Local Anti–PD-1 Delivery Prevents Progression of Premalignant Lesions in a 4NQO-Oral Carcinogenesis Mouse Model

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ABSTRACT

Although the principle of systemic treatment to prevent the progression of oral premalignant lesions (OPL) has been demonstrated, there remains a lack of consensus about an optimal approach that balances clinical efficacy with toxicity concerns. Recent advances in cancer therapy using approaches targeting the tumor immune microenvironment (TIME) including immune-checkpoint inhibitors indicate that these agents have significant clinically activity against different types of cancers, including oral cancer, and therefore they may provide an effective oral cancer prevention strategy for patients with OPLs. Our past work showed that systemic delivery of a monoclonal antibody to the pro–programmed death receptor 1 (PD-1) immune checkpoint can inhibit the progression of OPLs to oral cancer in a syngeneic murine oral carcinogenesis model. Here we report a novel approach of local delivery of a PD-1 immune-checkpoint inhibitor loaded using a hydrogel, which significantly reduces the progression of OPLs to carcinomas. In addition, we detected a significant infiltration of regulatory T cells associated with oral lesions with p53 mutation, and a severe loss of expression of STING, which correlated with a decreased infiltration of dendritic cells in the oral lesions. However, a single local dose of PD-1 inhibitor was found to restore stimulator of interferon response cGAMP interactor 1 (STING) and CD11c expression and increase the infiltration of CD8⁺ T cells into the TIME irrespective of the p53 mutational status. Overall, we provide evidence for the potential clinical value of local delivery of biomaterials loaded with anti–PD-1 antibodies to prevent malignant progression of OPLs.

Prevention Relevance: Oral cancer is an aggressive disease, with an overall survival rate of 50%. Preinvasive histologic abnormalities such as tongue dysplasia represent an early stage of oral cancer; however, there are no treatments to prevent oral carcinoma progression. Here, we combined biomaterials loaded with an immunotherapeutic agent preventing oral cancer progression.

Introduction

Oral cancer represents the sixth most common cancer worldwide with approximately 630,000 new patients diagnosed annually, resulting in more than 350,000 deaths every year (1). More than 90% of head and neck cancers are squamous cell carcinomas (HNSCC), many of which arise from the progression of oral premalignant lesions (OPL) through the accumulation of genomic alterations that arise as a result of the host’s genetic predisposition to accumulation of DNA damage and environmental exposure to carcinogens including tobacco and alcohol (2–4).

Studies of the mutational landscape of non–HPV–associated head and neck squamous cell carcinomas (HNSCC) have demonstrated genomic alterations in TP53 in ~85% of cases (5). Notably, high-risk TP53 mutations have been associated with poor survival and lack of response to chemotherapy in patients with head and neck cancer, suggesting that cancers of the oral cavity carrying high-risk TP53 mutations can be refractory to standard therapeutic approaches (6–10). Mutations in TP53 have been detected in approximately 30% of the OPLs, suggesting that these mutations arise early during oral tumor development and might influence the progression of OPLs, and their response to preventive strategies (11, 12).

The current standard treatments for OPLs include surgical excision and carbon dioxide laser ablation (13). Unfortunately, 0.13%–34% of these lesions still progress to oral cancer despite treatment (14). Given that treatment failure can also occur as a
result of mucosal areas adjacent to the excised OPL proceeding down the multistep carcinogenesis pathway to oral cancer, systemic therapy approaches that can reach the entire mucosal field at risk have been thought to have value in this setting. Although this approach has been proven in principle with administration of retinoids, a recent clinical prevention trial in oral cancer using erlotinib did not improve cancer-free survival in high-risk patients with OPLs, suggesting that inhibition of epidermal growth factor receptor is not sufficient to prevent oral cancer progression (15).

Immunotherapy, in the form of immune-checkpoint inhibitors (ICI), such as PD-1 and cytotoxic T-lymphocyte-associated protein 4 (CTLA-4), has recently been introduced as a promising therapeutic option for patients with solid tumors, including oral squamous cell carcinoma (OSCC; refs. 16–19), which arises from OPLs through a multistep carcinogenesis process (20, 21).

A recent immunoprevention study demonstrated that PD-1 blockade reduces the incidence of OPLs in a carcinogen mouse model. This study used Trp53fl/fl heterozygous mutant mice to accelerate OPLs in response to carcinogen. Systemic PD-1 treatment significantly reduced the incidence of malignant lesions with a high infiltration of activated cytotoxic CD8+ T cells, supporting the preventive potential of immune-checkpoint inhibition to contain oral cancer development (22). Furthermore, other studies using the same mouse model had demonstrated that cytokine levels are modulated in response to PD-1 treatment and this treatment also prevented oral lesion progression into carcinoma (23). In a more recent study, PD-1 treatment significantly reduced the formation of oral premalignant lesions, and the TIME can affect the tumor immune microenvironment (TIME) and consist of self-assembled peptides that mimic the extracellular matrix by generating a nanofibrous network to create a hydrogel. The hydrogel can encapsulate drugs, cytokines, and growth factors and control their sustained release to permit a sustained payload release in vivo settings (26–30). Although we have used this MDP to deliver a variety of small molecules and/or proteins in the past, reported for the first time here, we load the MDP hydrogel with anti–PD-1 immune-checkpoint inhibitor to enable local delivery with sustained release. To study the efficacy of local anti–PD-1 immunotherapy delivery to OPLs and define the role of mutant p53 in responsiveness to this treatment, we used a carcinogen, 4NQO, induced oral cancer development model in wild-type and mutant p53R172H mice that mimics the tobacco-mediated oral carcinogenesis observed in humans (31). In addition, we used MDP-based hydrogels as a novel biomaterial to control the payload delivery in a controlled manner, slowing the release of antibody locally. In this study, we monitored oral tumor development, analyzed immune cell infiltration in the premalignant lesions, and determined whether p53 mutations modulate the immunoprevention potential of ICIs.

Materials and Methods

Mouse models

We generated mice in which the endogenous p53R172H mutation was activated using K14Cre mice, which drives the activation of p53R172H mutant and floxed alleles in the oral epithelia. The following groups of mice were used: (i) mice with oral activation of p53R172H and deletion of the floxed wild-type p53 allele (K14Cre; p53R172Hfl/flox); (ii) mice with two p53 wild-type alleles (K14Cre; p53flox/flox). Mouse genotyping and activation of conditional alleles were determined by PCR using genomic DNA purified from mouse tails as previously described (32–33).

To induce oral lesions, mice were exposed to the carcinogen (4NQO, 100 µg/mL) in the drinking water containing 1% sucrose (Fisher Scientific) for 8 weeks, and monitored for oral lesions. A stock solution of 4NQO (50 mg/mL) was prepared by dissolving 4NQO powder (Sigma-Aldrich) in DMSO, which was stored at −20°C until used. All comparative studies were conducted using littermates with the appropriate genotypes. All animal studies were approved by the Institutional Animal Care and Use Committee (IACUC) of the University of Texas MD Anderson Cancer Center (Houston, TX).

Immunoprevention preclinical studies

In our study, we used 8–16-week-old mice and each mouse group consisted of seven mice and were distributed and treated as follows: K14Cre; p53 wild-type (5 females and 2 males) were treated with isotype IgG control-hydrogel; K14Cre; p53 wild-type (4 females and 3 males) were treated with PD1 gel; K14Cre; p53 R172H (4 females and 3 males*) were treated with...
isotype IgG control-hydrogel [*one male was dead during PD-1 treatment; unknown cause]; and K14Cre; p53 R172H (4 females** and 3 males**) were treated with PD-1 gel [*one male and one female were dead during PD-1 treatment; unknown cause]. Oral lesions were induced by 4NQO, as previously described. Four weeks after completion of the 4NQO treatment the mice were injected once in the tongue using 60 μL. MDP hydrogel containing 300 μg anti–PD-1 (Bio Cell; cat. #BE0146) or IgG2a (Bio Cell; cat. #BE0089). Because tongue microscopic lesions cannot be visualized at 4 weeks after exposure to 4NQO, we decided to inject 20 μL in three distinct sites (two dorsal and one central tongue) to facilitate drug distribution within the tongue.

All animals underwent weekly examination of the oral cavity followed by necropsy for tissue retrieval five weeks after completion of treatment. Tumor-bearing mice were euthanized by CO2. Finally, oral tissues were surgically excised, measured, photographed, and formalin fixed for histopathologic studies.

**Peptide synthesis**

Reagents for peptide synthesis reagents were acquired from EMD Chemicals. Both manual synthesis (typically performed at 0.15 mmol scale) and an Apex Focus XC (Aapptec) automatic synthesizer (typically performed at 0.45 mmol scale) were used for synthesis of multidomain peptides K2(SL)6K2 (K2-MDP). Peptides were synthesized using standard F-MOC chemistry and solid-phase peptide synthesis methods described previously (34–37). All peptides were N-terminally acetylated and C-terminally amidated. Peptides were cleaved from solid-phase resin by shaking for 3 hours with trifluoroacetic acid (TFA) and protecting scavengers, in a 2:1:1:18 ratio of Milli-Q (MQ) H2O: trisopropylsilane (TIPS): anisole: TFA. Rotary evaporation was used to remove excess TFA, and trituration with cold diethyl ether yielded crude peptide. The peptide was pH adjusted to approximately 7.0, and dialyzed against MQ water for 5–7 days to remove small-molecule scavengers, TFA, and other contaminants using 100–300 Da MWCO dialysis tubing (Spectra/Per, Spectrum Laboratories Inc.). After dialysis, peptide solutions were pH adjusted to pH 7.2–7.4, after which 0.2-μm filters were used to sterile filter the solutions, which were frozen at −80°C and lyophilized to dried powder for storage at −20°C. Peptides were analyzed before and after dialysis by Autoflex MALDI-TOF MS (Bruker Instruments) for confirmation of expected peptide mass.

**Hydrogel preparation and drug loading**

All chemicals not otherwise specified were purchased from Sigma-Aldrich. For preparation of sterile MDP hydrogels, 2% wt/vol solutions were dissolved in 298 mmol/L sucrose to support cytocompatibility. Anti–PD-1 checkpoint inhibitor antibodies and IgG isotype controls were purchased from Bio X Cell. Peptide stock solutions (K2-MDP) were first prepared at 4% wt/vol in 298 mmol/L sucrose. The target final loading concentrations of checkpoint inhibitor antibodies within the gels were 300 μg anti–PD-1 per 60 μL gel (5 μg/μL for PD-1) or the same loading concentration of respective isotype IgG for negative control tests. The antibody stocks were provided in PBS, a buffer that is compatible for peptide hydrogelation via phosphate anion cross-linking of charged peptide nanofibers.

The volume of antibody stock solution (usually provided at stock concentrations of ~7–9 μg/μL) required to achieve the final desired dose in the gel was then calculated, typically requiring a mixing of 1 part 4% wt/vol peptide with 2–3 parts antibody stock solution (with any leftover volume supplemented with additional PBS). The final antibody-loaded hydrogel formulations were 0.75X PBS, 75 mmol/L sucrose, 1% wt/vol peptide (10 mg/mL, ~5–6 mmol/L), 5 μg/μL PD-1 or isotype control.

**Histology and IHC analysis**

The tongues were fixed in 10% neutral-buffered formalin at room temperature for at least 24 hours. Then, the tissue was transferred to 70% ethanol and embedded in paraffin. Histologic sections (5 μm) were stained with hematoxylin and eosin (H&E) or processed for IHC analysis. H&E staining was used for histopathologic analysis. Tongue lesions were diagnosed based on worse diagnosis based on the evaluation of the whole tongue and three slides per mouse were analyzed by a pathologist blinded to the treatment group. The oral lesions were classified as previously described (22). Slides were subjected to IHC staining with indicated antibodies. The primary antibodies used for IHC were CD8a (Synaptic Systems, 1:1,000, cat. # 361003), CD4 (Cell Signaling Technology, 1:100, cat. #25229), FOXP3 (eBioscience, 1:100, cat. #14-5773-82), PD-1 (Cell Signaling Technology, 1:100, cat. #84651S), CD11C (Cell Signaling Technology, 1:200, cat. #97585), and STING (Cell Signaling Technology, 1:100, cat. #13647). Images were captured on a DMLA microscope equipped with a DFC310 FX camera (Leica Microsystems). We used 10× objective to take images and included 100 mm bar scale to frame 1 mm² area. Next, the images were analyzed in a bigger screen, and a grid was superimposed to delimit area and facilitate immunostained cell quantification. We also consider subcellular staining marker like FOXP3 (nuclear) CD4, CD8, PD-1, CD11C (cell surface), and STING (intracellular). The quantification score was based on the number of positive stained cells per mm² area. The low- and high-grade oral lesions were heterogeneous in length and width; therefore, IHC quantification scored as density of cells, defined as the number of positive cells per mm². Quantification was evaluated by the same pathologist in the previous report (22).

**Statistical analysis**

Statistical analysis was performed using SPSS 19.0 (SPSS) and GraphPad Prism 8.0 (GraphPad Software). All data are presented as mean ± standard error. Two-tailed Student t test, one-way ANOVA, and χ² test were used to analyze the data. P values <0.05 were considered significant, defined * P < 0.05; ** P < 0.01; *** P < 0.001; **** P < 0.0001.
Results

Biomaterial loaded with anti-PD-1 antibody significantly inhibits malignant progression of 4NQO-induced oral lesions

To evaluate the effect of a PD-1 immune-checkpoint inhibitor in preventing the transition of oral premalignant to carcinoma lesions, we exposed mutant p53R172H and p53 wild-type mice to carcinogen 4NQO (100 μg/mL) in drinking water for 8 weeks (Fig. 1A). After four weeks of carcinogen treatment termination, we evaluated the efficacy of local immunotherapy administration using MDP-based hydrogels as a novel biomaterial to control the payload release of anti–PD-1 antibody and isotype control IgG2a to reduce off-target toxicity in the oral cavity. We administered a single dose of PD-1 antibody and isotype control IgG2a in hydrogel, and five weeks later mouse tongues were harvested, fixed, and processed to compare the incidence of both low-grade and high-grade lesions in the control and treated groups (Fig. 1A). During the collection of tissues, we observed relatively higher numbers of macroscopic lesions in the tongues of the mice treated with isotype control with the greatest number of macroscopic lesions seen in mutant p53R172H mice (Supplementary Fig. S1). Although male and female mice were used, we did not observe a difference in OPLs by sex. Next, blinded histopathologic analysis by independent individuals revealed that control and treatment groups developed low-grade lesions, consisting of mild and/or moderate dysplasia and high-grade lesions consisting of severe dysplasia and/or in situ carcinoma (Fig. 1B).

The histopathologic analysis revealed that p53 wild-type mice treated with PD-1 gel had a reduced lesion percentage (14.29%) in low and high-grade lesions and 71.42% of the mice were diagnosed with normal tissue. Wild-type mice treated with IgG control had relatively higher numbers of low and high-grade lesions (57.14 and 28.57% respectively) suggesting that treatment with the PD-1 antibody prevented or even eliminated the incidence of oral lesions (Fig. 1C). Moreover, mutant p53R172H mice treated with PD-1 gel showed a reduced frequency of high-grade lesions compared with control mice (20% vs. 60% respectively; Fig. 1C). Overall, these results strongly indicate that local immunotherapy delivery can prevent the malignant progression of OPLs in mutant and wild-type p53 mice. Furthermore, mutant p53R172H mice developed more high-grade lesions compared with wild-type mice and a significant increase in low-grade lesions even after PD-1 treatment, suggesting that mutant p53R172H accelerates carcinogen-induced oral tumor development than p53 wild-type mice and drives resistance to immunopreventive treatment (Fig. 1C). Finally, we can conclude the PD-1 treatment in p53 wild-type mice is more efficient compared with p53 mutant mice treated with isotype IgG control (Supplementary Fig. S2). Interestingly, we have confirmed experimentally that local and systemic delivery show very similar results. The results seen with delivery of a single PD-1 gel dose is comparable with those seen with eight anti–PD-1 doses systemically administered in p53 wild-type mice as shown in Supplementary Fig. S3. Finally, mice treated with intraperitoneal administration of anti–PD-1 showed a trend in weight loss at the end of the study; however, it was not statistically significant (Supplementary Fig. S4).

Increased infiltration of effector and cytotoxic T lymphocytes after local delivery of anti–PD-1

To determine the effect of anti–PD-1 treatment on the activation of the immune system, we stained tissue sections of oral lesions induced by 4NQO to detect the infiltration of CD4+ and CD8+ T cells. We observed that wild-type and mutant p53 mice treated with IgG isotype control showed similar infiltration of CD4+ T cells in low- and high-grade oral lesions (Fig. 2A). This was surprising because we assumed that oncogenic p53R172H might prevent infiltration of immune cells in the tumor microenvironment. However, an increased infiltration of CD4+ T cells was detected after PD-1 gel treatment in low- and high-grade oral lesions in wild-type and mutant p53R172H mice. As previously observed, there was no difference of CD4+ T-cell infiltration between wild-type and mutant p53 mice.

Next, immunostaining of CD8+ T cells showed that local delivery of PD-1 was associated with increased infiltration in low- and high-grade oral dysplasia demonstrating the efficacy of immune cells to infiltrate tumorigenic lesions (Fig. 2B). Interestingly, CD8+ T-cell infiltration was not affected by mutant p53R172H expressed in oral dysplasia or carcinoma, suggesting that oncogenic mutant p53 might affect an alternative pathway to disrupt tumor immunity in oral cancer.

Foxp3 regulatory T-cell infiltration is altered in response to PD-1 blockade

To explore mechanisms of immunosuppression in OPLs, first we explored the levels of the immune-checkpoint inhibitor PD-1. Interestingly, after local delivery of anti–PD-1, we observed a significant reduction levels of PD-1 in low- and high-grade oral lesions; this was confirmed by immunostaining of PD-1 in oral lesions, in which high levels are detected in control-treated mice (Fig. 3). These results suggest that local delivery by hydrogels maintains anti–PD-1 antibody activity and the slow release by a single dose shows significant impact in the oral lesion microenvironment, which alters the infiltration of different immune cells.

Regulatory T cells (Treg) are suppressors of antitumor responses by disrupting maturation of dendritic cells (DC) and preventing activation of CD4+ effector and CD8+ cytotoxic cells in the tumor microenvironment (38). To assess the role of PD-1 blockade on Treg infiltration in oral lesions, we conducted an IHC analysis with specific antibodies to Foxp3, a specific Treg marker. First, we noted that low-grade dysplasia has infiltration of Foxp3+ Tregs; nevertheless, it is significantly higher in premalignant lesions expressing mutant p53R172H, and strongly indicating that oncogenic activity of p53 might influence the environment to promote a higher infiltration of immune suppressor cells (Fig. 4). Similar results were observed
in high-grade lesions (severe dysplasia and carcinoma). Furthermore, PD-1 gel treatment significantly reduced the number of Foxp3\(^{+}\) T cells in both low- and high-grade dysplasia, indicating that disruption of the PD-1/PD-L1 immunosuppression axis prevents infiltration of Foxp3\(^{+}\) cells.

**Local delivery of anti-PD-1 antibody restores STING expression and infiltration of CD11c DCs**

The mechanism by which the immune system is alerted to the presence of a developing malignant lesions is by the classic “danger signals” such as type I interferons (IFN), which are induced early during tumor development and mediated by the cGAS–STING pathway (39–41). These cytokines activate DCs and promote induction of adaptive CD4 and CD8 T-cell antitumor immune responses (42). A defect or genetic alterations in the cGAS–STING pathway will impair interferon secretion preventing DC maturation, allowing transformation of premalignant cells to a tumorigenic phenotype. We analyzed the expression levels of STING by IHC, an important activator of TANK-binding kinase 1 (TBK1), which initiates the downstream signaling to produce type I interferons. In the IgG control–treated mice, the levels of STING were significantly lower in oral lesions expressing mutant p53R172H compared with wild-type p53. Interestingly, after PD-1 treatment, we...
observed a remarkable expression of STING in the low- and high-grade oral lesions of p53 wild-type mice (Fig. 5). In addition, we observed modest levels of STING staining in the oral dysplasia and carcinomas in mutant p53 mice, which was significantly lower than the intensity of STING staining when compared with wild-type mice (Fig. 5). The normal tongue epithelium exposed to 4NQO did not show any expression levels of STING; it was detected only in the OPLs (Supplementary Fig. S5).

Furthermore, we stained oral lesions for CD11c, a specific DC marker. Strikingly, PD-1 inhibition was associated with a significantly higher degree of DC infiltration in the low- and high-grade oral dysplasia of wild-type p53 mice, which was significantly lower than the intensity of STING staining when compared with wild-type mice (Fig. 5). The normal tongue epithelium exposed to 4NQO did not show any expression levels of STING; it was detected only in the OPLs (Supplementary Fig. S5).

Discussion

We used a 4NQO oral carcinogenesis mouse model that allows the development of OPLs and oral cancers which consistently recapitulates histologic abnormalities observed in human carcinogenesis to examine the impact of a locally administered anti–PD-1 antibody-loaded hydrogel on the development of oral neoplastic lesions (31, 43). In our study, histopathologic analysis demonstrated that local delivery of loaded hydrogels with anti–PD-1 reduced the incidence of OPLs and carcinoma in p53 wild-type mice. Likewise, we observed a modest reduction of oral lesions in mutant p53R172H
mice, suggesting that oncogenic p53 activates mechanisms of resistance to an inhibitor of the PD-1 immune checkpoint. Mice expressing mutant p53R172H have a higher incidence of high-grade lesions, strongly suggesting that oncogenic p53 has a role in immunosuppressive mechanisms that enable neoplastic progression.

In this study, we also examined the efficacy of local immunotherapy delivery by loading anti–PD-1 in nanofibrous biomaterials called MDP hydrogels that consist of polymerized multipeptide domain macromolecules. As previously reported, the MDP hydrogels mimic the native extracellular matrix of the body, and prolong the release of small-molecule drugs to achieve long-term drug delivery, and minimize side effects through localized and site-specific targeting of drug (36). We speculate that injection of this biomaterial into the tongue of mice generates an inflammatory response that might favor...
infiltration of immune cells, and together with the effect of the PD-1 blockade prevents Tregs to inactivate CD4 effector and CD8 cytotoxic T cells in the oral malignant lesions.

We observed an increased infiltration of CD4⁺ and CD8⁺ T cells in low- and high-grade lesions after PD1 local blockade. We previously observed similar findings using the 4NQO carcinogen model in heterozygous p53 mice with loss of one p53 wt allele, with systemic administration (eight intraperitoneal injections) of anti–PD-1 antibodies, which led to an increased infiltration of activated T cells (22). Other studies using anti–PD-1 in mouse oral cancer models observed similar results in which PD1 blockade reduces OPL incidence and increase production of CD4⁺ and CD8⁺ T cells in the spleen and lymph nodes and circulation levels (24); or showed a temporal increase in CD4⁺ and CD8⁺ T cells in mice treated with anti–PD-1 in secondary lymph nodes (23).

Figure 5.
Local delivery of anti–PD-1 increases STING protein levels in low- and high-grade lesions. Representative IHC images of STING expression in oral lesions. A, significant difference of STING expression was detected between untreated and PD-1 treated mice groups. Immunostaining signal was defined as the number of positive cells per mm². Right graphs: **, P < 0.01; *** P < 0.001; **** P < 0.0001. Scale bar, 50 µm.

Figure 6.
PD-1 blockage promoted recruitment of CD11c⁺ DCs into oral lesions of wild-type and mutant p53 mice. Representative IHC images of CD11c⁺ DCs in oral lesions. A significant difference of CD11c⁺ cells was detected between untreated and PD1-treated mice groups. Immunostaining signal was defined as the number of positive cells per mm². Right graphs: **, P < 0.01; *** P < 0.001; **** P < 0.0001. Scale bar, 50 µm.
In aggregate, these findings indicate that early genomic alteration in the p53 gene of oral epithelial cells promotes immunosuppressive pathways that disrupt antitumor immunity mechanisms, preventing the activation of adaptive immune response. Interestingly, there were no significant differences in the numbers of CD4 and CD8 T lymphocytes in oral dysplasia or carcinoma in mutant and wild-type p53 mice, suggesting that immunosuppressive mechanisms might qualitatively impair the effector and cytotoxic function of infiltrated T cells, rather than decrease their number.

Tregs (CD4\(^+\), CD25\(^+\), Foxp3\(^+\)) negatively regulate tumor immunity, leading to tumor growth in mice through multiple suppressive mechanisms (44–46). The transcription factor Foxp3 is a phenotypic marker that correlates with tumor immunosuppression and worse prognosis (46, 47). Moreover, studies have shown that anti-CD25 immunotherapy can induce tumor rejection in mice and humans. In our study, we observed high infiltration of Foxp3\(^+\) Tregs in low-grade OPLs, with significantly higher numbers of Treg in oral lesions expressing mutant p53\(^{R172H}\), indicating that Tregs may be important immune cell population in early as well as later stages of oral neoplastic progression. Moreover, hydrogel/anti-PD-1 treatment dramatically reduced Foxp3\(^+\) Tregs in oral dysplasia and carcinoma, suggesting that local delivery and slow antibody release can provide an efficient therapy to block PD-1/PD-L1 interaction. A recent study using an orthotopic oral cancer mouse model showed that a lower dose of intratumoral of immunotherapy showed similar effects as systemic delivery of a higher dose. Furthermore, intratumoral injection led to higher distribution of the antibody in the tumor and cervical lymph nodes, but less in the spleen, a secondary lymphoid organ. This approach enhanced the recruitment and infiltration lymphoid and myeloid immune cells in the tumor microenvironment, resulting in a significant tumor volume reduction. These results demonstrate that local delivery of immunotherapeutic agents might represent a novel approach to improve efficacy with favorable tumor response (48).

The elimination phase of cancer immunoeediting is best described as an updated version of cancer immunosurveillance, in which the innate and adaptive immune systems work together to detect the presence of a developing tumor and destroy it before it becomes highly immunosuppressive. One of the mechanisms by which the immune system is alerted to the presence of a developing tumor is the production and secretion of type I interferons, which is regulated by the cGAS-STING pathway (39). In our study, OPLs with mutant p53\(^{R172H}\) showed a dramatic decrease in expression of STING, a critical regulatory molecule in the activation of interferon production. PD-1 blockade rescued the STING expression to a greater degree in lesions expressing wild-type p53 as compared with mice expressing oncogenic p53, providing direct evidence of the role of mutant p53\(^{R172H}\) in OPL progression, and suggest that STING expression is relatively low in the early stages of oral cancer development and throughout cancer progression in mutant p53\(^{R172H}\) mice.

Interestingly, a single dose of local immune-checkpoint inhibitory antibody delivery (anti-PD-1–hydrogel) also promotes a significantly higher number of DCs infiltrating into oral dysplasia, especially in wild-type p53 mice compared with mutant p53\(^{R172H}\) mice. DCs have a critical role in linking innate and adaptive immunity. They have a key role in capturing and presenting antigens in the form of MHC II receptors to T cells, stimulating them to develop into effector T cells. In this role, DCs have a significant role in the tumor microenvironment, surrounding tumors and providing signals to other immune cells that stimulate or suppress T-cell activation. Tumor-tolerant DCs are one reason that nascent tumors can escape immune surveillance (49). DCs can also have prognostic value in treating patients. Potent DCs are required for a strong antitumor immune response after chemotherapy or radiation. There is a great degree of variability in the location and number of DCs in tumors (50). Studies have shown that tumors with DCs expressing low levels of CD86 and greater levels of IL10, generally are resistant to therapy and have a poor outcome (51). All of these reasons make DC stimulation or prevention of DC tolerance potential good strategies for tumor immune prevention.

Although ICIs can induce robust antitumor immune responses, their systemic delivery can induce cytokine release syndrome and abnormal liver function (52). To minimize off-tissue effects, biomaterials like multipeptide domain hydrogels are delivery systems designed for local and sustained release in vivo as demonstrated in this and other studies (25). Therefore, we have developed a novel method to locally deliver immunotherapy in a mouse oral cancer model using a novel biomaterial (hydrogel). The novel local anti-PD-1 delivery is relevant because a single dose using biomaterials that regulate a slow antibody release for weeks reduces the incidence of lesions to progress into carcinomas. We have demonstrated that a single PD-1 gel delivery shows similar therapeutic effect as eight doses of anti–PD-1 by systemic delivery. This approach could eliminate the need for repeated systemic administration of the anti–PD-1 immune-checkpoint inhibitory antibody. In clinical practice, local immunotherapy administration in patients with OPLs could potentially decrease the cost and reduce unnecessary and undesirable systemic side effects in patients with oral leukoplakia.

Authors' Disclosures
D.G. Leach reports grants from National Science Foundation during the conduct of the study; in addition, D.G. Leach has a patent for hydrogel delivery of Sting immunotherapy for treatment of cancer, serial no. 62/520,834 pending. S. Young reports a patent for 62/950,718 pending and a patent for 62/520,834 pending. A.G. Sikora reports other support from SQZ, other support from Pelican, and other support from Tessa Therapeutics outside the submitted work. J.D. Hartgerink reports a patent 20200146975 pending. No disclosures were reported by the other authors.

Authors' Contributions
Y. Shi: Formal analysis, validation, investigation, methodology, writing—original draft. T. Xie: Conceptualization, validation, investigation, and

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methodology. D.G. Leach: Resources, investigation, methodology, and writing-review and editing. B. Wang: Investigation and methodology. S. Young: Conceptualization, resources, writing-review and editing. A.A. Osman: Investigation, methodology, writing-review and editing. A.G. Sikora: Conceptualization, investigation, writing-review and editing. X. Ren: Investigation. J.D. Hartgerink: Conceptualization, resources, supervision, investigation, methodology, writing-review and editing. J.N. Myers: Conceptualization, resources, formal analysis, supervision, funding acquisition, investigation, writing-original draft, and project administration. R. Rangel: Conceptualization, formal analysis, supervision, validation, investigation, methodology, writing-original draft, project administration, writing-review and editing.

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