

Genome-wide Association Analysis of Proinflammatory Cytokines and Gene–lifestyle Interaction for Invasive Breast Cancer Risk: The WHI dbGaP Study

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ABSTRACT

Immune-related etiologic pathways to influence invasive breast cancer risk may interact with lifestyle factors, but the interrelated molecular genetic pathways are incompletely characterized. We used data from the Women's Health Initiative Database for Genotypes and Phenotypes Study including 16,088 postmenopausal women, a population highly susceptible to inflammation, obesity, and increased risk for breast cancer. With 21,784,812 common autosomal single-nucleotide polymorphisms (SNP), we conducted a genome-wide association (GWA) gene–environment interaction ($G \times E$) analysis in six independent GWA Studies for proinflammatory cytokines [IL6 and C-reactive protein (CRP)] and their gene–lifestyle interactions. Subsequently, we tested for the association of the GWA SNPs with breast cancer risk. In women overall and stratified by obesity status (body mass index, waist circumference, and waist-to-hip ratio) and obesity-related lifestyle factors (exercise and high-fat diet), 88 GWA SNPs in 10 loci were associated with proinflammatory cytokines: 3 associated with IL6 (1 index

SNP in *MAPK1* and 1 independent SNP in *DECI*); 85 with CRP (3 index SNPs in *CRP1*, *CRP*, *RP11-419N10.5*, *HNF1A-AS1*, *HNF1A*, and *C1q2orf43*; and two independent SNPs in *APOE* and *APOC1*). Of those, 27 in *HNF1A-AS1*, *HNF1A*, and *C1q2orf43* displayed significantly increased risk for breast cancer. We found a number of novel top markers for CRP and IL6, which interacted with obesity factors. A substantial proportion of those SNPs' susceptibility influenced breast cancer risk. Our findings may contribute to better understanding of genetic associations between pro-inflammation and cancer and suggest intervention strategies for women who carry the risk genotypes, reducing breast cancer risk.

Prevention Relevance: The top GWA-SNPs associated with pro-inflammatory biomarkers have implications for breast carcinogenesis by interacting with obesity factors. Our findings may suggest interventions for women who carry the inflammatory-risk genotypes to reduce breast cancer risk.

Introduction

Chronic inflammation may play an important role in the pathogenesis of noninflammatory diseases, including specific types of cancers such as colorectal, liver, and breast cancers, from tumor initiation through progression (1, 2). Activation of innate immunity involves a number of inflammatory cells, growth and transcription factors, chemokines, and proinflammatory mediators, creating a tissue microenvironment high in reactive oxygen and nitrogen species, leading to potential DNA alterations in nearby cells (3). The inflammatory response also elevates the circulating levels of cancer-promoting inflammatory cytokines such as C-reactive protein (CRP) and IL6 (2). These key biomarkers reflect different molecular pathways in the immune cascade in acute and chronic immune responses but may be inter-related in carcinogenesis. For example, IL6, upregulated by macrophages and adipose tissue, has promoted breast tumor initiation and progression (4). CRP, a major acute-phase reactant and a biomarker of chronic low-grade inflammation, partially induced by IL6, has been associated with increased risk of breast cancer (5). The molecular mechanisms of these markers in carcinogenesis have not been confirmed and are

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partially understood. IL6 regulates estrogen synthesis and aromatase activity responsible for estrogen production in adipose tissue, which is important in postmenopausal breast cancer (6). CRP levels are reduced when COX-2 action (promoting estrogen formation in adipose tissue) is inhibited (7). Thus, IL6 and CRP may provide a link between inflammatory pathways and breast cancer tumorigenesis.

The heritability of CRP and IL6 levels was 25%–40% (8, 9) and 60% (10), respectively, in Europeans on the basis of family and twin studies. Previous genome-wide association studies (GWAS) explain about 5% (11) and <2% (12) of interindividual variability of CRP and IL6 levels, respectively, suggesting that additional genetic contributions to CRP and IL6 concentrations remain largely undetermined. CRP and IL6 levels are also determined by obesity (overall and central; refs. 11–16), lipid metabolism (15), and obesity-related lifestyle factors such as exercise, high-fat diet, smoking, and alcohol (12, 17, 18). Thus, studying whether these lifestyle factors interact with genetic markers to influence CRP and IL6 concentrations may uncover the complicated genotype–phenotype pathway.

Few GWA gene–environment interaction ($G \times E$) studies have examined those pleiotropic cytokines' gene–phenotype relationship with obesity factors. Particularly, genomic studies for CRP have investigated obesity-specific stratifications among GWA-based single-nucleotide polymorphisms (SNP) only (19, 20) or tested the $G \times E$ effect by introducing the interaction term into the gene–phenotype association (11). A full genome-wide scan within obesity strata at the initial stage throughout the GWA analysis may help to provide a more profound molecular basis for gene–phenotype pathways that are influenced by environment. In addition, a GWA analysis examining inflammatory markers as binary outcomes (e.g., normal range vs. chronic low-grade inflammation) rather than examining as a continuous variable, could address a nonlinearity issue that has frequently been violated in traditional linear regression and further identify SNPs whose effect reflects an allele-based risk magnitude of chronic low-grade inflammation.

Immune-related etiologic pathways that influence breast cancer may differ by menopausal status, probably owing to the role of sex hormones in mediating the innate and adaptive immune systems (21). Furthermore, in postmenopausal women, inflammatory cytokines and the genetic markers have displayed different associations with breast cancer according to obesity-related factors such as obesity status (overall and visceral; ref. 22), physical activity, and dyslipidemia (23, 24). Thus, our $G \times E$ study has focused on the risk for breast cancer among postmenopausal women, a population vulnerable to a high incidence of inflammation (25), obesity, and breast cancer (e.g., 80% of new cases occur in women age 50 years and older; ref. 26).

In this study, we hoped to identify SNPs explaining additional interindividual variability in cancer-promoting inflammatory biomarkers, including CRP and IL6. We conducted a GWA $G \times E$ study to characterize the genetic architecture of the biomarkers that interact with obesity factors. Next, we evaluated whether the identified SNPs in particular behavioral settings are

associated with breast cancer risk in the identical behavioral setting. This may avoid bias derived from a different population structure by examining an identical genomic structure of the population for association with inflammatory cytokines and cancer risk simultaneously. We tested an empirical hypothesis that a substantial proportion of the susceptibility of GWA-based SNPs in CRP and IL6 affects breast cancer risk and that obesity lifestyle factors modify the relationship (Supplementary Fig. S1).

Materials and Methods

Study population

Our study included postmenopausal women who enrolled in the Women's Health Initiative (WHI) Harmonized and Imputed GWASs coordinated by the database of Genotypes and Phenotypes (dbGaP). These studies encompass the WHI Observational Studies and Clinical Trials to contribute a joint imputation and harmonization effort to the GWASs. Detailed rationale and study design have been discussed elsewhere (27, 28). The WHI study included women enrolled between 1993 and 1998 at 40 clinical centers across the United States. Eligible women were 50–79 years old, postmenopausal, expected to reside near the clinical centers for at least 3 years after enrollment, and able to provide written informed consent. The Harmonized and Imputed studies involved 6 GWASs (Table 1). Of the 16,088 who reported their race or ethnicity as non-Hispanic white (Supplementary Fig. S2), we excluded 2,714 who had been diagnosed with diabetes at or after enrollment. We also excluded 1,301 whose genetic information was found to be duplicated in the 6 GWASs and/or those with first- and second-degree relatives. In addition, we excluded 1,275 women whose genetic data did not pass the quality assurance (QA) test [outliers based on Principal Components (PCs)], leaving 10,798 (90% of the eligible 12,073) for our GWA $G \times E$ analysis. For the association with breast cancer, we next excluded 619 women who had been followed up for < 1 year and/or had been diagnosed with any type of cancer at enrollment, leaving a total of 10,179 women (94% of the 10,798 GWA participants). These women had been followed up through August 29, 2014, with a mean of 16 years follow-up, and 537 of them had developed invasive breast cancer. The Institutional Review Boards of each WHI participating clinical center and the University of California, Los Angeles, approved this study.

Data collection

The WHI coordinating center collected information by using standardized written protocols and conducted data QA with periodic visits. Participants had completed at enrollment self-administered questionnaires on the following information (listed for only those selected for our study): demographic and socioeconomic (age, education, race/ethnicity, family income, and family history of breast cancer), lifestyle (depressive symptom, smoking, and exercise), dietary [dietary alcohol intake in g/day and % calories from saturated fatty acids (SFA)/day] factors, and reproductive histories (history of hysterectomy, ages at menarche and menopause, oral contraceptive use, and

Table 1. Distributions of proinflammatory phenotypes in six GWASs ($N = 10,798$).

Study ^a	N	Phenotype	
		IL6	CRP
Circulating plasma level			
		pg/mL, mean (SD)	mg/L, mean (SD)
AS264	1,603	4.478 (2.22)	3.883 (1.74)
GARNET	2,382	4.380 (3.16)	4.189 (7.50)
GECCO-CYTO	1,177	4.315 (3.04)	3.911 (3.81)
GECCO-INIT	216	3.221 (4.35)	4.323 (5.33)
HIPFX	1,909	4.548 (7.68)	3.752 (3.62)
WHIMS	3,511	4.472 (3.26)	3.360 (5.42)
Binary analysis			
		<4.4 pg/mL/≥4.4 pg/mL^b n (%) / n (%)	≤3.0 mg/L/>3.0 mg/L^b n (%) / n (%)
AS264	1,603	1,195 (74.5)/408 (25.5)	517 (32.3)/1,086 (67.7)
GARNET	2,382	1,748 (73.4)/634 (26.6)	1,446 (60.7)/936 (39.3)
GECCO-CYTO	1,177	901 (76.6)/276 (23.4)	497 (42.2)/680 (57.8)
GECCO-INIT	216	187 (86.6)/29 (13.4)	124 (57.4)/92 (42.6)
HIPFX	1,909	1,436 (75.2)/473 (24.8)	813 (42.6)/1,096 (57.4)
WHIMS	3,511	2,649 (75.4)/862 (24.6)	2,325 (66.2)/1,186 (33.8)

^aGenotyping was run on several platforms: AS264 via Affymetrix Gene Titan, Axiom Genome-Wide Human CEU I Array Plate; GARNET via Illumina HumanOmni1-Quad v1-0 B; GECCO via Illumina 610 and Cytochip 370K; HIPFX via Illumina 550K and 610K; and WHIMS via HumanOmniExpress Exome-8v1_B.

^bEach phenotype was categorized via the corresponding cut-off value (4.4 pg/mL for IL6 and 3.0 mg/L for CRP); higher blood level than the threshold to be considered either the fourth quartile of the participants ($\geq 75\%$, 4.4 pg/mL for IL6) or high immune response and chronic low-grade inflammation (> 3.0 mg/L for CRP; refs. 39, 40).

2 types of exogenous estrogen use: unopposed estrogen-only and opposed estrogen plus progestin). Anthropometric data, including height, weight, and waist and hip circumferences, were measured at baseline by trained staff. Those 17 variables (Table 2) were initially selected on the basis of their association with inflammation and breast cancer among postmenopausal women through the literature review (23, 29, 30) and were confirmed by univariate, stepwise, and multicollinearity regression testing to be analyzed in our study.

Breast cancer development was our cancer outcome of interest. The time between enrollment and breast cancer development, censoring, or study endpoint was measured as the number of days and then converted into years. Breast cancer diagnosis was determined using a centralized review of medical charts by a committee of physicians on the basis of pathology or cytology reports. Cancer cases were coded according to the National Cancer Institute's Surveillance, Epidemiology, and End-Results guidelines (31).

Genotyping and laboratory methods

Genotyped data generated on several different platforms were extracted from the WHI dbGaP Harmonized and Imputed GWASs (Table 1). The genotypes were normalized to the reference panel GRCh37 and genotype imputation was performed via 1,000 genomes reference panels. (28) SNPs were checked for harmonization with pairwise concordance among all samples across the 6 GWASs. We crosschecked the self-reported race/ethnicity with PCs. If any discrepancy or admixed participants were detected, an additional analysis was performed with a follow-up demographic questionnaire (32). In the initial quality control (QC) process, SNPs were filtered with a

missing-call rate of $< 2\%$ and a Hardy-Weinberg Equilibrium of $P \geq 1E-04$. In the secondary QC, we retained SNPs with $\hat{R}^2 \geq 0.6$ imputation quality (33). Within high-quality SNPs that included only HapMap3 SNPs with $\hat{R}^2 \geq 0.9$ (34), we estimated relatedness between samples. To reduce potential confounding due to shared environment, we excluded individuals with a kinship estimate > 0.088 on the basis of the KING robust kinship estimator (35). We next computed 10 PCs in each GWAS using the same set of high-quality SNPs, but without relatedness, within the linkage disequilibrium (LD) and excluded outlier samples that fell outside of 6 SD on the basis of Mahalanobis distances. Finally, a total of 21,784,812 autosomal SNPs from 10,798 individuals were examined in this study.

Participants' fasting blood samples at enrollment were drawn by trained phlebotomists. Serum concentrations of IL6 and CRP were measured by using, respectively, the Quantitative Sandwich Enzyme Immunoassay technique (Quantikine HS Immunoassay Kit; R&D Systems, Inc.) and a high sensitivity immunoturbidimetric assay (Kimiya Biomedical Company) read with a Roche analyzer (Roche Diagnostics), with the median interassay coefficients of variation of 12.4% and 2.3%, respectively. About 30% of the phenotypes were replaced by imputation values using an unsupervised splitting of Random Survival Forest imputation (<https://github.com/ehrlinger/randomForestSRC/blob/master/R/impute.rsrc.R>; ref. 36). In the overall and the obesity-specific strata of the GWA analysis, sensitivity testing was performed with and without imputed values for IL6 and CRP, producing regression estimates, genomic control inflation factors (λ_{GC}), Q-Q plots, and Manhattan plots; no significant differences were observed.

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Table 2. Participants' characteristics stratified by breast cancer.

Characteristics	Participants without breast cancer	Participants with breast cancer	P
	(n = 9,642) n (%)	(n = 537) n (%)	
Age in years, median (range)	67 (50–81)	67 (50–79)	0.085
Education			0.011
≤High school	3,476 (36.1)	164 (30.5) ^a	
>High school	6,166 (63.9)	373 (69.5)	
Family income			0.002
<\$35,000	4,344 (46.1)	207 (39.2) ^a	
≥\$35,000	5,088 (53.9)	321 (60.8)	
Family history of breast cancer			0.032
No	7,838 (81.3)	416 (77.5) ^a	
Yes	1,804 (18.7)	121 (22.5)	
BMI in kg/m ² , median (range)	26.84 (16.83–58.49)	28.23 (17.55–49.31) ^a	<0.001
BMI ^b			<0.001
<30.0 kg/m ²	6,859 (71.1)	320 (59.6) ^a	
≥30.0 kg/m ²	2,783 (28.9)	217 (40.4)	
WHR, median (range)	0.807 (0.444–1.393)	0.813 (0.640–1.263) ^a	0.013
WHR ^b			0.011
≤0.85	6,895 (71.5)	356 (66.3) ^a	
>0.85	2,747 (28.5)	181 (33.7)	
Waist in cm, median (range)	85.00 (62.00–125.00)	88.50 (63.50–125.00) ^a	<0.001
Waist ^b			<0.001
≤88 cm	5,756 (59.7)	268 (49.9) ^a	
>88 cm	3,886 (40.3)	269 (50.1)	
METS-hour/week ^c	7.25 (0.00–134.17)	6.75 (0.00–81.67)	0.487
METS-hour/week ^c			0.844
≥10.0	4,001 (41.5)	220 (41.0)	
<10.0	5,641 (58.5)	317 (59.0)	
How many cigarettes per day			<0.001
<15 cigarettes	5,432 (56.3)	250 (46.6) ^a	
≥15 cigarettes	4,210 (43.7)	287 (53.4)	
Depressive symptoms ^d , median (range)	0.002 (0.0004–0.937)	0.002 (0.0005–0.880)	0.836
Dietary alcohol per day in g, median (range)	1.04 (0.00–183.76)	1.86 (0.00–127.15) ^a	0.001
% calories from SFA, median (range)	11.33 (2.22–32.39)	11.46 (3.73–21.50)	0.967
% calories from SFA ^e			0.297
<9.0%	2,174 (22.5)	132 (24.6)	
≥9.0%	7,468 (77.5)	405 (75.4)	
Age at menarche in years, median (range)	13 (≤ 9–≥ 17)	12 (≤9–≥17) ^a	0.002
Hysterectomy ever			0.004
No	6,143 (63.7)	376 (70.0) ^a	
Yes	3,499 (36.3)	161 (30.0)	
Age at menopause in years, median (range)	50 (20–60)	50 (21–63)	0.050
Oral contraceptive duration in years, median (range)	5.66 (0.08–47.00)	5.18 (0.08–21.00) ^a	<0.001
Exogenous estrogen use (E only use)			0.001
Never use	6,697 (69.5)	411 (76.5) ^a	
<5 years	1,361 (14.1)	51 (9.5)	
5 to <10 years	516 (5.4)	17 (3.2)	
10 + years	1,068 (11.1)	58 (10.8)	
Exogenous estrogen use (E + P use)			0.001
Never use	7,940 (82.3)	412 (76.7) ^a	
<5 years	927 (9.6)	64 (11.9)	
5 to <10 years	406 (4.2)	30 (5.6)	
10 + Years	369 (3.8)	31 (5.8)	

Abbreviations: E, estrogen; E+P, estrogen plus progestin.

^aP < 0.05, χ^2 or Wilcoxon rank-sum test.^bBMI, WHR, and WST were categorized using 30 kg/m², 0.85, and 88 cm, respectively, where cut-off levels or higher fall within the overall or visceral obese range (<https://www.cdc.gov/obesity/adult/defining.html>).^cPhysical activity was estimated from recreational physical activity combining walking and mild, moderate, and strenuous physical activity. Each activity was assigned a MET value corresponding to intensity; the total MET-hours/week was calculated by multiplying the MET level for the activity by the hours exercised per week and summing the values for all activities. The total MET was stratified into two groups, with 10 METs as the cutoff according to current American College of Sports Medicine and American Heart Association recommendations (63).^dDepression scales were estimated via a short form of the Center for Epidemiologic Studies Depression Scale.^ePercent calories from SFA was classified by 9%, addressing low sample power (i.e., containing a quarter in one side) and adherent to the American Heart Association and American College of Cardiology dietary guidelines, which are aligned with the 2015–2020 Dietary Guidelines for Americans to help cardiovascular and metabolic diseases reductions (64).

Statistical analysis

Participants' baseline characteristics by breast cancer development were examined by unpaired two-sample *t* tests for continuous variables and χ^2 tests for categorical variables; if continuous variables were skewed or had outliers, Wilcoxon rank-sum test was used. A GWA analysis in each study was conducted via multiple linear and logistic regressions, adjusting for age and 10 PCs to produce effect sizes and ORs, respectively, and 95% confidence intervals (CI) of IL6 and CRP concentrations, with additive and minor allele-dominant and -recessive genetic models. In this study, we present the results from only the logistic regressions due to the high genomic control inflation factors ($\lambda_{GC} > 1.0$) from the linear regressions. Multiple testing was corrected by adjusting *P* values to the genome-wide significance level ($P < 5E-08$). The inverse variance-weighted fixed-effects meta-analysis was conducted to combine the findings across the 6 GWASs. Heterogeneity among studies was tested via Cochran Q statistic (37) at $P < 8E-03$ after Bonferroni corrections.

For all the SNPs at the initial stage of analysis, gene-obesity interactions were examined in each study (i) by introducing an interaction term into a regression model, adjusting for 10 PCs, and (ii) within the obesity strata defined by body mass index (BMI; cutoff, 30 kg/m²), waist circumference (WST; cutoff, 88 cm), waist-to-hip ratio (WHR; cutoff, 0.85), metabolic equivalents (MET; cutoff, 10-hours/week), and % calories from SFA (cutoff, 9%). Meta-analysis was performed assuming fixed effects to combine the interaction results across the 6 GWASs. Multiple comparisons were adjusted by the Benjamini-Hochberg method (38). The LDs between the top GWA signals were computed, and regional plots were created using LOCUS-ZOOM (<http://locuszoom.org/>). Gene enrichment and functional annotation analyses were performed using DAVID v6.8 (<https://david.ncifcrf.gov/>).

With the top signals, we further conducted a multiple Cox proportional hazards regression in the combined 6 GWASs to obtain HRs and 95% CIs predicting a risk for breast cancer by adjusting for 17 confounding factors (Table 2). The proportional hazards assumptions were tested by a Schoenfeld residual plot and rho. Given that the SNP-cancer pathway was a hypothesis-driven question, the *P* value was not subject to multiple testing corrections; thus, a two-tailed $P < 0.05$ was considered statistically significant. During the data QC process, dose2plink script (<http://genepi.qimr.edu.au/staff/sarahMe/dose2plink>), PLINK1.9/2.0, KING kinship estimator, and R *stats* package on UCLA's Hoffman2 high performance computing cluster were used. For GWA, phenotype imputation, and SNP-cancer association tests, PLINK1.9/2.0 (glm/interaction/meta-analysis) and R3.5.1. (qqman/manhattan, randomForestSRC, and survival packages) were used.

Results

Distributions of the pro-inflammatory cytokines (CRP and IL6) in the 6 GWASs are displayed in Table 1. Blood-level thresholds were determined as reflecting either the chronic

low-grade inflammation status (>3.0 mg/L for CRP) or the fourth quartile of the participants (≥ 4.4 pg/mL for IL6; refs. 39, 40). Allele frequencies in each study (Supplementary Table S1) were cross-checked with those in Europeans in the 1000 Genomes Project Ensembl GRCh37 (http://grch37.ensembl.org/Homo_sapiens/Info/Index). Participants' baseline characteristics by breast cancer development are shown in Table 2. Women who developed breast cancer were more likely to have greater incomes and a family history of breast cancer, be obese overall and viscerally, smoke ≥ 15 cigarettes/day, consume more dietary alcohol/day, have experienced menarche at earlier age, have undergone hysterectomy less frequently, and have shorter durations of oral contraceptive and E-only use but longer durations of estrogen + progestin use.

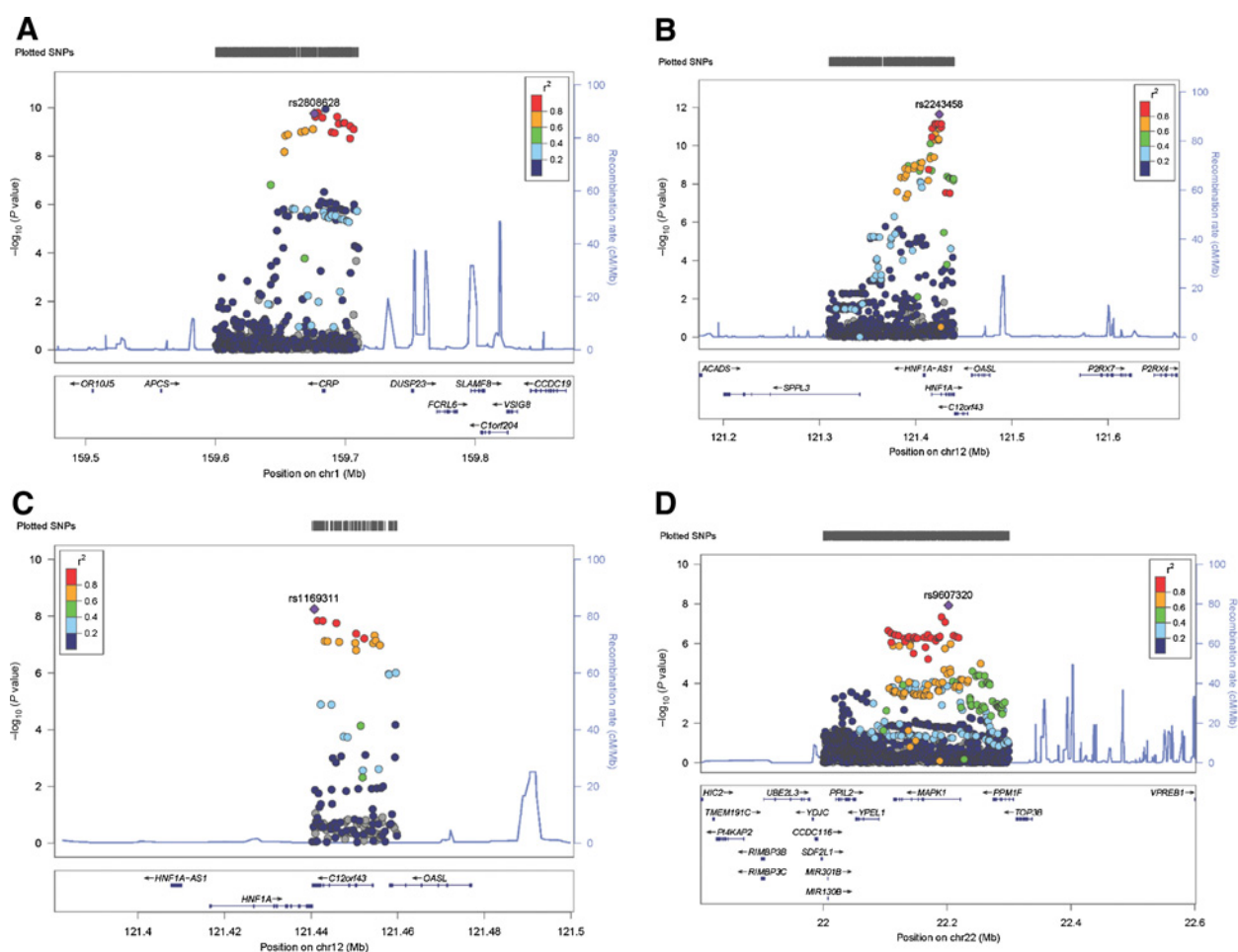
Of 21,784,812 common autosomal SNPs, 88 SNPs reached genome-wide significance (including 4 index SNPs and 3 SNPs, independent of each other) in 10 loci, 7 of which were novel, and 3 loci (SNPs near *CRP*, *HNFI1A*, and *APOE*) that were previously observed. The genomic control inflation factor (λ_{GC}) was 1.0, indicating no evidence of type 1 error inflation. Those top genetic markers did not overlap between the two cytokine phenotypes. Identified loci explained 5.8% and 0.7% of interindividual variability of CPR and IL6, respectively.

CRP: GWAS G \times E results and association with breast cancer

In the overall analysis, 82 top signals were detected in 7 loci of *CRP1*, *CRP*, *HNFI1A-AS1*, *HNFI1A*, *C12orf43*, *APOE*, and *APOC1* (Supplementary Table S2; Supplementary Fig. S3A-S3H). Specifically, 8 SNPs near *CRP1*, 1 near *CRP1/CRP*, and 11 in intron, 3'-UTR, or 3' flanking regions of *CRP* (index: rs2808628 with $r^2 > 0.7$, Fig. 1A) were associated with increased risk of chronic inflammation (approximately 20%). On the contrary, the following loci at 12q and 19q displayed about 20% of decreased risk: 23 in an intronic region of *HNFI1A-AS1* and 31 in intronic or 3'-UTR regions of *HNFI1A* (index: rs2243458 with $r^2 > 0.7$, Fig. 1B), 6 in intronic or 3'-UTR regions of *C12orf43* (index: rs1169311 with $r^2 > 0.9$; Fig. 1C), *APOE* rs429358, and *APOC1* rs5117.

When stratified by BMI (Supplementary Table S3), a number of SNPs near *HNFI1A-AS1* and *HNFI1A* (18 and 27 SNPs, respectively, with index: rs2243458) and one (rs1169311) in *C12orf43* were detected; these partially overlapped those identified from the overall analysis, with about 20% of decreased risk for chronic inflammation, only in the nonobese (BMI < 30) subgroup. In the obese subgroup (BMI ≥ 30), 3 novel SNPs in *CRP/RP11-419N10.5* in LD ($r^2 > 0.9$) were identified with 70% of increased risk for chronic inflammation. When stratified by WHR or WST, similar patterns were shown for SNPs on inflammation: SNPs near *CRP* and *CRP/RP11-419N10.5* with 35%–50% of increased risk in the viscerally obese subgroup only. SNPs near *HNFI1A* were shown with decreased risk for inflammation, also only in the viscerally obese subgroup (Supplementary Table S3).

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**Figure 1.**

Regional plots for the SNP association [Note: LD (r^2) shown by color intensity gradient]. **A**, Twenty SNPs nearby *CRP* and *CRP1* ($r^2 > 0.7$) with *CRP*. **B**, Fifty-four SNPs nearby *HNF1A-AS1* and *HNF1A* ($r^2 > 0.7$) with *CRP*. **C**, Six SNPs nearby *CI2orf43* ($r^2 > 0.9$) with *CRP*. **D**, Two SNPs nearby *MAPK1* ($r^2 > 0.8$) with *IL6*.

Table 3. Genome-wide associated SNPs with *CRP* and their associations with breast cancer risk in overall analysis.

SNP	Chr	Position ^b	Allele ^a (Ref/Alt)	OR ^c	P	Q	HR ^d (95% CI)	P
<i>HNF1A</i> rs2259852	12	121434833	A/G	0.84	4.95E-09	0.347	1.13 (1.00–1.27)	0.058
<i>HNF1A</i> rs2464195	12	121435475	A/G	0.84	4.96E-09	0.358	1.13 (1.00–1.27)	0.057
<i>HNF1A</i> rs2259816	12	121435587	T/G	0.84	5.16E-09	0.360	1.13 (1.00–1.28)	0.054
<i>HNF1A</i> rs1169306	12	121438311	T/C	0.84	5.13E-09	0.371	1.13 (1.00–1.27)	0.058
<i>HNF1A</i> rs735396	12	121438844	C/T	0.84	5.25E-09	0.375	1.13 (1.00–1.27)	0.058
<i>HNF1A</i> rs1169309	12	121439192	T/G	0.84	6.61E-09	0.356	1.13 (1.00–1.27)	0.057
<i>HNF1A</i> rs1169310	12	121439433	A/G	0.84	5.28E-09	0.376	1.13 (1.00–1.27)	0.058
<i>CI2orf43</i> rs1169311	12	121440731	T/C	0.84	5.64E-09	0.375	1.13 (1.00–1.27)	0.057

Note: Only SNPs significant on genome-wide level included ($N = 10,179$).

Abbreviations: Alt, alternative; Chr, chromosome; CI, confidence interval; Q, Cochran Q; Ref, reference.

^aAdditive genetic model regressed.

^bGRCh 37 coordinated.

^cGWA analysis for *CRP* adjusted for age and 10 PCs.

^dHR adjusted by age, education, annual family income, family history of breast cancer, BMI, waist-to-hip ratio, physical activity, depressive symptoms, How many cigarettes per day, dietary alcohol in g/day, % calories from SFA/day, hysterectomy, ages at menarche and menopause, oral contraceptive use, and exogenous estrogen only-use and plus progestin use.

Table 4. Genome-wide associated SNPs with CRP and their relationships with breast cancer risk in obesity-stratified (BMI and WST) analysis.

SNP	Chr	Position ^a	Allele ^b (Ref/Alt)	Non-obese group			Obese group			P
				OR ^c	P ^d	Q	HR ^e (95% CI)	P	Q	
Non overall-obese group, BMI < 30 (n = 7,179)										
HNF1A-AS1 rs11065365	12	121392040	G/A	0.79	1.42E-08	0.209	0.92 (0.76-1.10)	0.360	0.76	<Overall obese group, BMI ≥ 30 (n = 3,000)
HNF1A-AS1 rs7953249	12	121403724	G/A	0.81	6.45E-09	0.154	0.94 (0.80-1.10)	0.432	0.78	2.75E-05
HNF1A-AS1 rs7135337	12	121404155	A/C	0.80	3.19E-09	0.150	0.95 (0.81-1.11)	0.505	0.77	7.58E-05
HNF1A-AS1 rs1920792	12	121404584	C/T	1.22	1.56E-08	0.062	1.06 (0.91-1.24)	0.451	1.28	4.75E-05
HNF1A-AS1 rs10774579	12	121405210	C/T	1.22	1.72E-08	0.062	1.06 (0.91-1.24)	0.457	1.28	0.0001
HNF1A-AS1 rs2393792	12	121406293	A/G	1.22	2.96E-08	0.081	1.04 (0.89-1.22)	0.605	1.27	0.0002
HNF1A-AS1 rs142632970	12	121413345	G/A	0.80	8.81E-09	0.805	0.96 (0.81-1.14)	0.633	0.80	0.001
HNF1A-AS1 rs7139079	12	121415293	G/A	0.80	1.37E-09	0.140	0.95 (0.81-1.12)	0.566	0.75	5.26E-06
HNF1A-AS1 rs2464190	12	121415390	G/C	0.80	3.22E-09	0.163	0.94 (0.80-1.10)	0.441	0.75	1.02E-05
HNF1A rs1169289	12	121416622	G/C	0.80	1.68E-08	0.156	0.96 (0.81-1.12)	0.580	0.77	3.62E-05
HNF1A rs169473	12	121420260	A/G	0.80	7.62E-10	0.123	1.01 (0.86-1.18)	0.944	0.74	3.95E-06
HNF1A rs7979478	12	121420263	A/G	0.79	1.17E-10	0.167	0.99 (0.84-1.16)	0.864	0.75	1.15E-05
HNF1A rs7970695	12	121423376	G/A	0.79	5.40E-10	0.139	1.01 (0.86-1.18)	0.928	0.75	4.96E-06
HNF1A rs9738226	12	121423659	A/G	0.79	5.94E-10	0.130	1.01 (0.86-1.19)	0.902	0.75	4.96E-06
HNF1A rs2393791	12	121423956	C/T	0.79	5.16E-10	0.139	1.01 (0.86-1.18)	0.916	0.75	4.80E-06
HNF1A rs2393776	12	121424406	G/A	0.79	3.23E-10	0.218	1.01 (0.86-1.18)	0.918	0.74	2.31E-06
HNF1A rs2393775	12	121424574	G/A	0.79	3.25E-10	0.225	1.01 (0.86-1.18)	0.915	0.74	2.30E-06
HNF1A rs7310409	12	121424861	A/G	0.79	3.30E-10	0.225	1.01 (0.86-1.18)	0.915	0.74	2.31E-06
HNF1A rs2264782	12	121432603	T/C	0.82	4.64E-08	0.241	1.06 (0.90-1.24)	0.512	0.76	1.50E-05
HNF1A rs2259852	12	121434833	A/G	0.81	2.91E-08	0.273	1.05 (0.90-1.24)	0.519	0.76	2.45E-05
HNF1A rs2464195	12	121435475	A/G	0.81	2.82E-08	0.283	1.05 (0.90-1.24)	0.517	0.76	2.51E-05
HNF1A rs2259816	12	121435587	T/G	0.81	3.05E-08	0.281	1.06 (0.90-1.24)	0.498	0.76	2.54E-05
HNF1A rs1169306	12	121438311	T/C	0.81	2.81E-08	0.305	1.05 (0.90-1.24)	0.522	0.76	2.54E-05
HNF1A rs735396	12	121438844	C/T	0.81	2.83E-08	0.309	1.05 (0.90-1.24)	0.520	0.76	2.54E-05
HNF1A rs1169309	12	121439192	T/G	0.81	3.11E-08	0.294	1.05 (0.90-1.24)	0.513	0.77	2.97E-05
HNF1A rs1169310	12	121439433	A/G	0.81	2.85E-08	0.311	1.05 (0.90-1.24)	0.520	0.76	2.54E-05
C12orf43 rs1169311	12	121440731	T/C	0.81	2.98E-08	0.312	1.05 (0.90-1.24)	0.514	0.76	2.60E-05
Non visceral-obese group, WST ≤ 88 (n = 6,024)										
HNF1A rs2393776	12	121424406	G/A	0.82	1.25E-06	0.168	0.97 (0.81-1.15)	0.708	0.75	Visceral obese group, WST > 88 (n = 4,155)
HNF1A rs2393775	12	121424574	G/A	0.82	1.27E-06	0.173	0.97 (0.81-1.15)	0.711	0.75	3.47E-08
HNF1A rs7310409	12	121424861	A/G	0.82	1.28E-06	0.172	0.97 (0.81-1.15)	0.711	0.75	3.42E-08
										0.530
										0.528
										0.530
										1.23 (1.04-1.46)
										1.23 (1.04-1.46)
										1.23 (1.04-1.47)

Note: Only SNPs significant on genome-wide level included. Numbers in bold face are statistically significant.

Abbreviations: Alt, alternative; Chr, chromosome; CI, confidence interval; G, Cochran Q; Ref, reference.

^aGRCh 37 coordinated.

^bAdditive genetic model regressed.

^cGWA analysis for CRP adjusted for age and 10 PCs.

^dP values were adjusted to correct for multiple testing via the Benjamini-Hochberg approach.

^eHR adjusted by age, education, annual family income, family history of breast cancer, body mass index (not in BMI-stratified), WHR (not in WST-stratified), physical activity, depressive symptoms, how many cigarettes per day, dietary alcohol in g/day, % calories from SFA/day, hysterectomy, ages at menarche and menopause, oral contraceptive use, and exogenous estrogen only-use and plus progestin use.

In relation to breast cancer risk with the top signals, the loci at 12q, including *HNF1A-ASI*, *HNF1A*, and *C12orf43* were associated, while the loci at 1q (*CRPP1*, *CRP*, and *RP11-419N10.5*) were not. In particular, CRP-decreasing SNPs near *HNF1A* in the overall analysis were associated with 13% increased risk for breast cancer in the identical overall group (Table 3). CRP-decreasing SNPs near *HNF1A-ASI*, *HNF1A*, and *C12orf43* in the nonobese (BMI < 30) subgroup displayed a slightly decreased risk without reaching statistical significance, but in the counterpart obese subgroup (BMI ≥ 30), approximately 25% increased risk for breast cancer was observed (Table 4). In addition, CRP-decreasing SNPs near *HNF1A* in the viscerally obese subgroup showed a 23% increased risk for breast cancer in the same viscerally obese subgroup (Table 4).

Similar genetic associations for the CRP phenotype were observed when stratified by physical activity and % calories from SFA (Table 5). SNPs near *HNF1A-ASI* and *HNF1A* (index: rs2393776 with $r^2 > 0.7$) were associated with 35% decreased risk for chronic inflammation (except for few SNPs *HNF1A-ASI* with increased risk) in the less-fat diet subgroup, whereas SNPs near *CRPP1* and *CRP* (index: rs2808628) were associated with 20% increased risk for chronic inflammation in the physically inactive and high-fat diet subgroups. No associations of those SNPs with breast cancer were detected (Supplementary Table S4).

IL6: GWAS G × E results and association with breast cancer

Three of the most significant signals were detected in two loci of *DECI* and *MAPK1* (Table 6; Supplementary Fig. S3I–S3L). In all participants and in the nonviscerally obese and high-fat diet subgroups, rs149109490 near *DECI* was strongly associated with increased IL6 levels, reflecting high risk for chronic low-grade inflammatory status (OR = 3.61; $P = 3.42E-08$; $Q = 0.078$). Two *MAPK1* SNPs (rs56398890 and rs9607320) in LD ($r^2 > 0.9$) displayed 25% decreased risk in the physically active subgroup only. None of the top SNPs were associated with breast cancer risk (Supplementary Table S5).

Gene-ontology annotation analyses (Supplementary Tables S6 and S7) indicated *CRP* involvement in the acute-phase inflammation response; *HNF1A*, *APOC1*, and *APOE* in glucose, lipoprotein, triglyceride, and cholesterol metabolisms; and *DECI* and *MAPK1* in the regulation of mitogen-activated protein kinase/PI3K/ERK 1/2 cascades, apoptotic process, cellular response to DNA damage stimulus, and cell differentiation and migration.

Discussion

A growing number of population-based genomic studies have emphasized the role of environmental factors in modifying gene-phenotype pathways. To our knowledge, this is the first study to characterize genetic determinants of inflammatory cytokines and subsequently their genomic associations with breast cancer development in an identical

postmenopausal population, at the genome-wide level, by incorporating obesity factors as an effect modifier. A number of novel top-GWA signals have been detected in relation to chronic low-grade inflammation on the basis of CRP and IL6 concentrations. Some of the top loci for their associations with chronic inflammation and breast cancer would not have been detected without the incorporation of the obesity factors.

Many genes annotated to the CRP-related variants mainly clustered in the innate and adaptive immune functions (*CRP*) or the glucose and lipid metabolisms (*HNF1A* and *APOC1*; refs. 11, 13). In particular, the *CRP* gene encodes a member protein of the pentaxin family involving host defense functions during the acute-phase response to tissue infection to promote phagocytosis and the complement system by interacting with the DNA in circulating cells (41). Several *CRP* and *CRPP1* SNPs detected in our study are located at 1q21 to 25 region, which contains genes encoding proteins with immune- and inflammation-associated functions (42). Furthermore, the CRP-related SNPs were mainly clustered in the 3'-UTL or 3' flanking region (e.g., rs1205) of the *CRP* gene involved in the post-transcriptional process by regulating mRNA stability, localization, and translational efficiency; thus, those SNPs in this region may affect CRP production (19). The *CRP* gene is a single-copy gene. Within about 16-kb upstream and downstream from the *CRP* gene, only one other sequence is *CRPP1*, a pseudogene, with 50%–80% homology to *CRP* (43). CRP-related SNPs near *CRPP1* have thus far been reported only in African Americans (14). We found a different set of *CRPP1* SNPs that were novel in non-Hispanic whites.

A positive reciprocal feedback role between CRP and adiposity pathophysiology has been suggested (15, 20), and CRP-related genes were correlated with BMI/WST/WHR-related genes (13). Consistent with previous findings (19, 44), we found that *CRP/CRPP1* SNPs were related to elevated CRP levels (i.e., greater risk for chronic inflammation) among those who were obese or had obesity lifestyles, indicating that obesity factors play a role in regulating the effect of *CRP/CRPP1* genes on CRP levels. Of note, the *CRP/CRPP1* gene-phenotype relationships were more profound in women with central obesity, suggesting an important role of adipose tissue distribution in CRP gene-phenotype pathways. Furthermore, the novel SNPs we found near *CRP/RP11-419N10.5* were associated with up to 70% elevated risk for chronic inflammation only in women with overall and central obesity, but the biological implications of those SNPs on *RP11-419N10.5*, a processed pseudogene, are unclear.

Hepatic nuclear factor 1 alpha (*HNF1A*) is a key regulator of CRP protein. It binds to promotor regions of the *CRP* gene to regulate CRP synthesis in liver cells (45). Thus, *HNF1A* is required for *CRP* gene expression (46). This implicates the functional role of *HNF1A* SNPs in altering the expression of its target gene, *CRP*; *HNF1A* mutations led to loss of function in reducing CRP levels (47). This provides a biologically plausible basis for our findings that SNPs in *HNF1A/HNF1A-ASI* were

Table 5. GWA analysis for the association with CRP, stratified by obesity-related lifestyle factors (physical activity and fat diet).

SNP	Chr	Position ^a	Allele ^b (Ref/Alt)	Interaction test		Active/less-fat diet group		Inactive/high-fat diet group	
				OR ^c	P	OR ^c	P ^d	OR ^c	P ^d
Interaction test for PA									
CRP1 rs2808628	1	159676011	G/A	0.97	0.354	1.16	0.002	1.26	9.13E-09
CRP1 rs2808629	1	159676796	G/A	0.97	0.368	1.16	0.002	1.26	1.28E-08
CRP1/CRP rs2794520	1	159678816	C/T	0.97	0.361	1.16	0.002	1.26	9.62E-09
CRP rs1205	1	159682233	C/T	0.97	0.335	1.16	0.002	1.26	1.03E-08
CRP rs3091244	1	159684665	G/A	1.03	0.354	0.85	0.001	0.80	1.01E-08
CRP rs1341665	1	159691559	G/A	0.97	0.377	1.15	0.003	1.25	4.85E-08
CRP rs2211320	1	159693605	G/A	0.98	0.450	1.17	0.001	1.26	2.24E-08
CRP rs7551731	1	159694779	T/C	0.97	0.322	1.15	0.003	1.26	1.57E-08
CRP rs7553007	1	159698549	G/A	0.97	0.350	1.16	0.003	1.26	1.86E-08
CRP rs4546916	1	159699249	G/T	0.97	0.354	1.16	0.003	1.26	1.87E-08
CRP rs4287174	1	159703442	T/A	0.97	0.344	1.16	0.003	1.26	2.06E-08
CRP rs12037186	1	159706230	A/G	0.97	0.332	1.15	0.004	1.26	2.01E-08
Interaction test for SFA									
CRP1 rs2592887	1	159652939	C/T	0.99	0.725	1.15	0.033	1.21	1.27E-08
CRP1 rs1470515	1	159653599	C/T	0.99	0.787	1.18	0.015	1.22	4.70E-09
CRP1 rs2592902	1	159655726	G/T	0.99	0.773	1.18	0.015	1.22	4.59E-09
CRP1 rs2808624	1	159665921	C/G	0.99	0.741	1.17	0.017	1.22	3.53E-09
CRP1 rs11265257	1	159668984	C/T	0.99	0.730	1.17	0.018	1.22	3.17E-09
CRP1 rs876537	1	159674933	C/T	0.99	0.730	1.17	0.017	1.22	2.73E-09
CRP1 rs2808628	1	159676011	G/A	1.01	0.854	1.24	0.002	1.23	4.53E-09
CRP1 rs2808629	1	159676796	G/A	1.01	0.766	1.24	0.002	1.22	7.62E-09
CRP1/CRP rs2794520	1	159678816	C/T	1.01	0.732	1.24	0.002	1.22	6.36E-09
CRP rs1205	1	159682233	C/T	1.01	0.736	1.24	0.002	1.22	9.46E-09
CRP rs3091244 ^e	1	159684665	G/A	0.99	0.826	0.81	0.002	0.83	3.12E-08
CRP rs2027471 ^e	1	159689388	T/A	1.01	0.758	1.23	0.003	1.21	2.74E-08
CRP rs1341665 ^e	1	159691559	G/A	1.01	0.797	1.22	0.004	1.21	2.49E-08
CRP rs2211320	1	159693605	G/A	1.01	0.677	1.26	0.001	1.23	9.94E-09
CRP rs7551731 ^e	1	159694779	T/C	1.01	0.659	1.25	0.002	1.22	1.82E-08
CRP rs7553007 ^e	1	159698549	G/A	1.02	0.636	1.25	0.001	1.22	2.04E-08
CRP rs4546916 ^e	1	159699249	G/T	1.02	0.639	1.25	0.001	1.22	1.99E-08
CRP rs4287174 ^e	1	159703442	T/A	1.01	0.662	1.25	0.002	1.22	2.30E-08
CRP rs4428887 ^e	1	159703462	A/G	1.01	0.753	1.23	0.003	1.21	4.50E-08
CRP rs12037186 ^e	1	159706230	A/G	1.02	0.581	1.25	0.002	1.22	3.00E-08
HNF1A-AS1 rs11065365	12	121592040	G/A	1.13	0.0003	0.64	5.91E-10	0.87	0.0002
HNF1A-AS1 rs7953249	12	121403724	G/A	1.10	0.002	0.68	7.19E-09	0.88	0.0001
HNF1A-AS1 rs7135337	12	121404155	A/C	1.10	0.002	0.67	5.16E-09	0.87	6.09E-05
HNF1A-AS1 rs1920792	12	121404584	C/T	0.91	0.002	1.45	7.64E-09	1.12	0.0004
HNF1A-AS1 rs10774579	12	121405210	C/T	0.91	0.002	1.43	2.92E-08	1.12	0.0004
HNF1A-AS1 rs2393792 ^e	12	121406293	A/G	0.91	0.002	1.43	4.67E-08	1.12	0.001
HNF1A-AS1 rs2255531	12	121414915	A/G	1.11	0.002	0.67	1.05E-08	0.86	2.73E-05
HNF1A-AS1 rs7139079	12	121415293	G/A	1.10	0.001	0.66	8.91E-10	0.87	2.04E-05
HNF1A-AS1 rs2464190	12	121415390	C/T	1.11	0.001	0.66	8.24E-10	0.87	5.72E-05
HNF1A rs1169289	12	121416622	G/C	1.09	0.006	0.68	1.69E-08	0.87	6.50E-05

(Continued on the following page)

Table 5. GWA analysis for the association with CRP, stratified by obesity-related lifestyle factors (physical activity and fat diet). (Cont'd)

SNP	Chr	Position ^a	Allele ^b (Ref/Alt)	Interaction test		Active/less-fat diet group		Inactive/high-fat diet group		
				OR ^c	P	OR ^c	P ^d	OR ^c	P ^d	Q
<i>HNF1A</i> rs1169288	12	121416650	C/A	1.10	0.005	0.66	2.90E-08	0.84	1.94E-06	0.160
<i>HNF1A</i> rs2244608	12	121416988	G/A	1.09	0.006	0.67	2.07E-08	0.84	9.16E-07	0.291
<i>HNF1A</i> rs1169284	12	121419926	C/T	1.10	0.003	0.66	6.49E-09	0.84	1.05E-06	0.242
<i>HNF1A</i> rs7979473	12	121420260	A/G ^f	1.11	0.001	0.65	2.29E-10	0.86	1.50E-05	0.158
<i>HNF1A</i> rs7979478	12	121420263	A/G ^f	1.11	0.001	0.65	1.88E-10	0.86	9.31E-06	0.299
<i>HNF1A</i> rs1183910	12	121420807	A/G	1.10	0.003	0.65	3.58E-09	0.84	1.33E-06	0.236
<i>HNF1A</i> rs11065384	12	121423285	T/C	1.10	0.003	0.65	4.11E-09	0.84	1.24E-06	0.226
<i>HNF1A</i> rs7970695	12	121423376	G/A ^f	1.10	0.001	0.65	3.60E-10	0.86	1.09E-05	0.149
<i>HNF1A</i> rs11065385	12	121423386	A/G	1.10	0.003	0.65	4.01E-09	0.84	1.13E-06	0.228
<i>HNF1A</i> rs9738226	12	121423659	A/G ^f	1.10	0.002	0.65	4.49E-10	0.86	1.11E-05	0.146
<i>HNF1A</i> rs2393791	12	121423956	C/T ^f	1.10	0.001	0.65	3.13E-10	0.86	1.16E-05	0.141
<i>HNF1A</i> rs2393776	12	121424406	G/A ^f	1.11	0.001	0.65	1.80E-10	0.86	8.07E-06	0.218
<i>HNF1A</i> rs2243458	12	121424490	T/A ^f	1.11	0.002	0.65	1.58E-09	0.84	1.09E-06	0.300
<i>HNF1A</i> rs2393775	12	121424574	G/A ^f	1.11	0.001	0.65	1.84E-10	0.86	8.16E-06	0.223
<i>HNF1A</i> rs7310409	12	121424861	A/G ^f	1.11	0.001	0.65	1.84E-10	0.86	8.23E-06	0.222
<i>HNF1A</i> rs1169292	12	121426478	T/C	1.10	0.004	0.66	1.08E-08	0.84	1.74E-06	0.295
<i>HNF1A</i> rs1169294	12	121426594	A/G	1.10	0.004	0.66	1.16E-08	0.84	1.20E-06	0.284

Note: Only genome-wide significant SNPs included. Numbers in bold face are statistically significant.

Abbreviations: Alt, alternative; Chr, chromosome; PA, physical activity; Q, Cochran Q; Ref, reference.

^aGRCh 37 coordinated.

^bAdditive genetic model regressed.

^cGWA analysis for CRP adjusted for age and 10 PCs.

^dP values were adjusted to correct for multiple testing via the Benjamini-Hochberg approach.

^eSNPs did not reach the genome-wide significant level after multiple testing was corrected.

^fThe results from the additive model are presented; the dominant model had similar effect sizes and P values.

Table 6. GWA analysis for the association with IL6.

SNP	Chr	Position ^a	Allele ^b (Ref/Alt)	Overall analysis/Interaction test			Nonobese/less-fat diet/Active group			Obese/high-fat diet/Inactive group		
				OR ^c	P	Q	OR ^c	P ^d	Q	OR ^c	P ^d	Q
Overall analysis												
<i>DEC1</i> rs149109490	9	118330052	T/C	2.81	3.77E-08	0.36						
Interaction test for WHR												
							Non-visceral-obese group (WHR ≤ 0.85, n = 7,251)			Visceral obese group (WHR > 0.85, n = 2,928)		
<i>DEC1</i> rs149109490	9	118330052	T/C	0.70	0.056	0.169	3.61	3.42E-08	0.078	1.74	0.105	0.770
Interaction test for SFA												
							Less-fat diet group (% cal. from SFA < 9.0, n = 2,306)			High-fat diet group (% cal. from SFA ≥ 9.0, n = 7,873)		
<i>DEC1</i> rs149109490	9	118330052	T/C	1.14	0.607	0.891	2.01	0.199	0.907	3.19	4.21E-08	0.144
Interaction test for PA												
							Active group (MET ≥ 10, n = 4,221)			Inactive group (MET < 10, n = 5,958)		
<i>MAPK1</i> rs56398890	22	22190785	A/T	1.16	5.29E-06	0.533	0.74	4.54E-08	0.978	1.00	0.909	0.335
<i>MAPK1</i> rs9607320	22	22202164	T/C	1.17	3.73E-06	0.718	0.73	1.18E-08	0.938	0.99	0.897	0.292

Note: Only genome-wide significant SNPs included in overall and interaction tests (G × E or stratified analysis). Numbers in bold face are statistically significant. Abbreviations: Alt, alternative; Chr, chromosome; PA, physical activity; Q, Cochran Q; Ref, reference.

^aGRCh 37 coordinated.

^bAdditive genetic model regressed.

^cGWA analysis for IL6 adjusted for age and 10 PCs.

^dP values were adjusted to correct for multiple testing via the Benjamini-Hochberg approach.

associated with reduced risk for chronic inflammation (i.e., decreased CRP levels). Furthermore, similar to previous literature (48, 49), we found that the decreased effects of the *HNF1A* SNPs on CRP levels were more profound in the low-fat diet subgroup, conferring the regulating role of CRP on expression of proprotein convertase subtilisin/kexin type-9, a key regulator of low-density lipoprotein (LDL) metabolism, via the p38MAPK-HNF1A pathway in hepatic cells, as recently reported by *in vivo* studies (50, 51). Also, we found novel SNPs at *C12orf43*, which is located downstream of *HNF1A* in a tail-to-tail manner (52), that were associated with a reduced risk for chronic inflammation (i.e., decreased CRP levels); this finding warrants further biological mechanistic study of a link between the particular gene, its mutation, and CPR phenotype.

For many genes discovered by GWASs for metabolic traits, such as chronic inflammation, the mechanism by which the encoded proteins affect disease risk is controversial. Some intergenic or intronic SNPs may affect the function of transcriptional control structures, including enhancers and silencers (53). Of the 88 top markers in our study, 27 in *HNF1A/HNF1A-AS1/C12orf43* displayed the increased risk for breast cancer. Their functional significance and the molecular mechanisms of the target genes that mediated breast carcinogenesis remain unclear. The *HNF1A* gene regulates tissue-specific expression of multiple genes in the liver, pancreas (45), proximal tubule of the kidney (54), and epithelial cells of the intestine (55), and its mutation has been associated with maturity-onset diabetes of the young type 3 (56), hepatocellular adenoma (HCC; ref. 57), and endometrial (58) and pancreatic carcinomas (59). Whether the *HNF1A* gene and its mutation

act as a tumor suppressor (59) or an oncogene (55, 60) on the specific types of tumor (HCC and pancreatic cancer) has not been determined. For example, one study (61) reported that *HNF1A* silencing in HCC cells led to overexpression of several genes encoding growth factor receptors, components of translational machinery, cell cycle, and angiogenesis regulators, which promote cell proliferation and suppress apoptosis. Another study (60) presented high *HNF1A* gene expression in pancreatic cancer stem cells and ductal adenocarcinoma. Our study is the first to show that SNPs near *HNF1A/HNF1A-AS1* in overall and visceraally obese subgroups were associated with increased risk for breast cancer, calling for study of the biological molecular mechanisms in the gene-breast cancer pathways.

In relation to increased risk for chronic low-grade inflammation on the basis of IL6, we found one novel SNP near *DEC1*, whose expression was suppressed in esophageal cancer (62), suggestive of a candidate tumor suppressor. *DEC1* mutations' functional implications in other tumor cells have not been determined. In our study, the SNP near *DEC1* had a substantial suppressive effect on breast cancer development, although that effect did not reach statistical significance.

Because of the restrictions of the available data, our study focused on two key inflammatory biomarkers (CRP and IL6) and examined obesity factors at screening in cross-section fashion. In addition, our study was confined to non-Hispanic white postmenopausal women, so the generalizability of our results to other populations is limited. All identified loci contributed to 6.3% of interindividual variability in proinflammatory cytokines. Future DNA methylation studies may address this low heritability. In addition,

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the chronic inflammation pathway involved in breast cancer among postmenopausal women may differ by the cancer's molecular subtype and the exogenous estrogen intake; our study could not delineate such differences due to insufficient statistical power. Finally, some meta-analysis results indicated pleiotropic effects of inflammatory cytokines.

In conclusion, our study suggests that a number of newly identified top signals may exert their effect on chronic low-grade inflammation by interacting with obesity factors and may have implications for breast carcinogenesis in postmenopausal women. Our findings may contribute to better understanding of the molecular genetic associations between proinflammation and cancer and suggest potential intervention strategies, such as body weight control, for women who carry the inflammatory-risk genotypes, thus reducing their risk for breast cancer.

Authors' Disclosures

No disclosures were reported.

Authors' Contributions

S.Y. Jung: Conceptualization, resources, data curation, formal analysis, funding acquisition, investigation, visualization, methodology, writing-original draft, project administration, writing-review and editing. **P.A. Scott:** Data curation, software, formal analysis, validation, investigation, visualization, methodology, writing-original draft, writing-review and editing. **J.C. Papp:** Conceptualization, resources, software, supervision, validation, writing-original draft, writing-review and editing. **E.M. Sobel:** Conceptualization, resources, data curation, supervision, investigation, writing-original draft, writing-review and editing. **M. Pellegrini:** Conceptualization, resources, supervision, writing-original draft, writing-review and editing. **H. Yu:** Conceptualization, resources, supervision, investigation, writing-original draft, writing-review and editing. **S. Han:** Formal analysis, visualization, methodology, writing-original draft, writing-review and editing. **Z.-F. Zhang:** Conceptualization, resources, supervision, validation, visualization, methodology, writing-original draft, writing-review and editing.

References

- Coussens LM, Werb Z. Inflammation and cancer. *Nature* 2002;420:860–7.
- Disis ML. Immune regulation of cancer. *J Clin Oncol* 2010;28:4531–8.
- Grivnennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. *Cell* 2010;140:883–99.
- Chao T, Ladd JJ, Qiu J, Johnson MM, Israel R, Chin A, et al. Proteomic profiling of the autoimmune response to breast cancer antigens uncovers a suppressive effect of hormone therapy. *Proteomics Clin Appl* 2013;7:327–36.
- Nelson SH, Brasky TM, Patterson RE, Laughlin GA, Kritz-Silverstein D, Edwards BJ, et al. The Association of the C-Reactive protein inflammatory biomarker with breast cancer incidence and mortality in the women's health initiative. *Cancer Epidemiol Biomarkers Prev* 2017;26:1100–6.
- Purohit A, Reed MJ. Regulation of estrogen synthesis in postmenopausal women. *Steroids* 2002;67:979–83.
- Bogaty P, Brophy JM, Noel M, Boyer L, Simard S, Bertrand F, et al. Impact of prolonged cyclooxygenase-2 inhibition on inflammatory

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- markers and endothelial function in patients with ischemic heart disease and raised C-reactive protein: a randomized placebo-controlled study. *Circulation* 2004;110:934–9.
- Retterstol L, Eikvar L, Berg K. A twin study of C-Reactive Protein compared to other risk factors for coronary heart disease. *Atherosclerosis* 2003;169:279–82.
- Dupuis J, Larson MG, Vasan RS, Massaro JM, Wilson PW, Lipinska I, et al. Genome scan of systemic biomarkers of vascular inflammation in the Framingham Heart Study: evidence for susceptibility loci on 1q. *Atherosclerosis* 2005;182:307–14.
- Worms MA, Victor A, Galle PR, Hohler T. Genetic and environmental contributions to plasma C-reactive protein and interleukin-6 levels—a study in twins. *Genes Immun* 2006;7:600–5.
- Dehghan A, Dupuis J, Barbalic M, Bis JC, Eiriksdottir G, Lu C, et al. Meta-analysis of genome-wide association studies in >80,000 subjects identifies multiple loci for C-reactive protein levels. *Circulation* 2011;123:731–8.
- Amaral WZ, Krueger RF, Ryff CD, Coe CL. Genetic and environmental determinants of population variation in interleukin-6, its

- soluble receptor and C-reactive protein: insights from identical and fraternal twins. *Brain Behav Immun* 2015;49:171–81.
13. Ligthart S, Vaez A, Vosa U, Stathopoulou MG, de Vries PS, Prins BP, et al. Genome analyses of >200,000 individuals identify 58 loci for chronic inflammation and highlight pathways that link inflammation and complex disorders. *Am J Hum Genet* 2018;103:691–706.
 14. Doumatey AP, Chen G, Tekola Ayele F, Zhou J, Erdos M, Shriner D, et al. C-reactive protein (CRP) promoter polymorphisms influence circulating CRP levels in a genome-wide association study of African Americans. *Hum Mol Genet* 2012;21:3063–72.
 15. Hu M, Lee MH, Mak VW, Tomlinson B. Effect of central obesity, low high-density lipoprotein cholesterol and C-reactive protein polymorphisms on C-reactive protein levels during treatment with Rosuvastatin (10 mg Daily). *Am J Cardiol* 2010;106:1588–93.
 16. Iyengar NM, Arthur R, Manson JE, Chlebowski RT, Kroenke CH, Peterson L, et al. Association of body fat and risk of breast cancer in postmenopausal women with normal body mass index: a secondary analysis of a randomized clinical trial and observational study. *JAMA Oncol* 2019;5:155–63.
 17. Bermudez EA, Rifai N, Buring JE, Manson JE, Ridker PM. Relation between markers of systemic vascular inflammation and smoking in women. *Am J Cardiol* 2002;89:1117–9.
 18. Stewart SH, Mainous AG 3rd, Gilbert G. Relation between alcohol consumption and C-reactive protein levels in the adult US population. *J Am Board Fam Pract* 2002;15:437–42.
 19. Wu Y, McDade TW, Kuzawa CW, Borja J, Li Y, Adair LS, et al. Genome-wide association with C-reactive protein levels in CLHNS: evidence for the CRP and HNF1A loci and their interaction with exposure to a pathogenic environment. *Inflammation* 2012;35:574–83.
 20. Prasad G, Giri AK, Basu A, Tandon N, Bharadwaj D. Genome-wide association study for C-reactive protein in Indians replicates known associations of common variants. *J Genet* 2019;98:20.
 21. Gong Z, Quan L, Yao S, Zirpoli G, Bandera EV, Roberts M, et al. Innate immunity pathways and breast cancer risk in African American and European-American women in the Women's Circle of Health Study (WCHS). *PLoS One* 2013;8:e72619.
 22. Connor AE, Baumgartner RN, Baumgartner KB, Pinkston CM, Boone SD, John EM, et al. Associations between ALOX, COX, and CRP polymorphisms and breast cancer among Hispanic and non-Hispanic white women: The breast cancer health disparities study. *Mol Carcinog* 2015;54:1541–53.
 23. Fairey AS, Courneya KS, Field CJ, Bell GJ, Jones LW, Martin BS, et al. Effect of exercise training on C-reactive protein in postmenopausal breast cancer survivors: a randomized controlled trial. *Brain Behav Immun* 2005;19:381–8.
 24. Healy LA, Ryan AM, Carroll P, Ennis D, Crowley V, Boyle T, et al. Metabolic syndrome, central obesity and insulin resistance are associated with adverse pathological features in postmenopausal breast cancer. *Clin Oncol (R Coll Radiol)* 2010;22:281–8.
 25. Ellis J, Lange EM, Li J, Dupuis J, Baumert J, Walston JD, et al. Large multiethnic Candidate Gene Study for C-reactive protein levels: identification of a novel association at CD36 in African Americans. *Hum Genet* 2014;133:985–95.
 26. American Cancer Society. *Cancer Facts and Figures, 2019*. Atlanta, GA: American Cancer Society; 2019.
 27. Design of the Women's Health Initiative clinical trial and observational study. The Women's Health Initiative Study Group. *Control Clin Trials* 1998;19:61–109.
 28. NCBI: WHI Harmonized and Imputed GWAS Data. A sub-study of Women's Health Initiative; 2019. Available at: https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000746.v3.p3.
 29. Orchard TS, Andridge RR, Yee LD, Lustberg MB. Diet quality, inflammation, and quality of life in breast cancer survivors: a cross-sectional analysis of pilot study data. *J Acad Nutr Diet* 2018;118:578–881.
 30. Gunter MJ, Wang T, Cushman M, Xue X, Wassertheil-Smoller S, Strickler HD, et al. Circulating adipokines and inflammatory markers and postmenopausal breast cancer risk. *J Natl Cancer Inst* 2015;107:djv169.
 31. National Cancer Institute. SEER Program: Comparative Staging Guide For Cancer June 1993. Available at: https://seer.cancer.gov/archive/manuals/historic/comp_stage1.1.pdf.
 32. Women's Health Initiative: WHI Follow-Up Dataset: Form 41 - Addendum to Personal Information (Race). Available at: <https://www.whi.org/doc/F041-v1.1.pdf>.
 33. Schumacher FR, Al Olama AA, Berndt SI, Benlloch S, Ahmed M, Saunders EJ, et al. Association analyses of more than 140,000 men identify 63 new prostate cancer susceptibility loci. *Nat Genet* 2018;50:928–36.
 34. Loh PR, Bhatia G, Gusev A, Finucane HK, Bulik-Sullivan BK, Pollack SJ, et al. Contrasting genetic architectures of schizophrenia and other complex diseases using fast variance-components analysis. *Nat Genet* 2015;47:1385–92.
 35. Manichaikul A, Mychaleckyj JC, Rich SS, Daly K, Sale M, Chen WM. Robust relationship inference in genome-wide association studies. *Bioinformatics* 2010;26:2867–73.
 36. Ishwaran H, Kogalur UB. Random Forests for survival, regression, and classification. October 17, 2017. Available at: <https://cran.r-project.org/web/packages/randomForestSRC/randomForestSRC.pdf>.
 37. Higgins JP, Thompson SG. Quantifying heterogeneity in a meta-analysis. *Stat Med* 2002;21:1539–58.
 38. Wiens BL, Dmitrienko A, Marchenko O. Selection of hypothesis weights and ordering when testing multiple hypotheses in clinical trials. *J Biopharm Stat* 2013;23:1403–19.
 39. Tao Q, Ang TFA, DeCarli C, Auerbach SH, Devine S, Stein TD, et al. Association of chronic low-grade inflammation with risk of alzheimer disease in ApoE4 carriers. *JAMA Netw Open* 2018;1:e183597.
 40. Buckley DI, Fu R, Freeman M, Rogers K, Helfand M. C-reactive protein as a risk factor for coronary heart disease: a systematic review and meta-analyses for the U.S. Preventive Services Task Force. *Ann Intern Med* 2009;151:483–95.
 41. CRP gene. C-reactive protein. Genetics Home Reference 2019. Available at: <https://ghr.nlm.nih.gov/gene/CRP#conditions>.
 42. Walsh MT, Divane A, Whitehead AS. Fine mapping of the human pentraxin gene region on chromosome 1q23. *Immunogenetics* 1996;44:62–9.
 43. Goldman ND, Liu T, Lei KJ. Structural analysis of the locus containing the human C-reactive protein gene and its related pseudogene. *J Biol Chem* 1987;262:7001–5.
 44. Curocichin G, Wu Y, McDade TW, Kuzawa CW, Borja JB, Qin L, et al. Single-nucleotide polymorphisms at five loci are associated with C-reactive protein levels in a cohort of Filipino young adults. *J Hum Genet* 2011;56:823–27.
 45. Shih DQ, Bussen M, Sehayek E, Ananthanarayanan M, Shneider BL, Suchy FJ, et al. Hepatocyte nuclear factor-1alpha is an essential regulator of bile acid and plasma cholesterol metabolism. *Nat Genet* 2001;27:375–382.
 46. Nishikawa T, Hagihara K, Serada S, Isobe T, Matsumura A, Song J, et al. Transcriptional complex formation of c-Fos, STAT3, and hepatocyte NF-1 alpha is essential for cytokine-driven C-reactive protein gene expression. *J Immunol* 2008;180:3492–501.
 47. Juszczak A, Pavic T, Vuckovic F, Bennett AJ, Shah N, Pape Medvidovic E, et al. Plasma fucosylated glycans and C-reactive protein as biomarkers of HNF1A-MODY in young adult-onset nonautoimmune diabetes. *Diabetes Care* 2019;42:17–26.
 48. Zhou YJ, Yin RX, Hong SC, Yang Q, Cao XL, Chen WX. Association of the HNF1A polymorphisms and serum lipid traits, the risk of

Jung et al.

- coronary artery disease and ischemic stroke. *J Gene Med* 2017;19:e2941.
49. Huang T, Wang T, Heianza Y, Sun D, Ivey K, Durst R, et al. HNF1A variant, energy-reduced diets and insulin resistance improvement during weight loss: The POUNDS Lost trial and DIRECT. *Diabetes Obes Metab* 2018;20:1445–52.
 50. Cui CJ, Li S, Zhu CG, Sun J, Du Y, Zhang Y, et al. Enhanced pro-protein convertase subtilisin/kexin type 9 expression by C-reactive protein through p38MAPK-HNF1alpha pathway in HepG2 cells. *J Cell Mol Med* 2016;20:2374–83.
 51. Shende VR, Wu M, Singh AB, Dong B, Kan CF, Liu J. Reduction of circulating PCSK9 and LDL-C levels by liver-specific knock-down of HNF1alpha in normolipidemic mice. *J Lipid Res* 2015;56:801–9.
 52. Erdmann J, Grosshennig A, Braund PS, Konig IR, Hengstenberg C, Hall AS, et al. New susceptibility locus for coronary artery disease on chromosome 3q22.3. *Nat Genet* 2009;41:280–2.
 53. O'Brien RM. Moving on from GWAS: functional studies on the G6PC2 gene implicated in the regulation of fasting blood glucose. *Curr Diab Rep* 2013;13:768–77.
 54. Terryn S, Tanaka K, Lengele JP, Olinger E, Dubois-Laforgue D, Garbay S, et al. Tubular proteinuria in patients with HNF1alpha mutations: HNF1alpha drives endocytosis in the proximal tubule. *Kidney Int* 2016;89:1075–89.
 55. Yang R, Kerschner JL, Harris A. Hepatocyte nuclear factor 1 coordinates multiple processes in a model of intestinal epithelial cell function. *Biochim Biophys Acta* 2016;1859:591–8.
 56. Bellanne-Chantelot C, Carette C, Riveline JP, Valero R, Gautier JF, Larger E, et al. The type and the position of HNF1A mutation modulate age at diagnosis of diabetes in patients with maturity-onset diabetes of the young (MODY)-3. *Diabetes* 2008;57:503–8.
 57. Bluteau O, Jeannot E, Bioulac-Sage P, Marques JM, Blanc JF, Bui H, et al. Bi-allelic inactivation of TCF1 in hepatic adenomas. *Nat Genet* 2002;32:312–5.
 58. Rebouissou S, Rosty C, Lecuru F, Boisselier S, Bui H, Le Frere-Belfa MA, et al. Mutation of TCF1 encoding hepatocyte nuclear factor 1alpha in gynecological cancer. *Oncogene* 2004;23:7588–92.
 59. Janky R, Binda MM, Allemeersch J, Van den Broeck A, Govaere O, Swinnen JV, et al. Prognostic relevance of molecular subtypes and master regulators in pancreatic ductal adenocarcinoma. *BMC Cancer* 2016;16:632.
 60. Abel EV, Goto M, Magnuson B, Abraham S, Ramanathan N, Hotaling E, et al. HNF1A is a novel oncogene that regulates human pancreatic cancer stem cell properties. *Elife* 2018;7:e33947.
 61. Pelletier L, Rebouissou S, Paris A, Rathahao-Paris E, Perdu E, Bioulac-Sage P, et al. Loss of hepatocyte nuclear factor 1alpha function in human hepatocellular adenomas leads to aberrant activation of signaling pathways involved in tumorigenesis. *Hepatology* 2010;51:557–66.
 62. Nishiwaki T, Daigo Y, Kawasoe T, Nakamura Y. Isolation and mutational analysis of a novel human cDNA, DEC1 (deleted in esophageal cancer 1), derived from the tumor suppressor locus in 9q32. *Genes Chromosomes Cancer* 2000;27:169–76.
 63. Haskell WL, Lee IM, Pate RR, Powell KE, Blair SN, Franklin BA, et al. Physical activity and public health: updated recommendation for adults from the American College of Sports Medicine and the American Heart Association. *Med Sci Sports Exerc* 2007;39:1423–34.
 64. Van Horn L, Carson JA, Appel LJ, Burke LE, Economos C, Karmally W, et al. Recommended dietary pattern to achieve adherence to the American Heart Association/American College of Cardiology (AHA/ACC) guidelines: A scientific statement from the American Heart Association. *Circulation* 2016;134:e505–9.

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Genome-wide Association Analysis of Proinflammatory Cytokines and Gene–lifestyle Interaction for Invasive Breast Cancer Risk: The WHI dbGaP Study

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