosis and survivin. This earlier study also showed that FKA treatment inhibited the anchorage-independent growth and *in vivo* xenograft growth of bladder cancer EJ cells (26). Tang et al. now show that the kava chalcone FKA (50 mg/kg of body weight) strongly decreased the *in vivo* growth rate of a bladder cancer xenograft (RT4 cells) in athymic nude mice, without causing toxicity. The studies of Tang et al. and Johnson et al. clearly and convincingly suggest the strong preventive and therapeutic potential of kava against the major tobacco-related diseases lung and bladder cancers.

Smoking exposure (among other factors) results in cancer initiation via a number of molecular changes including mutations that inactivate tumor suppressor genes [such as *p53*, retinoblastoma (*Rb*), and *INK4*] and/or mutations that activate various oncogenes [such as epidermal growth factor receptor (*EGFR*), *Ras*, and *cyclin D1*; ref. 27]. Through these changes, cancer cells acquire the capability of uncontrolled multiplication, apoptosis evasion, and constitutive activation of survival signaling pathways such as nuclear factor  $\kappa$ B and Akt, which

is followed by neoangiogenesis and metastatic spread (28). Johnson et al. (17) and Tang et al. (18) showed that kava or its constituent FKA could inhibit proliferation and nuclear factor- $\kappa$ B activation and induce apoptosis in tobacco-related lung and bladder cancer cells. The antiproliferative effect of FKA was more prominent in bladder cancer cell lines harboring mutations in both *p53* and *Rb*, which are frequent in tobacco-related human cancers (17, 18, 27).

Targeting the deregulated cell cycle has emerged as an ideal prevention strategy for checking the development and uncontrolled growth of cancer cells (29). Tang et al. (18) showed that FKA treatment differentially induced G<sub>1</sub> cell cycle arrest in *p53* wild-type and G<sub>2</sub>-M cell cycle arrest in *p53*-mutant bladder cancer cells. Furthermore, in low-grade bladder cancer cells carrying wild-type *p53*, FKA treatment increased the levels of cyclin-dependent kinase (CDK) inhibitors (p21 and p27) and decreased CDK2 kinase activity (18). However, in *p53*-mutant, high-grade bladder cancer cells, FKA treatment reduced the expression of CDK1 inhibitory kinases, Myt1 and Wee1, and increased cyclin B1 levels leading to CDK1 activation (18). These results suggest that FKA is potentially a G<sub>2</sub> checkpoint abrogator in cancer cells carrying mutant *p53*, which is consistent with the induction by FKA of M-phase arrest in cancer cells. Of interest, FKA induced M-phase arrest through signaling events downstream of widely known cellular checkpoints (Chk1 and Chk2). Whether this unscheduled entry into M-phase in response to FKA treatment leads to activation of spindle checkpoint and results in mitotic catastrophe remains to be examined. The effect of FKA treatment on the expression of key mitotic kinesins and kinases (Plk and Aurora kinases) must also be examined to understand the mechanistic details underlying M-phase arrest. FKA treatment also promoted mitotic slippage in bladder cancer cells, which needs to be studied more closely because mitotic slippage not only could lead to cell cycle arrest and apoptosis but also could promote genetic instability and cancer progression (30). All of the *in vitro* and *in vivo* evidence, along with the epidemiologic data from Pacific island populations, supports the cancer chemopreventive potential of kava and its constituents. Other issues, however, remain to be addressed. Little or no literature is available about the bioavailability of kava constituents in plasma and other organs of interest. Of particular importance to prevention, kava has

potential toxicity. The use of kava as an herbal supplement was banned by many Western countries in 2002 after reports of its severe hepatotoxicity. The numerous studies of kava toxicity (21, 25, 31–33) have shown, for example, that this toxicity is linked to kava formulation/extraction (acetone/ethanol extraction or extraction from the stem or leaves of the kava plant), to genetic background (e.g., CYP2D6 deficiency is prevalent in 7–9% of Europeans but is rare in Polynesians and Asians), and strongly to the interaction of kava with other drugs (21, 31–33).

The articles in this issue of the journal raise the question of whether it is better to screen and develop natural products for their cancer preventive or therapeutic activity or to take a targeted, mechanistic approach in developing specific inhibitors of known critical molecules in cancer cells. Either approach has the potential to identify effective cancer preventive or therapeutic agents. Many scientists or pharmaceutical houses, however, prefer one or the other of these strategies. The benefits of screening specific targets (e.g., performing a screen of all known kinases to develop specific kinase inhibitor drugs) are that (a) a target is known once a lead compound is selected, and (b) extensive mechanistic data on the particular target in relation to cancer cell growth may be available. However, if multiple signaling molecules are required to effectively prevent or treat cancer, this single-target screen may ultimately fail, and screening a natural compound with the potential to affect many signaling pathways at once may be more productive. The advantages, disadvantages, and technical issues involved in specific molecular-targeted versus natural-agent development would be a worthy topic for a future perspective or commentary.

In conclusion, although the ultimate success of kava will depend on the outcomes of further preclinical and clinical studies, this herb exemplifies the principle of “nature to bedside” and supports the identification and preclinical and clinical testing of natural agents for cancer chemoprevention. Kava presents as well a venue for examining the value of robust mechanistic studies in advancing rational natural-agent development.

## Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

## References

1. Tsao AS, Kim ES, Hong WK. Chemoprevention of cancer. *CA Cancer J Clin* 2004;54:150–80.
2. Naithani R, Huma LC, Moriarty RM, McCormick DL, Mehta RG. Comprehensive review of cancer chemopreventive agents evaluated in experimental carcinogenesis models and clinical trials. *Curr Med Chem* 2008;15:1044–71.
3. Thomasset SC, Berry DP, Garcea G, Marczylo T, Steward WP, Gescher AJ. Dietary polyphenolic phytochemicals-promising cancer chemopreventive agents in humans? A review of their clinical properties. *Int J Cancer* 2007;120:451–8.
4. Szabo E. Assessing efficacy in early-phase cancer prevention trials: the case of oral premalignancy. *Cancer Prev Res* 2008;1:312–5.
5. Wellington K, Jarvis B. Silymarin: a review of its clinical properties in the management of hepatic disorders. *BioDrugs* 2001;15:465–89.
6. Agarwal R, Agarwal C, Ichikawa H, Singh RP, Aggarwal BB. Anticancer potential of silymarin: from bench to bedside. *Anticancer Res* 2006;26:4457–98.
7. Deep G, Agarwal R. Chemopreventive efficacy of silymarin in skin and prostate cancer. *Integr Cancer Ther* 2007;6:130–45.
8. Lahiri-Chatterjee M, Katiyar SK, Mohan RR, Agarwal R. A flavonoid antioxidant, silymarin, affords exceptionally high protection against tumor promotion in the SENCAR mouse skin tumorigenesis model. *Cancer Res* 1999;59:622–32.
9. Singh RP, Agarwal R. Mechanisms of action of novel agents for prostate cancer chemoprevention. *Endocr Relat Cancer* 2006;13:751–78.
10. Velmurugan B, Singh RP, Tyagi A, Agarwal R. Inhibition of azoxymethane-induced colonic aberrant crypt foci formation by silibinin in male Fisher 344 rats. *Cancer Prev Res* 2008;1:376–84.
11. Agarwal R. Cell signaling and regulators of cell

- cycle as molecular targets for prostate cancer prevention by dietary agents. *Biochem Pharmacol* 2000;60:1051–9.
12. Qi L, Singh RP, Lu Y, et al. Epidermal growth factor receptor mediates silibinin-induced cytotoxicity in a rat glioma cell line. *Cancer Biol Ther* 2003;2:526–31.
13. Petrelli A, Giordano S. From single- to multi-target drugs in cancer therapy: when aspecificity becomes an advantage. *Curr Med Chem* 2008;15:422–32.
14. Lee HY, Oh SH, Woo JK, et al. Chemopreventive effects of deguelin, a novel Akt inhibitor, on tobacco-induced lung tumorigenesis. *J Natl Cancer Inst* 2005;97:1695–9.
15. Oh SH, Woo JK, Yazici YD, et al. Structural basis for depletion of heat shock protein 90 client proteins by deguelin. *J Natl Cancer Inst* 2007;99:949–61.
16. Fang N, Casida JE. Anticancer action of cube insecticide: correlation for rotenoid constituents between inhibition of NADH:ubiquinone oxidoreductase and induced ornithine decarboxylase activities. *Proc Natl Acad Sci U S A* 1998;95:3380–4.
17. Johnson TE, Kassie F, O'Sullivan MG, et al. Chemopreventive effect of kava on 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone plus benzo(a)pyrene-induced lung tumorigenesis in A/J mice. *Cancer Prev Res* 2008;1:430–8.
18. Tang Y, Simoneau AR, Xie J, Shahandeh B, Zi X. Effects of the kava chalcone flavokawain A differ in bladder cancer cells with wild-type versus mutant p53. *Cancer Prev Res* 2008;1:439–51.
19. O'Sullivan HM, Lum K. The poisoning of 'awa: the non-traditional use of an ancient remedy. *Pac Health Dialog* 2004;11:211–5.
20. Tabudravu JN, Jaspars M. Anticancer activities of constituents of kava (*Piper methysticum*). *The South Pac J Natural Sci* 2005;23:26–9.
21. Clouatre DL. Kava kava: examining new reports of toxicity. *Toxicol Lett* 2004;150:85–96.
22. Henderson BE, Kolonel LN, Dworsky R, et al. Cancer incidence in the islands of the Pacific. *Natl Cancer Inst Monogr* 1985;69:73–81.
23. Steiner GG. The correlation between cancer incidence and kava consumption. *Hawaii Med J* 2000;59:420–2.
24. Singh YN. Kava: an overview. *J Ethnopharmacol* 1992;37:13–45.
25. Weiss J, Sauer A, Frank A, Unger M. Extracts and kavalactones of *Piper methysticum* G. Forst (kava-kava) inhibit P-glycoprotein *in vitro*. *Drug Metab Dispos* 2005;33:1580–3.
26. Zi X, Simoneau AR, Flavokawain A. A novel chalcone from kava extract, induces apoptosis in bladder cancer cells by involvement of Bax protein-dependent and mitochondria-dependent apoptotic pathway and suppresses tumor growth in mice. *Cancer Res* 2005;65:3479–86.
27. Mitra AP, Birkhahn M, Cote RJ. p53 and retinoblastoma pathways in bladder cancer. *World J Urol* 2007;25:563–71.
28. Naugler WE, Karin M. NF- $\kappa$ B and cancer—identifying targets and mechanisms. *Curr Opin Genet Dev* 2008;18:19–26.
29. Deep G, Agarwal R. New combination therapies with cell-cycle agents. *Curr Opin Investig Drugs* 2008;9:591–604.
30. Dalton WB, Nandan MO, Moore RT, Yang VW. Human cancer cells commonly acquire DNA damage during mitotic arrest. *Cancer Res* 2007;67:11487–92.
31. Whittaker P, Clarke JJ, San RH, et al. Evaluation of commercial kava extracts and kavalactone standards for mutagenicity and toxicity using the mammalian cell gene mutation assay in L5178Y mouse lymphoma cells. *Food Chem Toxicol* 2008;46:168–74.
32. Whitton PA, Lau A, Salisbury A, Whitehouse J, Evans CS. Kava lactones and the kava-kava controversy. *Phytochemistry* 2003;64:673–9.
33. Moulds RF, Malani J. Kava: herbal panacea or liver poison?. *Med J Aust* 2003;178:451–3.

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