

Comparison of Systemic Exposure to Toxic and/or Carcinogenic Volatile Organic Compounds (VOC) during Vaping, Smoking, and Abstention

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ABSTRACT

Comparisons of systemic exposure to toxicants during monitored cigarette smoking, electronic cigarette (e-cigarette) use, and abstention are needed to enhance our understanding of the risks of e-cigarette use (vaping). In a crossover study, we measured 10 mercapturic acid metabolites of volatile organic compounds (VOCs) in 24-hour urine samples collected from 36 dual users (8 women) of e-cigarettes and cigarettes during 2 days of *ad libitum* vaping or cigarette-only use, and 2 days of enforced abstention. Concentrations of VOC metabolites were higher during smoking compared with vaping, except for the methylating agents' metabolite. The fold-difference in concentrations when smoking relative to vaping ranged from 1.31 (1.06–1.61; geometric mean, 95% confidence interval; 1,3-butadiene) to 7.09 (5.88–8.54;

acrylonitrile). Metabolites of acrylamide [fold difference of 1.21 (1.03–1.43)] and benzene [1.46 (1.13–1.90)] were higher during vaping compared with abstention. The 1,3-butadiene and propylene oxide metabolites were higher in variable-power tank users compared with users of cig-a-likes. E-cigarettes expose users to lower levels of toxic VOCs compared with cigarette smoking, supporting their harm reduction potential among smokers. However, some e-cigarettes expose users to VOCs such as acrylamide, benzene, and propylene oxide, and may pose health risks to non-smoking users. The results of our study will inform regulators in assessing e-cigarettes with respect to the balance between its potential harm reduction for adult smokers and risk to nonsmoking users.

Introduction

Although nicotine is the primary addictive substance in tobacco smoke (1), and despite concerns about nicotine's potential deleterious effects (2, 3), the morbidity and mortality of smoking are attributable primarily to non-nicotine toxicants such as volatile organic compounds (VOC; refs. 4, 5). As such, cigarettes and other combustible tobacco products are the most harmful on the continuum of risk of tobacco products. On the other hand, noncombustible tobacco products, such as electronic cigarettes (e-cigarettes), are believed to be less harmful because their emissions contain low levels or none of the many toxicants present in tobacco smoke (6). On the basis of these

observations, the harm reduction potential of e-cigarettes has been proposed by some, including the FDA (7), as a way to reduce the public health burden of smoking. A further important consideration is the inherent toxicity of e-cigarettes and the potential risks they pose to nonsmokers who vape.

In general, e-cigarette users have lower systemic exposure to toxicants compared with smokers, which supports the idea that e-cigarettes have a lower risk profile relative to cigarettes (6). These findings have been largely derived from studies in which e-cigarette-naïve smokers switched from cigarettes to e-cigarettes and were followed prospectively in their naturalistic settings (i.e., switching studies), and include assessing changes in biomarkers of tobacco-related toxicants (8–11). In addition, cross-sectional comparisons of biomarkers of toxicant exposure in smokers and e-cigarette users who used their products in their naturalistic settings have been reported (12–15). Important limitations of these studies are that product use (and patterns of use) are not controlled or monitored, are self-reported, and potentially include use of multiple tobacco products, making it difficult to ascertain e-cigarette use and quantify the magnitude of e-cigarette-associated exposures and the risk of e-cigarette-only use. More accurate assessment of toxicant exposure associated with e-cigarette-only use relative to cigarette smoking or no product use will enhance our understanding of the public health risk of e-cigarettes. One between-subject industry study found no significant differences in levels of VOC metabolites in smokers

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who used blu e-cigarettes exclusively for 5 days in a research setting compared with smokers who were abstinent over that same period (11).

The primary objective of this study was to assess one aspect of e-cigarette safety by measuring urinary biomarkers of toxic and/or carcinogenic VOCs in a cross-over (within-subject) study where each participant used e-cigarettes only, cigarettes only, and had a period of enforced nicotine and tobacco product abstinence. Biochemical measures included mercapturic acid metabolites of acrolein, which is believed to be a major contributor to smoking-induced cardiopulmonary disease (5) and is a thermal breakdown product of glycerin in e-cigarettes (16), benzene, a known human carcinogen (17) that can be formed from e-cigarette constituents such as benzoic acid (18), and propylene oxide, an International Agency for Research on Cancer (IARC) class 2B carcinogen (possibly carcinogenic to humans; ref. 19) that can be formed by thermal degradation of propylene glycol (20).

Materials and Methods

Study design

We conducted a two-arm counterbalanced, cross-over study in 36 healthy dual users of e-cigarettes and cigarettes. Participants were asked to smoke cigarettes or vape e-cigarettes only for periods of 7 days, each. During each arm, use of the assigned product and subjective measures were tracked by self-report for 4 days as outpatients, followed by 3 days on a research ward where product use was monitored or abstinence enforced, and biosamples were collected for biomarker measurement. The hospital phase of each arm included a single-dose pharmacokinetic study on the first day of admission (21), followed by 2 days of *ad libitum* access to the assigned product. Furthermore, 2 days of enforced abstinence on the research ward were added immediately after the second arm to examine excretion of toxicant biomarkers during a period of no tobacco product use. In this study, we present biomarkers of toxicants measured in spot urine samples collected at baseline (before product assignment), in 24-hour urine samples collected during the 2 days of *ad libitum* access to the assigned product during each arm, and in 24-hour urine samples collected on the second day of 2 days of abstinence. Known elimination half-lives of the VOC mercapturic acid metabolites measured are 8 hours for the acrylonitrile metabolite (CNEMA; ref. 22), 9 hours for the acrolein (3-HPMA; ref. 23) and benzene metabolites (PMA; ref. 24), and 14 hours for the acrylamide metabolite (AAMA; ref. 23; see the Analytical Chemistry section for full names of the metabolites). Although VOC metabolite levels derived from noncompliant smoking during the at-home period of the e-cigarette arm would potentially carryover to levels measured during e-cigarette use on the research ward, or smoking during the second arm on the research ward would potentially carryover to the abstinence arm, 2–3 days of abstinence from cigarettes are sufficient to observe substantial reductions in urinary mercapturic acid levels to near baseline (25).

Participants

Thirty-six healthy participants recruited via Craigslist.com, Facebook, flyers, and college campus newspapers, completed the study. Participants had to be at least 21 years old, smoke at least five cigarettes per day (CPD) over the past 30 days and use the same e-cigarette device at least once daily on 15 of the past 30 days, and use e-liquids of at least 6 mg/mL nicotine concentration, have no intention to quit smoking or vaping, and at screening, have saliva cotinine and expired carbon monoxide (CO) of ≥ 50 ng/mL and ≥ 5 ppm, respectively, negative pregnancy test (if a woman), and negative urine illicit drug test, except for cannabis. The study was conducted in accordance with the principles of the Declaration of Helsinki, and the protocol was approved by the Institutional Review Board at the University of California San Francisco. Written, informed consent was obtained from each participant and all participants were financially compensated.

Products

Participants used their usual brands of e-cigarettes and cigarettes, provided by the study. The types of e-cigarettes used by study participants were as follows: cig-a-likes ($n = 12$ participants); fixed-power tanks ($n = 15$), variable-power tanks ($n = 6$), and pod e-cigarettes ($n = 3$, all JUULs). Details of the products have been described elsewhere (21).

Experimental procedure

We screened participants for eligibility in an outpatient research clinic where consent was obtained, questionnaires completed, and saliva samples were collected for cotinine measurement. Eligible participants returned for an orientation visit at which time the sequence of products was assigned, and a 4-day supply of the product assigned to the first arm was dispensed for at-home use. During the orientation visit, we also collected a spot urine sample for baseline assessment of exposure biomarkers.

On day 5 of each arm, participants were admitted to one of the clinical research center research smoking rooms at the Zuckerberg San Francisco General Hospital (San Francisco, CA) between 7:00 to 8:00 am. We asked participants to abstain from all tobacco product use starting at 10 pm the night before the hospital admission and we measured expired CO to verify abstinence from cigarettes (≤ 5 ppm). An intravenous line for blood sampling was placed in the forearm followed by a standardized session of product use to examine differences in nicotine pharmacokinetics and subjective effects between e-cigarettes and cigarette use (21).

During the second and third days of admission (of each arm), participants had *ad libitum* access to the assigned product from 8 am to midnight (hospital policy prohibits smoking in the hospital research smoking rooms after midnight). For these 2 days of *ad libitum* use (referred hereafter as *ad libitum* day 1 and day 2), cig-a-like users were given their usual brand of cartridges, fixed-power or variable-power tank users were provided with their usual brand of e-liquid in a vial, which

they used with their own device, and JUUL users were provided with their usual flavor of JUUL pods. During the cigarette arm, participants were given their usual brand of cigarettes. Because the participants were dual users, to meet their required daily nicotine intake, we anticipated an increased consumption of the assigned product compared with self-reported consumption of that product during the screening visit. Accordingly, participants were given an additional number of cartridges, e-liquid vials, pods, or cigarettes during these 2 *ad libitum* access days. All remaining products were collected by study nurses at midnight. No participant ran out of their cartridges, e-liquids, pods, or cigarettes during the day. After the second study arm, participants remained for an additional 2 days, during which they abstained from any nicotine or tobacco products. Twenty-four-hour urine was collected on the second day of *ad libitum* access of each arm and on the last day of abstinence.

Analytical chemistry

We measured mercapturic acid metabolites of VOCs in urine samples using LC-MS/MS by a method described previously (26). The mercapturic acid metabolites measured were as follows, shown as the mercapturic acid metabolite [abbreviation, parent compound(s), limit of quantitation (LOQ)]: 2-hydroxypropylmercapturic acid [2-HPMA, propylene oxide, 0.5 ng/mL]; 3-hydroxypropylmercapturic acid [3-HPMA, acrolein, 1 ng/mL]; 2-carbamoylmercapturic acid [AAMA, acrylamide, 0.5 ng/mL]; 2-cyanoethylmercapturic acid [CNEMA, acrylonitrile, 0.5 ng/mL]; 2-hydroxyethylmercapturic acid [HEMA, acrylonitrile, vinyl chloride, ethylene oxide, 0.5 ng/mL]; 3-hydroxy-1-methylpropylmercapturic acid [HPMMA, crotonaldehyde, 1 ng/mL]; sum of isomers 1-hydroxy-3-buten-2-ylmercapturic acid and 2-hydroxy-3-buten-1-ylmercapturic acid [MHBMA-1+2, 1,3-butadiene, 0.1 ng/mL]; 4-hydroxy-2-buten-1-ylmercapturic acid [MHBMA-3, 1,3-butadiene, 0.1 ng/mL]; methylmercapturic acid [MMA, methylating agents such as 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK), N-nitrosodimethylamine (NDMA), and endogenous methylating agents, 5 ng/mL]; and phenylmercapturic acid [PMA, benzene, 0.1 ng/mL].

Statistical analysis

We imputed biomarker values below the LOQ using the LOQ divided by the square root of 2 ($LOQ/\sqrt{2}$) and we normalized urinary biomarker concentrations by creatinine concentrations, including biomarkers measured in 24-hour urine. We normalized the 24-hour urine samples for creatinine because the spot urine sample collected at baseline had to be normalized for creatinine. However, differences in 24-hour urinary biomarker levels across arms were consistent with or without creatinine correction. Because the concentrations were approximately log-normally distributed, biomarker concentrations were log transformed.

Our primary analysis was a comparison of day 2 urinary biomarker concentrations over the three conditions via repeated measures ANCOVA. We focused on day 2 because, as stated

before, VOC metabolite concentrations derived from noncompliant smoking during the at-home phase of the e-cigarette arm would be reduced substantially by day 2 on the research ward. We conducted *post hoc* pairwise comparisons between study arms and applied Bonferroni correction for multiple comparisons. Covariates included sex and treatment order and a random effect of participants. We calculated geometric mean and 95% confidence intervals for the relative ratio of concentrations in cigarette versus e-cigarette arms and e-cigarette versus abstinence. We evaluated differences in urinary biomarker levels on day 2 between e-cigarette device types with Wilcoxon rank-sum tests. We computed Spearman correlation coefficients between 24-hour biomarker concentrations and corresponding 24-hour area under the plasma nicotine concentration-time curve (AUC) for the e-cigarette and cigarette arms, respectively, as a way to examine the relationship between product use on the research ward and VOC exposure. (Plasma nicotine AUC is reported in another article; ref. 27). Finally, we computed the frequencies of participants with e-cigarette to abstinence biomarker level ratios of at least 1.25 or at least 1.50 by device type and flavor category, representing at least 25% and 50% higher biomarker levels from e-cigarette use compared with abstinence respectively.

All analyses were considered significant at two-tailed $P < 0.05$ and were conducted in SAS Version 9.4 and R Version 3.4.

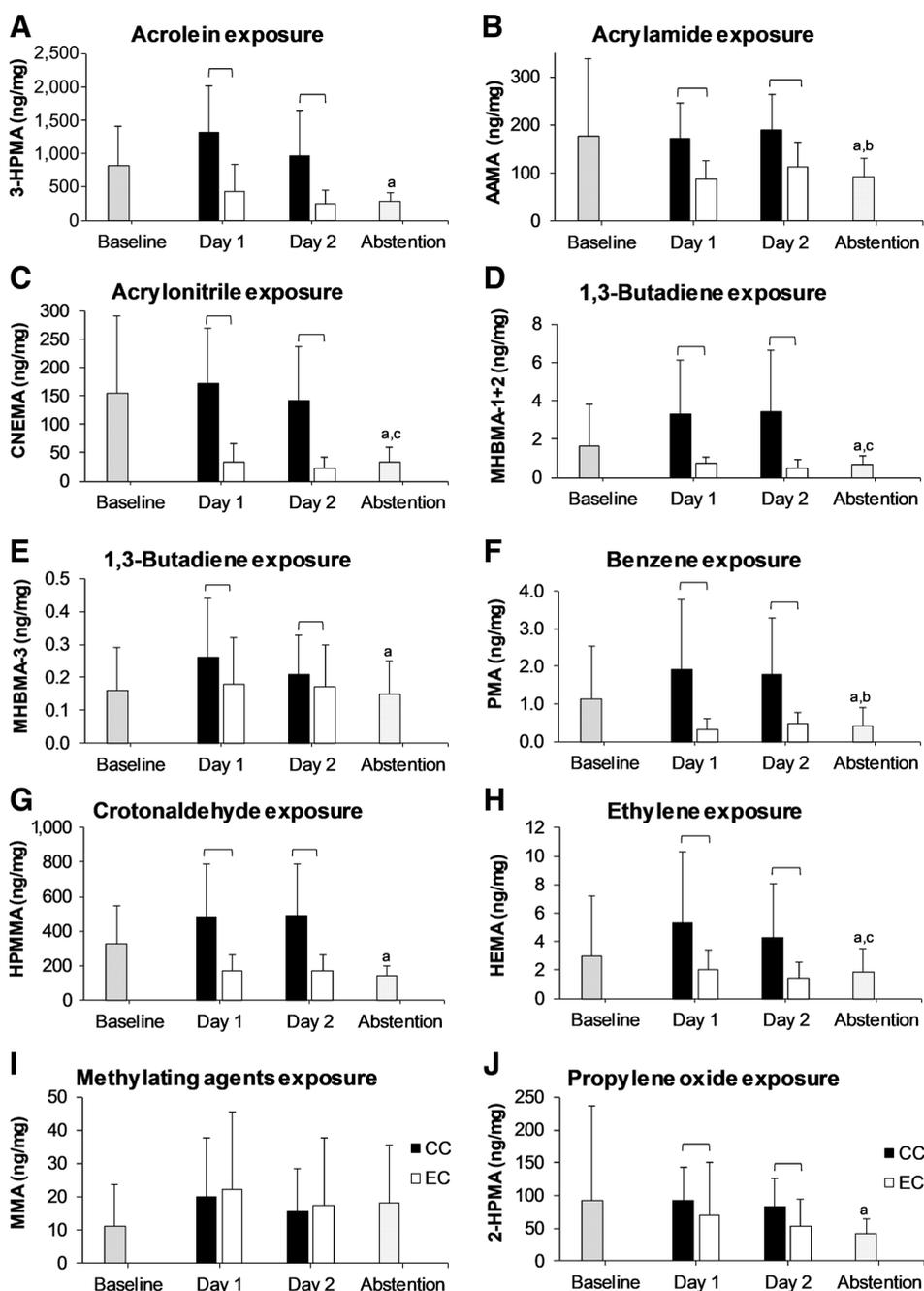
Results

Of 36 participants enrolled (8 women), 2 were Asian, 3 were African American/Black, 4 were Latino, 22 were White, and 5 were mixed race. On average, participants smoked 12.9 ± 6.4 (mean \pm SD) CPD, used e-cigarettes on 22.6 ± 7.3 days of the past 30 days, and on days that they used the e-cigarette, they used the e-cigarette 8.1 ± 7.2 times. Average screening saliva cotinine was 189 ± 92.8 ng/mL (range 119–248 ng/mL). Eight participants (22.2%) used a dessert/candy flavored e-liquid/e-cigarette, 5 (13.9%) used a fruit flavor, 5 (13.9%) used a menthol flavor, and 18 (50%) used a tobacco flavor.

VOC exposure from e-cigarettes versus cigarettes

Concentrations of metabolites of VOCs collected in spot urine samples at baseline and in 24-hour urine collected during *ad libitum* use of the assigned product on day 1 and day 2, respectively, and during enforced abstinence are shown in **Fig. 1A–J**. **Table 1** shows these VOC metabolite concentrations in 24-hour urine collected on day 2 during e-cigarette or cigarette use and during abstinence. Concentrations of all VOC metabolites were significantly higher during both days of cigarette use compared with e-cigarette use (all $P < 0.001$) except for MMA (the metabolite of methylating agents; **Fig. 1A–J**). The geometric means of the fold-difference in concentrations of these VOC metabolites when using cigarettes relative to that of e-cigarettes ranged from 1.31 for MHBMA-3 (one of the butadiene metabolites) to 7.09 for CNEMA (acrylonitrile metabolite; **Table 2**).

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**Figure 1.**

Concentration of metabolites of VOCs at baseline before study products were assigned, during days 1 and 2 of each arm, and during abstinence from nicotine and tobacco products. Square brackets, significant difference between combustible cigarettes (CC) and e-cigarettes (EC). During “baseline” participants smoked and/or vaped in their naturalistic environment. The black column represents smoking CC on days 1 and 2 while the white column represents EC use on days 1 and 2. During the “abstinence” arm, participants abstained from all tobacco products. For comparisons: a, significantly lower than combustible cigarette use on day 2; b, significantly lower than e-cigarette use on day 2; and c, significantly higher than e-cigarette use on day 2.

VOC exposure from e-cigarettes versus abstinence

Within-subject concentrations of metabolites of acrylamide (AAMA) and benzene (PMA) were significantly higher during e-cigarette use (day 2 of the e-cigarette arm) compared with abstinence, with average within-subject fold-difference of 1.21 ($P = 0.019$) and 1.46 ($P = 0.006$), respectively, among all participants (Table 1). Considering the absolute concentration of each metabolite, compared with abstinence, most participants had higher levels of AAMA (frequency = 63.9%), PMA (66.7%), and 2-HPMA (58.3%) during e-cigarette use

(Fig. 2A–C). Metabolites of acrylonitrile (CNEMA), 1,3-butadiene (MHBMA-1+2), and ethylene oxide (HEMA) were significantly lower during e-cigarette use than during abstinence, with average fold-differences of 0.64 ($P < 0.001$), 0.63 ($P = 0.001$), and 0.82 ($P = 0.010$), respectively.

The order of assigned products influenced the magnitude of changes in the concentrations of some metabolites measured during e-cigarette use compared with abstinence, indicative of potential carryover effect on biomarker levels from smoking cigarettes (Table 1; Fig. 2D–F). The within-subject

VOC Exposure from Vaping, Smoking, and Abstinence

Table 1. Concentrations of mercapturic acid metabolites of VOCs measured in 24-hour urine collected during cigarette smoking, e-cigarette use, and abstinence.

Exposure (biomarker) (ng/mg creatinine)	Cigarette (Mean, SD)	E-cigarette (Mean, SD)	Abstinence (Mean, SD)	CC to EC ratio (GM, 95% CI)	Ratio of EC to Abstinence (GM, 95% CI)		
					All subjects	CC at arm 2 ^a	EC at arm 2 ^b
Acrolein (3-HPMA)	965.7 (674.3)	258.8 (195.2)	279.9 (140.0)	3.70 (2.85–4.79) ^c	0.82 (0.67–1.01)	0.81 (0.62–1.04)	0.83 (0.61–1.14)
Acrylamide (AAMA)	190.2 (72.8)	112.9 (50.8)	92.8 (37.2)	1.70 (1.50–1.92) ^c	1.21 (1.03–1.43) ^d	1.15 (0.93–1.43)	1.27 (1.01–1.61)
Acrylonitrile (CNEMA)	140.9 (95.5)	21.8 (19.7)	32.9 (27.6)	7.09 (5.88–8.54) ^c	0.64 (0.56–0.74) ^e	0.50 (0.42–0.59)	0.81 (0.71–0.92)
1,3-Butadiene (MHBMA-1+2)	3.43 (3.23)	0.51 (0.42)	0.70 (0.40)	5.80 (3.73–9.00) ^c	0.63 (0.48–0.82) ^e	0.47 (0.32–0.68)	0.83 (0.61–1.13)
1,3-Butadiene (MHBMA-3)	0.21 (0.12)	0.17 (0.13)	0.15 (0.10)	1.31 (1.06–1.61) ^c	1.05 (0.84–1.30)	0.92 (0.66–1.29)	1.18 (0.91–1.53)
Benzene (PMA)	1.77 (1.52)	0.48 (0.31)	0.42 (0.48)	3.21 (2.53–4.07) ^c	1.46 (1.13–1.90) ^d	0.94 (0.64–1.37)	2.18 (1.71–2.77)
Crotonaldehyde (HPMMA)	489.9 (297.7)	168.1 (95.36)	145.6 (55.3)	2.77 (2.34–3.29) ^c	1.08 (0.94–1.25)	1.23 (0.99–1.54)	0.97 (0.82–1.14)
Ethylene oxide (HEMA)	4.28 (3.82)	1.47 (1.06)	1.84 (1.64)	2.55 (2.10–3.10) ^c	0.82 (0.71–0.95) ^e	0.78 (0.66–0.93)	0.86 (0.69–1.08)
Methylating agent (MMA)	15.51 (12.84)	17.30 (20.32)	18.0 (17.5)	1.01 (0.85–1.20)	0.94 (0.77–1.15)	1.18 (0.97–1.44)	0.77 (0.57–1.04)
Propylene oxide (2-HPMA)	82.7 (43.5)	53.6 (41.1)	41.7 (22.9)	1.69 (1.33–2.16) ^c	1.17 (0.98–1.39)	1.08 (0.88–1.33)	1.24 (0.95–1.64)

Note: During day 2 of the cigarette arm, per cent below the LOQ for each metabolite was as follows: 3-HPMA (0%); AAMA (0%); CNEMA (0%); MHBMA-1+2 (2.8%); MHBMA-3 (58.3%); PMA (0%); HPMMA (0%); HEMA (0%); MMA (36.1%); and 2-HPMA (0%). Per cent below LOQ on day 2 of the e-cigarette arm were as follows: 3-HPMA (0%); AAMA (0%); CNEMA (0%); MHBMA-1+2 (41.7%); MHBMA-3 (83.3%); PMA (22.2%); HPMMA (0%); HEMA (0%); MMA (33.3%); and 2-HPMA (0%). Per cent below LOQ during abstinence were as follows: 3-HPMA (0%); AAMA (0%); CNEMA (0%); MHBMA-1+2 (80.6%); MHBMA-3 (97.2%); PMA (30.6%); HPMMA (0%); HEMA (25.0%); MMA (33.3%); and 2-HPMA (0%).

Abbreviations: CC, combustible cigarette; EC, e-cigarette.

^aCC at arm 2 = participants who were assigned to smoke combustible cigarettes during arm 2 immediately before the abstinence days.

^bEC at arm 2 = participants who were assigned to vape e-cigarettes during arm 2, immediately before the abstinence days.

^cSignificant difference between combustible cigarettes and e-cigarettes.

^dSignificantly higher during e-cigarette use compared with abstinence.

^eSignificantly lower during e-cigarette use compared with abstinence.

fold-difference from e-cigarette use compared with abstinence in concentrations of metabolites of acrylamide, benzene, and propylene oxide were higher in participants who used e-cigarettes during the second arm (i.e., immediately before the abstinence days) compared with those who smoked cigarettes during the second arm (Table 1). Of note, the average concentration of the benzene metabolite (PMA) was 2.18-fold higher during e-cigarette use relative to abstinence among participants who were assigned e-cigarettes during the second arm, while it was a 0.94-fold difference in participants who smoked cigarettes during the second arm.

VOC exposure across different types of e-cigarettes

In Table 2, we present concentrations of VOC metabolites across users of different types of e-cigarettes during the e-

cigarette arm. When all participants were considered, the 1,3-butadiene metabolite (MHBMA-3) and the propylene oxide metabolite (2-HPMA) were significantly different across e-cigarette devices, with higher levels in variable-power tank users. When pod users were excluded from the analysis, 2-HPMA was the only VOC metabolite that differed significantly by device type.

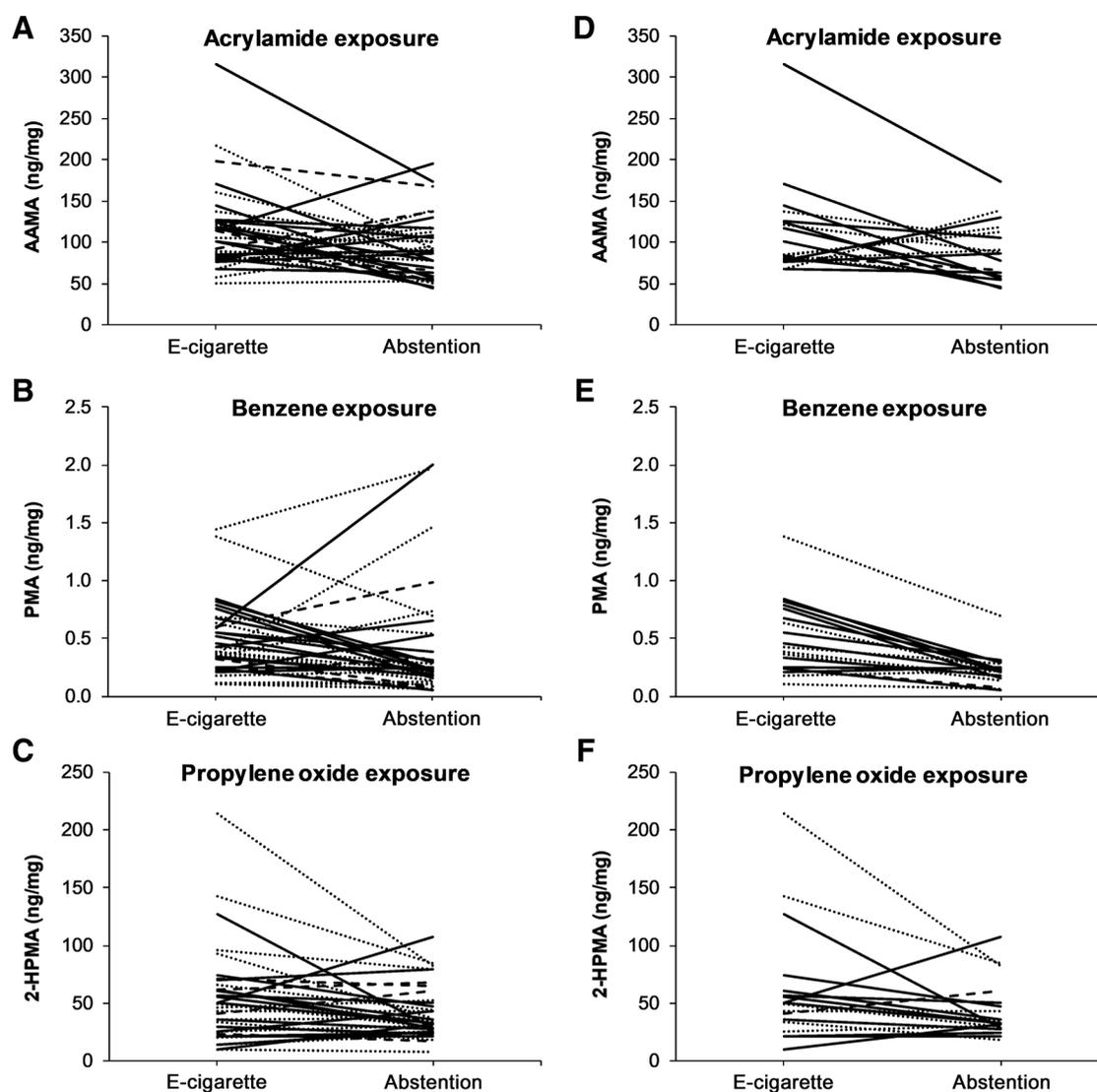
Correlations between VOCs and nicotine exposure

Spearman correlation coefficients between VOC metabolite levels and plasma nicotine AUC over 24 hours for day 2 of the cigarette and e-cigarette arms are shown in Table 3. For the cigarette arm, except for MHBMA-1+2 (1,3-butadiene) and MMA (methylating agents), correlation between plasma nicotine AUC and urinary VOC metabolites was moderate and

Table 2. Concentrations of mercapturic acid metabolites of VOCs measured in 24-hour urine during e-cigarette use.

Exposure (biomarker) (ng/mg creatinine)	Cig-a-like (mean, SD) (n = 12)	Fixed-power (mean, SD) (n = 15)	Variable-power (mean, SD) (n = 6)	Pod (mean, SD) (n = 3)	Difference (P)	
					All included	Pods excluded
Acrolein (3-HPMA)	260.6 (236.0)	238.7 (189.8)	340.7 (173.1)	188.0 (49.9)	0.081	0.100
Acrylamide (AAMA)	117.4 (65.9)	103.2 (43.6)	118.8 (35.1)	131.9 (58.9)	0.325	0.311
Acrylonitrile (CNEMA)	27.5 (28.6)	20.8 (15.6)	17.1 (3.79)	13.1 (13.4)	0.442	0.950
1,3-Butadiene (MHBMA-1+2)	0.51 (0.36)	0.57 (0.50)	0.50 (0.43)	0.19 (0.07)	0.055	0.730
1,3-Butadiene (MHBMA-3)	0.16 (0.09)	0.15 (0.09)	0.24 (0.23)	0.08 (0.05)	0.019	0.363
Benzene (PMA)	0.54 (0.22)	0.48 (0.41)	0.38 (0.20)	0.39 (0.19)	0.081	0.129
Crotonaldehyde (HPMMA)	152.4 (68.0)	194.2 (128.6)	132.5 (30.5)	171.3 (71.6)	0.477	0.242
Ethylene oxide (HEMA)	1.56 (0.64)	1.35 (1.25)	1.73 (1.31)	1.27 (1.40)	0.090	0.124
Methylating agent (MMA)	15.7 (9.3)	18.8 (27.0)	18.7 (23.7)	13.6 (12.8)	0.601	0.865
Propylene oxide (2-HPMA)	34.2 (17.4)	62.6 (55.0)	75.8 (26.4)	42.2 (19.6)	<0.001	0.001

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**Figure 2.**

Within-subject changes in mercapturic acid metabolites of acrylamide (A), benzene (B), and propylene oxide (C) for all participants and for participants who used e-cigarettes during arm 2, immediately before 2 days of abstinence, showing acrylamide (D), benzene (E), and propylene oxide (F). Solid black line, cig-a-like; dotted black line, fixed-power tank e-cigarette; solid gray line, variable-power tank e-cigarette; broken gray line, JUUL pod e-cigarette; A-C, all participants (left); and D-F, e-cigarette use in second arm (right).

statistically significant. For the e-cigarette arm, the correlation between plasma nicotine AUC and 2-HPMA (propylene oxide) was small and statistically significant but others were not significant.

Evaluation of elevated exposures from e-cigarette use by device type and flavors

We present the frequency of participants whose biomarker levels were at least 25% (Table 4 A) or 50% (Table 4B) higher during e-cigarette use relative to abstinence by device types and e-liquid flavors. Notably, 21 (58.3%) of 36 participants had at least 50% higher PMA (benzene) levels during e-cigarette use compared with abstinence, including 8 of 12 (66.7%) cig-a-like

users, 8 of 15 (53.3%) fixed-power tank users, 3 of 6 (50%) variable-power tank users, and 2 of 3 (66.7%) pod users. Across flavors, 3 of 8 (37.5%) users of dessert/candy e-liquids, 5 of 5 (100%) users of fruit flavors, 3 of 5 (60%) users of menthol flavor, and 11 of 18 (61.1%) users of tobacco flavors had PMA levels that were at least 50% higher during e-cigarette use than abstinence.

Discussion

The public health burden of e-cigarette use is a balance between their potential benefits as a form of harm reduction for smokers and their direct harm to nonsmokers who vape. The

Table 3. Spearman correlation coefficients between mercapturic acid metabolites of VOCs measured in urine collected during a day of combustible cigarette smoking or e-cigarette use and area under the plasma nicotine AUC measured during the corresponding day.

Exposure (biomarker)	Plasma nicotine during cigarette smoking		Plasma nicotine during e-cigarette use	
	Spearman	P	Spearman	P
Acrolein (3-HPMA)	0.42	0.011	0.31	0.066
Acrylamide (AAMA)	0.47	0.004	0.13	0.456
Acrylonitrile (CNEMA)	0.53	0.001	0.14	0.428
1,3-Butadiene (MHBMA-1+2)	0.27	0.107	0.18	0.295
1,3-Butadiene (MHBMA-3)	0.59	<0.001	0.11	0.530
Benzene (PMA)	0.44	0.007	0.18	0.307
Crotonaldehyde (HPMMA)	0.43	0.008	0.29	0.084
Ethylene oxide (HEMA)	0.20	0.244	0.16	0.352
Methylating agent (MMA)	0.41	0.012	0.17	0.331
Propylene oxide (2-HPMA)	0.36	0.029	0.37	0.027

Note: Correlations were between mercapturic acid levels measured in 24-hour urine and the area under the plasma nicotine concentration-time curve over 24 hours.

findings of our study support the harm reduction potential of e-cigarettes for smokers but also suggest that e-cigarettes may have deleterious effects in nonsmoking vapers. We found that dual users confined to a research ward were exposed to substantially lower levels of toxic and/or carcinogenic VOCs when they used e-cigarettes compared with when they smoked

cigarettes. These findings align with previous cross-sectional studies and longitudinal ambulatory switching studies which found that e-cigarette use resulted in lower systemic exposure to toxicants compared with smoking (10, 12).

Of note however, we found higher levels of metabolites of acrylamide, benzene, and possibly propylene oxide during e-cigarette use relative to enforced abstinence, suggesting that e-cigarette use results in higher systemic exposure to these toxic/carcinogenic VOCs. These findings are important because, as far as we know, this is the first nonindustry-associated assessment of toxicant exposure from use of commercial e-cigarettes in a setting where e-cigarette use is monitored and abstinence enforced.

We found no published study explaining how e-cigarette use can lead to increased exposure to acrylamide, an IARC group 2A carcinogen (probable human carcinogen). Sources of acrylamide exposure include manufacturing, chemical, and agricultural industries, but French fries, potato chips, cereals, and coffee are important dietary sources (28). Acrylamide is generated through the Maillard reactions of food products, which are heat-dependent reactions of glucose with amino acids, particularly asparagine, peptides, and aromatic amines (29, 30). These reactions are plausible during e-cigarette use. Amino acids are not a significant constituent of e-liquids but studies have reported greater excretion of aromatic amines from exclusive e-cigarette users compared with controls (31), suggesting that aromatic amines are given off in e-cigarette aerosols and might contribute to acrylamide formation. Another

Table 4. Frequency of participants whose biomarker levels were at least 25% or 50% higher during e-cigarette use relative to abstinence.

Exposure (biomarker)	All	E-cigarette type				Flavor type			
		Cig-a-like	Fixed-power	Variable-power	Pod	Dessert or Candy	Fruit	Menthol	Tobacco
Sample size (N)	36	12	15	6	3	8	5	5	18
A. Participants with $\geq 25\%$ increase in VOC biomarker levels during e-cigarette use compared with abstinence (n, %)									
Acrolein (3-HPMA)	8 (22.2)	2 (16.7)	4 (26.7)	2 (33.3)	0 (0.0)	1 (12.5)	2 (40.0)	1 (20.0)	4 (22.2)
Acrylamide (AAMA)	17 (47.2)	6 (50.0)	6 (40.0)	3 (50.0)	2 (66.7)	3 (37.5)	3 (60.0)	3 (60.0)	9 (50.0)
Acrylonitrile (CNEMA)	1 (2.8)	0 (0.0)	0 (0.0)	1 (16.7)	0 (0.0)	1 (12.5)	0 (0.0)	1 (20.0)	0 (0.0)
1,3-Butadiene (MHBMA-1+2)	7 (19.4)	2 (16.7)	3 (20.0)	2 (33.3)	0 (0.0)	2 (25.0)	1 (20.0)	2 (40.0)	2 (11.1)
1,3-Butadiene (MHBMA-3)	13 (36.1)	4 (33.3)	5 (33.3)	4 (66.7)	0 (0.0)	3 (37.5)	2 (40.0)	3 (60.0)	6 (33.3)
Benzene (PMA)	23 (63.9)	9 (75.0)	9 (60.0)	3 (50.0)	2 (66.7)	4 (50.0)	5 (100.0)	4 (80.0)	12 (66.7)
Crotonaldehyde (HPMMA)	15 (41.7)	5 (41.7)	6 (40.0)	2 (33.3)	2 (66.7)	5 (62.5)	2 (40.0)	5 (100)	6 (33.3)
Ethylene oxide (HEMA)	6 (16.7)	2 (16.7)	2 (13.3)	2 (33.3)	0 (0.0)	1 (12.5)	2 (40.0)	1 (20.0)	1 (5.6)
Methylating agent (MMA)	11 (30.6)	1 (8.3)	7 (46.7)	3 (50.0)	0 (0.0)	3 (37.5)	4 (80.0)	3 (60.0)	4 (22.2)
Propylene oxide (2-HPMA)	18 (50.0)	6 (50.0)	8 (53.3)	3 (50.0)	1 (33.3)	2 (25.0)	5 (100.0)	2 (40.0)	10 (55.6)
B. Participants with $\geq 50\%$ increase in VOC biomarker levels during e-cigarette use compared with abstinence (n, %)									
Acrolein (3-HPMA)	4 (11.1)	0 (0.0)	2 (13.3)	2 (33.3)	0 (0.0)	1 (12.5)	0 (0.0)	1 (20.0)	3 (16.7)
Acrylamide (AAMA)	12 (33.3)	5 (41.7)	4 (26.7)	2 (33.3)	1 (33.3)	3 (37.5)	0 (0.0)	3 (60.0)	6 (33.3)
Acrylonitrile (CNEMA)	1 (2.8)	0 (0.0)	0 (0.0)	1 (16.7)	0 (0.0)	1 (12.5)	0 (0.0)	1 (20.0)	0 (0.0)
1,3-Butadiene (MHBMA-1+2)	6 (16.7)	2 (16.7)	2 (13.3)	2 (33.3)	0 (0.0)	2 (25.0)	1 (20.0)	2 (40.0)	1 (5.6)
1,3-Butadiene (MHBMA-3)	10 (27.8)	3 (25.0)	4 (26.7)	3 (50.0)	0 (0.0)	2 (25.0)	2 (40.0)	2 (40.0)	5 (27.8)
Benzene (PMA)	21 (58.3)	8 (66.7)	8 (53.3)	3 (50.0)	2 (66.7)	3 (37.5)	5 (100.0)	3 (60.0)	11 (61.1)
Crotonaldehyde (HPMMA)	8 (22.2)	2 (16.7)	5 (33.3)	0 (0.0)	1 (33.3)	2 (25.0)	1 (20.0)	2 (40.0)	4 (22.2)
Ethylene oxide (HEMA)	4 (11.1)	2 (16.7)	1 (6.7)	1 (16.7)	0 (0.0)	1 (12.5)	1 (20.0)	1 (20.0)	0 (0.0)
Methylating agent (MMA)	7 (19.4)	1 (8.3)	5 (33.3)	1 (16.7)	0 (0.0)	0 (0.0)	4 (80.0)	0 (0.0)	3 (16.7)
Propylene oxide (2-HPMA)	11 (30.6)	3 (25.0)	5 (33.3)	3 (50.0)	0 (0.0)	1 (12.5)	4 (80.0)	1 (20.0)	6 (33.3)

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potential pathway of acrylamide formation is the reaction between acrylic acid, formed from oxidation of acrolein, and ammonia, which could be potentially generated by thermal decomposition of nitrogen-containing compounds (29). Of relevance to the latter route, ethyl acrylate, the ethyl ester of acrylic acid, is a major volatile constituent of organic passion fruit pulp (32), is found in pineapples, grapes, and vanilla (33), and is a flavor additive (34). Ethyl acrylate would react with ammonia much more readily than would acrylic acid. Furthermore, because acrylamide is used in the manufacturing of some plastics and adhesives, we cannot rule out the possibility that acrylamide residues that remain in plastics used in the manufacturing of e-cigarette devices or in plastic equipment and containers used in manufacturing and transporting of e-liquids is the source.

We did not see a significant correlation between plasma nicotine AUC over 24-hour and 24-hour urinary AAMA levels during the e-cigarette arm, suggesting that other sources could have contributed to acrylamide exposure. Also, the elimination half-life of AAMA is 14 hours (23), thus noncompliant smoking during the at-home phase of the e-cigarette arm could have influenced AAMA levels measured during the research ward phase of the e-cigarette arm. Nevertheless, during the smoking arm, there was no increase in AAMA levels from day 1 to day 2 (Fig. 1A–J), arguing against substantial carryover effect of smoking on AAMA levels. Furthermore, among those who were assigned the e-cigarette during the second arm, AAMA levels were significantly higher during e-cigarette use compared with abstinence, providing evidence of the contribution of e-cigarettes to acrylamide exposure. In summary, acrylamide formation in e-cigarettes is plausible and our findings are suggestive, but a firm conclusion cannot be made that e-cigarette use leads to acrylamide exposure.

Pankow and colleagues demonstrated that benzene can be generated from thermal degradation of the humectants, propylene glycol and vegetable glycerin, and additives, such as benzoic acid and benzaldehyde (18). Most participants with elevated levels of the benzene metabolite (PMA) during e-cigarette use relative to abstinence were users of cig-a-likes or fixed-power e-cigarettes (i.e., low-powered devices); users of fruit or tobacco flavors also showed elevated benzene exposure. Although generation of aldehydes and VOCs in e-cigarette aerosol is known to be temperature dependent (16), and thus higher exposure to these toxicants are expected in users of high-powered devices, our study raises questions about toxicant exposure from use of low-powered devices. Benzene has been detected in some refill e-liquids and cartridges (35), potentially serving as a source of benzene even in devices that operate at low power/temperature settings.

Users of variable-power e-cigarettes, which are typically operated at higher power and temperatures (36), had elevated excretion of the propylene oxide metabolite (2-HPMA) compared with users of the other types of e-cigarettes. Propylene

oxide, an IARC group 2B carcinogen (possibly carcinogenic to humans; ref. 19), can be derived from propylene glycol in the presence of weak bases and heat (37). Aerosol generation is greater in high power e-cigarettes, resulting in greater nicotine intake and potentially more propylene oxide generation. The significant correlation between 2-HPMA levels and plasma nicotine AUC during the e-cigarette arm is evidence for propylene oxide generation in e-cigarettes.

We found no evidence of significant differences in exposure to the other VOCs, including acrolein, across device types. This observation regarding acrolein was surprising because vaping machine studies have reported substantial acrolein generation from e-cigarettes, particularly at higher power settings (38–40). However, it is possible that background exposure to acrolein, primarily from food sources through thermal breakdown of animal and vegetable fats, carbohydrates, and amino acids, or even endogenous production of acrolein (41), could overwhelm the contribution of e-cigarettes to acrolein exposure. To minimize the contribution of food to toxicant exposure during the participants' stay on the research ward, we did not allow charbroiled meats and fried foods. Furthermore, because participants were admitted to the hospital on the same day of each week, they were served the same meals on each named day, thus reducing variation in diet-related exposures within- and between-participants.

Despite lower risks of e-cigarettes relative to cigarettes, questions remain of the inherent toxicity of e-cigarettes, and risks to nonsmoking adults and children who vape. Our findings of potentially increased systemic exposure to acrylamide, benzene, and propylene oxide from e-cigarette use are particularly concerning given that these VOCs are known or suspected human carcinogens. A previous study found higher levels of another benzene metabolite, trans,trans-muconic acid, in baseline urine samples of e-cigarette users compared with nontobacco users enrolled in a laboratory study, but polyuse of other tobacco products could not be ruled out (15). Our findings also raise concerns about benzene exposure among JUUL users because a major constituent of JUUL pod fluids is benzoic acid. Although the study by Pankow and colleagues did not detect benzene in JUUL aerosol, 2 of 3 JUUL users in our study had elevated PMA excretion during e-cigarette use relative to abstinence.

Using the same analytic chemistry methods as used in the current study, we measured all 10 VOC metabolites in smokers enrolled in a clinical trial at 10 sites across the United States. Levels of all metabolites in smokers in the former study were comparable with levels measured at baseline and during the cigarette arm in this study (42). In addition, of the same eight VOC biomarkers utilized between studies, average levels of 2-HPMA, 3-HPMA, and HEMA measured in e-cigarette-only users from a nationally representative sample were comparable with levels measured during the e-cigarette arm of this study; the average level of AAMA was about 2-times higher and

CNEMA was over 5-times higher during the e-cigarette arm of this study, while the average levels of HPMMA, MHBMA-1+2, and PMA were 2.6-, 8-, and 2-times higher, respectively, in e-cigarette-only users in the former study compared with during the e-cigarette arm of this study (12).

A strength of our study is its cross-over design, in which each participant served as their own control. However, while we counterbalanced the order of e-cigarette and cigarette arms, the 2 days of abstinence always followed the second arm. Because the VOC metabolites have half-lives of several hours, from at least 8 hours for CNEMA (acrylonitrile; ref. 22) to 14 hours for the AAMA (acrylamide; ref. 23), there was likely carryover from product use to abstinence, particularly when cigarettes were assigned immediately before the abstinence days (see Fig. 2A–F). Thus, we could have underestimated differences in VOC exposure from e-cigarette use compared with abstinence. On the other hand, during the cigarette and e-cigarette arms, the levels of biomarkers at day 1 and day 2 were consistent, potentially indicating minimal carryover from 1 day to the next. Another limitation of our study is that most participants were males, which limits assessment of sex differences. Furthermore, assessment of differences by device type was limited by the small sample size of variable-power tank users and JUUL pod users enrolled in the study.

In conclusion, e-cigarettes expose users to lower levels of toxic VOCs, supporting their harm reduction potential among smokers. However, some e-cigarettes potentially expose users to VOCs such as acrylamide, benzene, and propylene oxide, and may pose health risks to nonsmoking users. For example, more cig-a-like users had elevated benzene exposure compared with users of other types of e-cigarettes. Further studies are needed to examine what design features of e-cigarettes and user behaviors lead to elevated toxicant exposure. Regulation of e-cigarettes must include a balanced approach to maximize their potential for harm reduction among adult smokers and minimize their risk to nonsmoking users, including minimizing exposure to toxic VOCs.

References

- Benowitz NL. Nicotine addiction. *N Engl J Med* 2010;362:2295–303.
- Ahmad S, Zafar I, Mariappan N, Husain M, Wei C-C, Vetal N, et al. Acute pulmonary effects of aerosolized nicotine. *Am J Physiol Lung Cell Mol Physiol* 2018;316:L94–L104.
- Schweitzer KS, Chen SX, Law S, Van Demark M, Poirier C, Justice MJ, et al. Endothelial disruptive proinflammatory effects of nicotine and e-cigarette vapor exposures. *Am J Physiol Lung Cell Mol Physiol* 2015;309:L175–L187.
- Fowles J, Dybing E. Application of toxicological risk assessment principles to the chemical constituents of cigarette smoke. *Tob Control* 2003;12:424–30.
- Haussmann H-J. Use of hazard indices for a theoretical evaluation of cigarette smoke composition. *Chem Res Toxicol* 2012;25:794–810.
- National Academies of Sciences Engineering and Medicine. Public health consequences of e-cigarettes. Washington, DC:National Academies Press;2018.
- Gottlieb S, Zeller M. A nicotine-focused framework for public health. *N Engl J Med* 2017;377:1111–4.
- Cravo AS, Bush J, Sharma G, Savioz R, Martin C, Craige S, et al. A randomised, parallel group study to evaluate the safety profile of an electronic vapour product over 12 weeks. *Regul Toxicol Pharmacol* 2016;81:S1–S14.
- Goniewicz ML, Gawron M, Smith DM, Peng M, Jacob P, Benowitz NL. Exposure to nicotine and selected toxicants in cigarette smokers who switched to electronic cigarettes: a longitudinal within-subjects observational study. *Nicotine Tob Res* 2017;19:160–7.
- Pulvers K, Emami AS, Nollen NL, Romero DR, Strong DR, Benowitz NL, et al. Tobacco consumption and toxicant exposure of cigarette smokers using electronic cigarettes. *Nicotine Tob Res* 2018;20:206–14.
- D’Ruiz CD, Graff DW, Robinson E. Reductions in biomarkers of exposure, impacts on smoking urge and assessment of product use and tolerability in adult smokers following partial or complete substitution

Disclosure of Potential Conflicts of Interest

N.L. Benowitz is a consultant for Pfizer (paid consultant), Achieve Life Sciences, has provided expert testimony against tobacco companies. No potential conflicts of interest were disclosed by the other authors.

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- of cigarettes with electronic cigarettes. *BMC Public Health* 2016;16:543.
12. Goniewicz ML, Smith DM, Edwards KC, Blount BC, Caldwell KL, Feng J, et al. Comparison of nicotine and toxicant exposure in users of electronic cigarettes and combustible cigarettes. *JAMA Netw Open* 2018;1:e185937.
 13. Shahab L, Goniewicz ML, Blount BC, Brown J, McNeill A, Alwis KU, et al. Nicotine, carcinogen, and toxin exposure in long-term e-cigarette and nicotine replacement therapy users: a cross-sectional study. *Ann Intern Med* 2017;166:390–400.
 14. Rubinstein ML, Delucchi K, Benowitz NL, Ramo DE. Adolescent exposure to toxic volatile organic chemicals from e-cigarettes. *Pediatrics* 2018;141:e20173557.
 15. Lorkiewicz P, Riggs DW, Keith RJ, Conklin DJ, Xie Z, Sutaria S, et al. Comparison of urinary biomarkers of exposure in humans using electronic cigarettes, combustible cigarettes, and smokeless tobacco. *Nicotine Tob Res* 2019;21:1228–38.
 16. Sleiman M, Logue JM, Montesinos VN, Russell ML, Litter MI, Gundel LA, et al. Emissions from electronic cigarettes: key parameters affecting the release of harmful chemicals. *Environ Sci Technol* 2016;50:9644–51.
 17. McMichael A. Carcinogenicity of benzene, toluene and xylene: epidemiological and experimental evidence. *IARC Sci Publ* 1988;85:3–18.
 18. Pankow JF, Kim K, McWhirter KJ, Luo W, Escobedo JO, Strongin RM, et al. Benzene formation in electronic cigarettes. *PLoS One* 2017;12:e0173055.
 19. Smith C, Perfetti T, Rumble M, Rodgman A, Doolittle D. “IARC Group 2B carcinogens” reported in cigarette mainstream smoke. *Food Chem Toxicol* 2001;39:183–205.
 20. Laino T, Tuma C, Moor P, Martin E, Stolz S, Curioni A. Mechanisms of propylene glycol and triacetin pyrolysis. *J Phys Chem A* 2012;116:4602–9.
 21. St.Helen G, Nardone N, Addo N, Dempsey D, Havel C, Jacob P III, et al. Differences in nicotine intake and effects from electronic and combustible cigarettes among dual users. *Addiction*.2019.
 22. Jakubowski M, Linhart I, Pielas G, Kopecký J. 2-Cyanoethylmercapturic acid (CEMA) in the urine as a possible indicator of exposure to acrylonitrile. *Occup Environ Med* 1987;44:834–40.
 23. Watzek N, Scherbl D, Feld J, Berger F, Doroshenko O, Fuhr U, et al. Profiling of mercapturic acids of acrolein and acrylamide in human urine after consumption of potato crisps. *Mol Nutr Food Res* 2012;56:1825–37.
 24. Van Sittert N, Boogaard P, Beulink G. Application of the urinary S-phenylmercapturic acid test as a biomarker for low levels of exposure to benzene in industry. *Occup Environ Med* 1993;50:460–9.
 25. Carmella SG, Chen M, Han S, Briggs A, Jensen J, Hatsukami DK, et al. Effects of smoking cessation on eight urinary tobacco carcinogen and toxicant biomarkers. *Chem Res Toxicol* 2009;22:734–41.
 26. Jacob P, Raddaha AHA, Dempsey D, Havel C, Peng M, Yu L, et al. Comparison of nicotine and carcinogen exposure with water pipe and cigarette smoking. *Cancer Epidemiol Biomarkers Prev* 2013;22:765–72.
 27. Harvanko AM, St.Helen G, Nardone N, Addo N, Benowitz N. Twenty-four hour subjective and pharmacological effects of *ad libitum* electronic and combustible cigarette use among dual users. *Addiction* 2019.
 28. Sharp D. Acrylamide in food. *Lancet* 2003;361:361–2.
 29. Becalski A, Lau BP-Y, Lewis D, Seaman SW. Acrylamide in foods: occurrence, sources, and modeling. *J Agric Food Chem* 2003;51:802–8.
 30. Gokhale MY, Kearney WR, Kirsch LE. Glycosylation of aromatic amines I: Characterization of reaction products and kinetic scheme. *AAPS PharmSciTech* 2009;10:317–28.
 31. Fuller TW, Acharya AP, Meyyappan T, Yu M, Bhaskar G, Little SR, et al. Comparison of bladder carcinogens in the urine of e-cigarette users versus non e-cigarette using controls. *Sci Rep* 2018;8:507.
 32. Janzantti NS, Macoris MS, Garruti DS, Monteiro M. Influence of the cultivation system in the aroma of the volatile compounds and total antioxidant activity of passion fruit. *LWT - Food Sci Technol* 2012;46:511–8.
 33. Silano V, Bolognesi C, Castle L, Chipman K, Cravedi JP, Engel KH, et al. Safety of ethyl acrylate to be used as flavouring. *EFSA J* 2017;15:5012.
 34. Burdock GA. *Encyclopedia of food & color additives*: New York: CRC Press;2014.
 35. Lim H-H, Shin H-S. Determination of volatile organic compounds including alcohols in refill fluids and cartridges of electronic cigarettes by headspace solid-phase micro extraction and gas chromatography–mass spectrometry. *Anal Bioanal Chem* 2017;409:1247–56.
 36. Wagener TL, Floyd EL, Stepanov I, Driskill LM, Frank SG, Meier E, et al. Have combustible cigarettes met their match? The nicotine delivery profiles and harmful constituent exposures of second-generation and third-generation electronic cigarette users. *Tob Control* 2017;26:e23–e28.
 37. Yu Z, Xu L, Wei Y, Wang Y, He Y, Xia Q, et al. A new route for the synthesis of propylene oxide from bio-glycerol derivated propylene glycol. *Chem Commun* 2009;26:3934–6.
 38. Havel CM, Benowitz NL, Jacob P III, St.Helen G. An electronic cigarette vaping machine for the characterization of aerosol delivery and composition. *Nicotine Tob Res* 2017;19:1224–31.
 39. Conklin DJ, Ogunwale MA, Chen Y, Theis WS, Nantz MH, Fu X-A, et al. Electronic cigarette-generated aldehydes: the contribution of e-liquid components to their formation and the use of urinary aldehyde metabolites as biomarkers of exposure. *Aerosol Sci Technol* 2018;52:1219–32.
 40. Ogunwale MA, Li M, Ramakrishnam Raju MV, Chen Y, Nantz MH, Conklin DJ, et al. Aldehyde detection in electronic cigarette aerosols. *ACS Omega* 2017;2:1207–14.
 41. Abraham K, Andres S, Palavinskas R, Berg K, Appel KE, Lampen A. Toxicology and risk assessment of acrolein in food. *Mol Nutr Food Res* 2011;55:1277–90.
 42. St.Helen G, Benowitz NL, Ko J, Jacob P III, Gregorich SE, Pérez-Stable EJ, et al. Differences in exposure to toxic and/or carcinogenic volatile organic compounds between Black and White cigarette smokers. *J Expo Sci Environ Epidemiol* 2019 Aug 12 [Epub ahead or print].

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Comparison of Systemic Exposure to Toxic and/or Carcinogenic Volatile Organic Compounds (VOC) during Vaping, Smoking, and Abstinence

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