

Perspective

Perspective on Checkley et al., p. 1011

Rapamycin and mTORC1 Inhibition in the Mouse: Skin Cancer Prevention

Mohammad Athar¹ and Levy Kopelovich²

Abstract

Therapeutic and preventive effects of rapamycin include reduced risk of nonmelanoma skin cancer (NMSC). In this issue of the journal (beginning on page 1011), Checkley and colleagues report that rapamycin inhibits mTOR complex 1 in murine epidermis, thereby inhibiting tumor promotion mediated by tetradecanoyl phorbol-13 acetate in association with a strong anti-inflammatory effect. Rapamycin is an immunosuppressive drug for preventing graft rejection in organ transplant recipients and reduces the risk of NMSC and Kaposi's sarcoma in this population, albeit by mechanisms distinct from immunosuppression. Important future directions include identifying molecular predictors of rapamycin/rapalog sensitivity or resistance (potentially, for example, PI3K pathway alterations and *KRAS* mutations) and combined non-rapalog, mTOR-targeting approaches, all of which should increase efficacy and minimize toxicity. *Cancer Prev Res*; 4(7); 957–61. ©2011 AACR.

The lifetime risk for skin cancer exceeds the lifetime risk of all other cancers combined (1, 2). It is estimated that more than 1.2 million new cases of nonmelanoma skin cancer (NMSC), including squamous cell carcinoma (SCC) and basal cell carcinoma, are diagnosed annually in the United States (1). The risk for NMSC is greatly increased in chronically immune-suppressed organ transplant recipients (3). The molecular mechanisms involved in the pathogenesis of NMSC are not fully understood.

SCC pathogenesis is characterized by the inhibition of apoptosis and enhancement of cell proliferation, resulting in the recruitment of initiated premalignant cells during progression to cancer (4, 5). In murine skin, sustained activation of AKT because of exposure to tumor-promoting agents such as tetradecanoyl phorbol-13 acetate (TPA) and ultraviolet B (UVB) has been shown to cause benign lesions (papillomas), which progress at a certain frequency to SCC. AKT activation is accompanied by increased phosphorylation of its effector molecules including mTOR, glycogen synthase kinase 3 beta (GSK3 β), and BAD, suggesting the possibility that AKT and its downstream signaling pathways are involved in skin tumor promotion (6–8). Direct support for AKT involvement comes from studies showing that epidermal overexpression of wild-type AKT or a

constitutively active form of AKT (myristoylated-AKT) enhances susceptibility to chemically induced skin cancer (8). A major mechanism by which AKT can augment skin cancer is activation of mTOR, a serine/threonine protein kinase that regulates cell growth and proliferation by enhancing both Cap-dependent and 5'-terminal oligopyrimidine (TOP) mRNA-dependent protein synthesis (9, 10).

mTOR functions intracellularly as a physiologic sensor of nutrients, regulating metabolism and growth (10) and is the catalytic subunit comprising the distinct complexes mTOR complex 1 (mTORC1) and mTORC2. These complexes are distinguished by unique accessory proteins, regulatory associated protein of mTOR (RAPTOR) for mTORC1 and rapamycin-insensitive companion of mTOR (RICTOR) for mTORC2. The two mTOR pathways represent an intricate network with multiple feedback loops. In brief, mTORC1 immediate substrates S6 kinase 1 (S6K1) and eukaryotic translation initiation factor 4E (eIF4E)-binding protein 1 (4E-BP1) associate with mRNA and regulate mRNA translation and initiation, thereby controlling the rate of protein synthesis (10–13). Whereas unphosphorylated 4E-BP1 suppresses mRNA translation, phosphorylated 4E-BP1 (by mTORC1) disassociates from eIF4E and allows it to recruit the translation initiation factor eIF4G to the 5' end of mRNAs. Phosphorylation of S6K1 by mTORC1 promotes mRNA translation and, in turn, phosphorylates or binds multiple proteins including eukaryote elongation factor 2 kinase (eEF2K), S6K1 Aly/REF-like target (SKAR), 80-kDa nuclear cap-binding protein (CBP80), and eIF4B (14). mTORC1 also controls the activity of several transcription factors involved in lipid synthesis and mitochondrial metabolism (10, 15). Dysregulated mTORC1 signaling in humans is associated with the Peutz-Jeghers syndrome (due to LKB1-inactivating mutations) and with colorectal cancers having adenomatous

Authors' Affiliations: ¹Department of Dermatology, Skin Diseases Research Center and UAB Comprehensive Cancer Center, University of Alabama at Birmingham, Birmingham, Alabama; and ²Division of Cancer Prevention, National Cancer Institute, Bethesda, Maryland

Corresponding Author: Levy Kopelovich, NIH, National Cancer Institute, DCP, Executive Plaza North, Suite 2114, 6130 Executive Boulevard, Bethesda, MD 20892-7322. Phone: 301-594-0467; Fax: 301-402-0553; E-mail: kopelovl@mail.nih.gov

doi: 10.1158/1940-6207.CAPR-11-0266

©2011 American Association for Cancer Research.

Table 1. Summary of animal studies supporting the role of rapamycin-mediated inhibition of mTOR in cancer therapy/prevention

Cancer type	Description of effects	References
Skin	Reduction in tumor numbers, tumor size, and progression of benign lesions to SCCs in UVB-induced SKH-1 hairless, DMBA-TPA 2-stage and cyclosporine A-mediated carcinogenesis in various murine models	22, 26, 38–41
Head and neck	Reduction in the oral carcinogen-specific mouse model, human xenograft, and oral-specific Kras/p53 two-hit carcinogenesis murine models	22, 42–44
Gastrointestinal (liver, intestine, stomach, and pancreas)	Inhibition of hepatocellular carcinoma in xenograft and orthotopic murine models; reduction in intestinal polyp formation and mortality in Apc Δ 716 engineered mice; inhibition of gastric cancer in xenograft immune deficient mouse models; mimicked the anticancer effects of calorie restriction in a murine model of pancreatic cancer	28, 36, 45–47
Kidney	Inhibition of tumor growth in A/J Tsc2 ^{+/-} mice and xenograft models	48, 49
Breast	Inhibition of tumor growth of premalignant and malignant lesions in ductal carcinoma <i>in situ</i> and Erb2-dependent transgenic breast cancer mouse models	50, 51
Prostate	Diminution of prostate adenocarcinoma in Pten ^{+/-} and Pten ^{+/-} /Tsc2 ^{+/-} mice.	52
Bladder	Prevention of invasive bladder cancer in a murine model carrying bladder epithelium-specific deletion of p53 and Pten.	53
Rhabdomyosarcoma	Lack of published studies in murine models, however, a clear benefit of combinatorial chemotherapy in humans	54
Lung	Inhibition of benzo(a)pyrene-, tobacco carcinogen (NNK)-induced lung tumorigenesis in A/J as well as in Tsc1/Kras double-mutant mice	23, 55, 56
Anal	Rapamycin slowed the growth of anal cancer both in a murine model and in humans	57
Uterine	Eker rat model of uterine leiomyomata faithfully recapitulates the human transcriptional profile of this tumor. In this model, mTOR activation (followed by IRS-1 phosphorylation) occurs early, or in endometrial hyperplasia, and the rapamycin analogue WAY-129327 significantly decreased hyperplasia, proliferation, and tumor incidence, multiplicity, and size	58

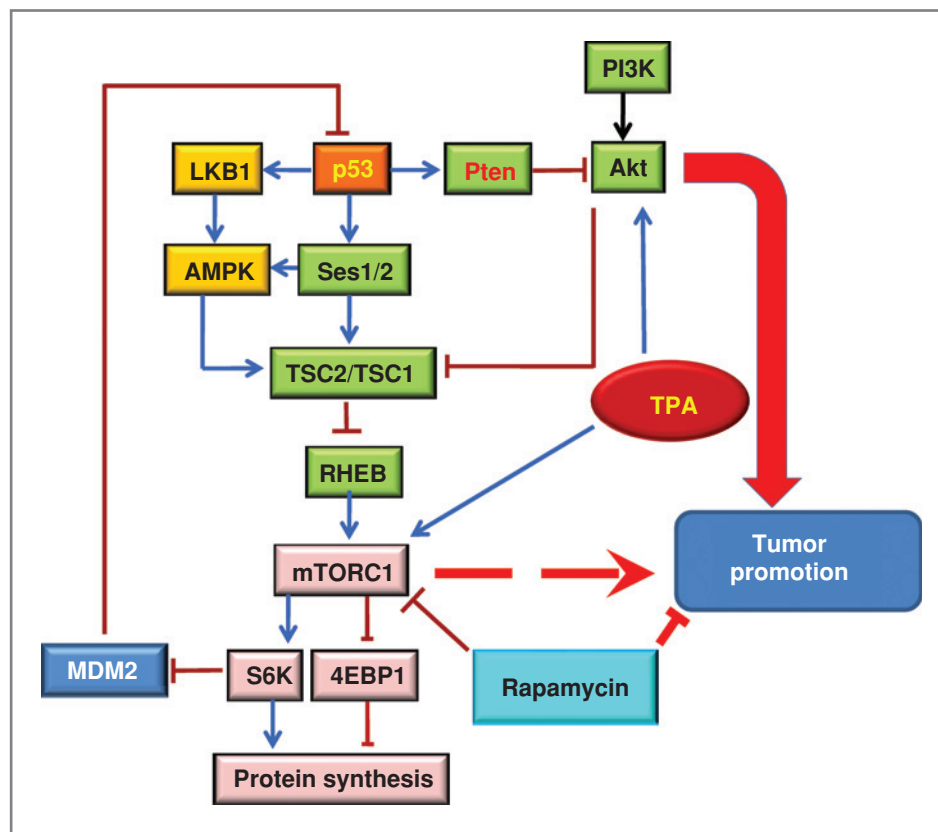
Abbreviations: DMBA, 7,12-dimethylbenz[α]anthracene; NNK, nicotine-derived nitrosamine ketone; IRS-1, insulin-receptor substrate 1.

polyposis coli (APC) loss-of-function mutations (10). Tuberous sclerosis complexes 1 and 2 (TSC1 and TSC2) function as tumor suppressors and are implicated in kidney cancer (9), presumably through their effect on the mTOR signaling pathway. The TSC1–TSC2 complex acts as a GTPase-activating protein. Following phosphorylation by 5'-AMP-activated protein kinase (AMPK), TSC2 stimulates Ras homologue enriched in brain (RHEB), thereby inhibiting mTORC1 signaling. TSC2 also can be phosphorylated by AKT or GSK3 β , which negatively regulates mTORC1, providing additional evidence for the existence of multiple regulatory mechanisms of the mTOR signaling pathway and cell growth.

mTORC2 is associated with the regulation of cytoskeletal organization and cell polarization. mTORC2 phosphorylates and activates AKT through a feedback loop [at serine 473 (Ser473)], including serum- and glucocorticoid-regu-

lated kinase and protein kinase C (PKC), thus playing an important role in regulating cell survival, cell-cycle progression, and anabolism. After phosphorylation at Ser478, AKT is primed for further phosphorylation at threonine 308 (Thr308), which leads to its full activation. TPA is a potent inducer of PKC in this context, enhancing phosphorylation of AKT at both Ser473 and Thr308. Rapamycin inhibits mTORC1, but chronic use of this drug may also inhibit mTORC2 (16). Rapamycin binds to a 12-kDa FK506-binding protein, also known as prolyl isomerase (PPIase), thereby inhibiting the ability of mTORC1 to phosphorylate its downstream effectors. mTORC1 inhibition by rapamycin or other rapalogs has antineoplastic effects, suggesting that mTORC1 is a relevant common effector protein complex involved in the pathogenesis of various cancers (Table 1). Specifically, epidermal transgenic expression of RHEB, a direct activator of mTORC1, sensitized murine

Figure 1. mTOR signaling cascade in the pathogenesis and prevention of skin cancer. The skin tumor promoter TPA induces mTORC1 activity, which in turn enhances the AKT-dependent cell survival signaling pathway. TPA-mediated mTORC1 activation augments epidermal hyperplasia and dermal inflammation. Rapamycin inhibits TPA-mediated inflammation and tumor promotion by inhibiting mTORC1 activity. MDM2, murine double minute 2.



skin to SCC induction (17). The phosphoinositide 3-kinase (PI3K)/AKT/mTOR signaling pathway is also activated by mutations in the p110 α subunit of *PI3K* called *PI3CA*, and preclinical investigations suggest that *PI3CA* mutations may predict the clinical outcome of PI3K/AKT/mTOR-axis inhibitors (18).

In this issue of the journal, a report by Checkley and colleagues further expands our understanding of the mechanisms by which rapamycin inhibits phorbol ester-mediated skin tumor promotion (19). These investigators showed that the inhibition by rapamycin of TPA-induced mTORC1 [characterized by enhanced phosphorylation of mTOR (Ser2448), p70S6K (Thr389), p4E-BP1 (Ser65 and Thr37/46), pS6-ribosomal protein (Ser240/244), and AKT (Thr308, Ser473)] resulted in a significant reduction in the epidermal labeling index as well as TPA-induced hyperplasia. Of importance, these events were followed by a significant decrease in dermal inflammation that was associated with decreased infiltration of inflammatory cells (T cells, macrophages, neutrophils, and mast cells). Chronic inflammation is a known risk factor for various human cancers, including skin cancer (20). These data suggest that mTORC1 is a key regulator of proliferation and inflammation at the site of TPA treatment of mouse skin. Presumably, therefore, rapamycin inhibits expression of multiple cytokines/chemokines and their receptors, known to promote proliferation and inflammation responses (21). However, the anti-inflammatory effects

of rapamycin on macrophage and T-cell infiltration may depend on the type of treatment protocols, including routes of administration (22, 23).

An important finding in this article was that, contrary to its inhibitory effects at lower doses on AKT phosphorylation, rapamycin increased AKT phosphorylation at both Ser473 and Thr308 when given at 200 nmol in multiple administrations. Although the mechanisms for these observations are unclear, the authors suggest that an mTOR-dependent negative feedback loop may be responsible. It is conceivable, however, that the effects of rapamycin on cell survival signaling are associated with a biphasic dose response which involves distinct mechanisms. A clarification of this as yet unresolved issue would require the demonstration that rapamycin-mediated inhibition of AKT phosphorylation at both Ser473 and Thr308 also occurs through its inhibition of mTORC2 and that rapamycin is able to inhibit TPA-induced PKC (vide supra). Loss of RICTOR results in a complete loss of AKT phosphorylation at Ser473, leading to suppression of phosphorylation of the transcription factors forkhead box O 1 (FOXO1) and FOXO3 but not of its other substrate TSC2 (tuberin). Apparently, mTORC2 favors cell survival through AKT-mediated inhibition of FOXO1 and FOXO3, which are known to activate genes that promote apoptosis (10, 20). This year, Zinzalla and colleagues provided evidence that mTORC2 is activated by its direct physical association with the ribosomal machinery which ensures

that mTORC2 activity is calibrated to complement the intrinsic growth capacity of the cell (24, 25).

Of importance, Checkley and colleagues also observed that rapamycin is a potent inhibitor of TPA-mediated tumor promotion in murine skin (Fig. 1). It abrogates papilloma formation and reduces the progression of papillomas to SCC (19). Similar anticarcinogenic effects of rapamycin (Table 1) were previously reported in UVB-induced photocarcinogenesis and 2-stage chemical carcinogenesis murine models (22, 26). Consistent with the observations of Checkley and colleagues, rapamycin has preventive and therapeutic effects on cancer in humans (27–31). A significant clinical effect of rapamycin treatment is its ability to delay the development of premalignancies and to reduce the incidence of new NMSCs in organ transplant recipients (3).

In conclusion, by using the TPA-induced skin cancer model in mice, Checkley and colleagues have provided new insights into the potential utility of rapalogs in preventing and treating environmentally induced skin cancer. Their data indicate that signaling through mTORC1 contributes significantly to inflammation and ultimately to skin tumor promotion (Fig. 1). On the basis of the understanding of compensatory feedback loops between mTORC1 and mTORC2, second-generation inhibitors of this pathway have been developed (32). These agents simultaneously inhibit both mTOR complexes, and some of them also inhibit PI3K, which is a key player in regulating this feedback loop (18). mTOR inhibitors require a word of caution, however: despite their strong effect in preventing cancer in both animals and humans, their chronic use in humans is associated with a number of side effects which often require discontinuation of therapy

and are a major concern in the cancer prevention setting (3, 18, 33). Metformin and other biguanides may have a role in addressing this concern. These agents are widely used in treating diabetes, where they have a tolerable risk profile, and recent data suggest that they may be effective cancer chemoprevention agents (34, 35). *In vivo*, metformin downregulates mTORC1 via several potential pathways, including AMPK-dependent and -independent mechanisms (34, 35). Important data show that caloric restriction partially mimics the anticancer effects of rapamycin in a murine model of pancreatic cancer (36). Substantial data on potential predictive markers of mTOR-inhibitor efficacy for cancer therapy and prevention have highlighted the predictive potential of KRAS mutations and markers of PI3K/AKT pathway activation (37). Predictive markers and combinations of non-rapalog approaches for targeting mTOR should increase efficacy and minimize toxicity. A specific cancer prevention approach combining metformin and caloric restriction modalities as part of an mTOR-targeted regimen should prove useful, particularly because safety is such a critical factor for chemoprevention trials.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Grant Support

This work was supported in part by 1R01CA138998, R21ES017494, NCI N01-CN-43300 and P30AR050948.

Received March 17, 2011; revised May 9, 2011; accepted May 24, 2011; published online July 6, 2011.

References

- Neville JA, Welch E, Leffell DJ. Management of nonmelanoma skin cancer in 2007. *Nat Clin Pract Oncol* 2007;4:462–9.
- Jemal A, Siegel R, Xu J, Ward E. Cancer statistics, 2010. *CA Cancer J Clin* 2010;60:277–300.
- Athar M, Walsh SB, Kopelovich L, Elmets CA. Pathogenesis of nonmelanoma skin cancers in organ transplant recipients. *Arch Biochem Biophys* 2011;508:159–63.
- Bickers DR, Athar M. Oxidative stress in the pathogenesis of skin disease. *J Invest Dermatol* 2006;126:2565–75.
- Narayanan DL, Saladi RN, Fox JL. Ultraviolet radiation and skin cancer. *Int J Dermatol* 2010;49:978–86.
- Segrelles C, Ruiz S, Perez P, Murga C, Santos M, Budunova IV, et al. Functional roles of Akt signaling in mouse tumorigenesis. *Oncogene* 2002;21:53–64.
- Lu J, Rho O, Wilker E, Beltran L, Digionvanni J. Activation of epidermal Akt by diverse mouse skin tumor promoters. *Mol Cancer Res* 2007;5:1342–52.
- Segrelles C, Lu J, Hammann B, Santos M, Moral M, Cascallana JL, et al. Deregulated activity of Akt in epithelial basal cells induces spontaneous tumors and heightened sensitivity to skin carcinogenesis. *Cancer Res* 2007;67:10879–88.
- Vignot S, Faivre S, Aguirre D, Raymond E. mTOR-targeted therapy of cancer with rapamycin derivatives. *Ann Oncol* 2005;16:525–37.
- Zoncu R, Efeyan A, Sabatini DM. mTOR: from growth signal integration to cancer, diabetes and ageing. *Nat Rev Mol Cell Biol* 2011;12:21–35.
- Facchinetti V, Ouyang W, Wei H, Soto N, Lazorchak A, Gould C, et al. The mammalian target of rapamycin complex 2 controls folding and stability of Akt and p190Rheb. *EMBO J* 2008;27:1932–43.
- Garcia-Martinez JM, Alessi DR. mTOR complex 2 (mTORC2) controls hydrophobic motif phosphorylation and activation of serum- and glucocorticoid-induced protein kinase 1 (SGK1). *Biochem J* 2008;416:375–85.
- Ikenoue T, Inoki K, Yang Q, Zhou X, Guan KL. Essential function of TORC2 in PKC and Akt turn motif phosphorylation, maturation and signaling. *EMBO J* 2008;27:1919–31.
- Dorrello NV, Peschiaroli A, Guardavaccaro D, Colburn NH, Sherman NE, Pagano M. S6K1- and betaTRCP-mediated degradation of PDCD4 promotes protein translation and cell growth. *Science* 2006;314:467–71.
- Gwinn DM, Shackelford DB, Egan DF, Mihaylova MM, Mery A, Vasquez DS, et al. AMPK phosphorylation of raptor mediates a metabolic checkpoint. *Mol Cell* 2008;30:214–26.
- Sarbassov DD, Ali SM, Sengupta S, Sheen JH, Hsu PP, Bagley AF, et al. Prolonged rapamycin treatment inhibits mTORC2 assembly and Akt/PKB. *Mol Cell* 2006;22:159–68.
- Lu ZH, Shvartsman MB, Lee AY, Shap JM, Murray MM, Kladney RD, et al. Mammalian target of rapamycin activator RHEB is frequently overexpressed in human carcinomas and is critical and sufficient for skin epithelial carcinogenesis. *Cancer Res* 2010;70:3287–98.
- Janku F, Tsimberidou AM, Garrido-Laguna I, Wang X, Luthra R, Hong DS, et al. PIK3CA mutations in patients with advanced cancers treated

- with PI3K/AKT/mTOR axis inhibitors. *Mol Cancer Ther* 2011;10:558–65.
19. Checkley LA, Rho O, Moore T, Hursting S, Digiovanni J. Rapamycin is a potent inhibitor of skin tumor promotion by 12-O-tetradecanoyl-phorbol-13-acetate. *Cancer Prev Res* 2011;4:1011–20.
 20. Aggarwal BB, Vijayalekshmi RV, Sung B. Targeting inflammatory pathways for prevention and therapy of cancer: short-term friend, long-term foe. *Clin Cancer Res* 2009;15:425–30.
 21. Hansell CA, Simpson CV, Nibbs RJ. Chemokine sequestration by atypical chemokine receptors. *Biochem Soc Trans* 2006;34:1009–13.
 22. Amornphimoltham P, Leelahavanichkul K, Molinolo A, Patel V, Gutkind JS. Inhibition of mammalian target of rapamycin by rapamycin causes the regression of carcinogen-induced skin tumor lesions. *Clin Cancer Res* 2008;14:8094–101.
 23. Granville CA, Warfel N, Tsurutani J, Hollander MC, Robertson M, Fox SD, et al. Identification of a highly effective rapamycin schedule that markedly reduces the size, multiplicity, and phenotypic progression of tobacco carcinogen-induced murine lung tumors. *Clin Cancer Res* 2007;13:2281–9.
 24. Zinzalla V, Stracka D, Oppliger W, Hall MN. Activation of mTORC2 by association with the ribosome. *Cell* 2011;144:757–68.
 25. Xie X, Guan K-L. The ribosome and TORC2: collaborators for cell growth. *Cell* 2011;144:640–2.
 26. Wulff BC, Kusewitt DF, VanBuskirk AM, Thomas-Ahner JM, Jason DF, Oberszyn TM. Sirolimus reduces the incidence and progression of UVB-induced skin cancer in SKH mice even with co-administration of cyclosporine A. *J Invest Derm* 2008;128:2467–73.
 27. Dennis PA. Rapamycin for chemoprevention of upper aerodigestive tract cancers. *Cancer Prev Res* 2009;2:7–9.
 28. Fujishita T, Aoki K, Lane HA, Aoki M, Takeoto MM. Inhibition of the mTORC1 pathway suppresses intestinal polyp formation and reduces mortality in *ApcDelta716* mice. *Proc Acad Sci USA* 2008;105:13544–49.
 29. Kudo M. Current status of molecularly targeted therapy for hepatocellular carcinoma: clinical practice. *Int J Clin Oncol* 2010;15:242–55.
 30. Yao JC, Lombard-Bohas C, Baudin E, Kvols LK, Rougier P, Ruzsniowski P, et al. Daily oral everolimus activity in patients with metastatic pancreatic neuroendocrine tumors after failure of cytotoxic chemotherapy: a phase II trial. *J Clin Oncol* 2010;28:69–76.
 31. Kulke MH, Bergsland EK, Yao JC. Glycemic control in patients in insulinoma treated with everolimus. *N Engl J Med* 2009;360:195–7.
 32. Vilar E, Perez-Garcia J, Tabernero J. Pushing the envelope in the mTOR pathway: The second generation of inhibitors. *Mol Cancer Ther* 2011;10:395–403.
 33. Kopelovich L, Fay JR, Sigman C, Crowell J. The mammalian target of rapamycin pathway as a potential target for cancer chemoprevention. *Cancer Epidemiol Biomarkers Prev* 2007;16:1330–40.
 34. Engelman JA, Cantley LC. Chemoprevention meets glucose control. *Cancer Prev Res* 2010;3:1066–76.
 35. Memmott RM, Mercado JR, Maier CR, Kawabata S, Fox SD, Dennis PA. Metformin prevents tobacco carcinogen-induced lung tumorigenesis. *Cancer Prev Res* 2010;3:1066–76.
 36. Lashinger LM, Malone LM, Brown GW, Daniels EA, Goldberg JA, Otto G, et al. Rapamycin partially mimics the anticancer effects of calorie restriction in a murine model of pancreatic cancer. *Cancer Prev Res* 2011;4:1041–51.
 37. Di Nicolantonio F, Arena S, Tabernero J, Grosso S, Molinari F, Macarulla T, et al. Deregulation of the PI3K and KRAS signaling pathways in human cancer cells determines their response to everolimus. *J Clin Invest* 2010;120:2858–66.
 38. Fernández A, Marcén R, Pascual J, Galeano C, Ocaña J, Arellano EM, et al. Conversion from calcineurin inhibitors to everolimus in kidney transplant recipients with malignant neoplasia. *Transplant Proc* 2006;38:2453–5.
 39. Wulff BC, Kusewitt DF, VanBuskirk AM, Thomas-Ahner JM, Duncan FJ, Oberszyn TM. Sirolimus reduces the incidence and progression of UVB-induced skin cancer in SKH mice even with co-administration of cyclosporine A. *J Invest Dermatol* 2008;128:2467–73.
 40. de Gruijl FR, Voskamp P. Photocarcinogenesis—DNA damage and gene mutations. *Cancer Treat Res* 2009;146:101–8.
 41. de Gruijl FR, Koehl GE, Voskamp P, Strik A, Rebel HG, Gaumann A, et al. Early and late effects of the immunosuppressants rapamycin and mycophenolate mofetil on UV carcinogenesis. *Int J Cancer* 2010;12:796–804.
 42. Amornphimoltham P, Patel V, Sodhi A, Nikitakis NG, Sauk JJ, Sausville EA, et al. Mammalian target of rapamycin, a molecular target in squamous cell carcinomas of the head and neck. *Cancer Res* 2005;65:9953–61.
 43. Czerninski R, Amornphimoltham P, Patel V, Molinolo AA, Gutkind JS. Targeting mammalian target of rapamycin by rapamycin prevents tumor progression in an oral-specific chemical carcinogenesis model. *Cancer Prev Res* 2009;2:27–36.
 44. Raimondi AR, Molinolo A, Gutkind JS. Rapamycin prevents early onset of tumorigenesis in an oral-specific K-ras and p53 two-hit carcinogenesis model. *Cancer Res* 2009;69:4159–66.
 45. Lang SA, Klein D, Moser C, Gaumann A, Glockzin G, Dahlke MH, et al. Inhibition of heat shock protein 90 impairs epidermal growth factor-mediated signaling in gastric cancer cells and reduces tumor growth and vascularization in vivo. *Mol Cancer Ther* 2007;6:123–32.
 46. Wang Z, Zhou J, Fan J, Qiu SJ, Yu Y, Huang XW, et al. Effect of rapamycin alone and in combination with sorafenib in an orthotopic model of human hepatocellular carcinoma. *Clin Cancer Res* 2008;14:5124–30.
 47. Saxena NK, Fu PP, Nagalingam A, Wang J, Handy J, Cohen C, et al. Adiponectin modulates C-jun N-terminal kinase and mammalian target of rapamycin and inhibits hepatocellular carcinoma. *Gastroenterology* 2010;139:762–73, 1773.
 48. Woodrum C, Nobil A, Dabora SL. Comparison of three rapamycin dosing schedules in *A/J Tsc2^{+/-}* mice and improved survival with angiogenesis inhibitor or asparaginase treatment in mice with subcutaneous tuberous sclerosis related tumors. *J Transl Med* 2010;8:14.
 49. Wysocki PJ. mTOR in renal cell cancer: modulator of tumor biology and therapeutic target. *Expert Rev Mol Diagn* 2009;9:231–41.
 50. Liu M, Howes A, Lesperance J, Stallcup WB, Hauser CA, Kadoya K, et al. Antitumor activity of rapamycin in a transgenic mouse model of ErbB2-dependent human breast cancer. *Cancer Res* 2005;65:5325–36.
 51. Namba R, Young LJ, Abbey CK, Kim L, Damonte P, Borowsky AD, et al. Rapamycin inhibits growth of premalignant and malignant mammary lesions in a mouse model of ductal carcinoma in situ. *Clin Cancer Res* 2006;12:2613–21.
 52. Blando J, Portis M, Benavides F, Alexander A, Mills G, Dave B, et al. PTEN deficiency is fully penetrant for prostate adenocarcinoma in C57BL/6 mice via mTOR-dependent growth. *Am J Pathol* 2009;174:1869–79.
 53. Seager CM, Puzio-Kuter AM, Patel T, Jain S, Cordon-Cardo C, McKiernan J, et al. Intravesical delivery of rapamycin suppresses tumorigenesis in a mouse model of progressive bladder cancer. *Cancer Prev Res* 2009;2:1008–14.
 54. Huh WW, Skapek SX. Childhood rhabdomyosarcoma: new insight on biology and treatment. *Curr Oncol Rep* 2010;12:402–10.
 55. Yan Y, Lu Y, Wang M, Vikis H, Yao R, Wang Y, et al. Effect of an epidermal growth factor receptor inhibitor in mouse models of lung cancer. *Mol Cancer Res* 2006;4:971–81.
 56. Liang MC, Ma J, Chen L, Kozlowski P, Qin W, Li D, et al. TSC1 loss synergizes with KRAS activation in lung cancer development in the mouse and confers rapamycin sensitivity. *Oncogene* 2010;29:1588–97.
 57. Stelzer MK, Pitot HC, Liem A, Lee D, Kennedy GD, Lambert PF. Rapamycin inhibits anal carcinogenesis in two preclinical animal models. *Cancer Prev Res* 2010;3:1542–51.
 58. McCampbell AS, Harris HA, Crabtree JS, Winneker RC, Walker CL, Broadus RR, et al. Loss of inhibitory insulin receptor substrate-1 phosphorylation is an early event in mammalian target of rapamycin-dependent endometrial hyperplasia and carcinoma. *Cancer Prev Res* 2010;3:290–300.

Cancer Prevention Research

Rapamycin and mTORC1 Inhibition in the Mouse: Skin Cancer Prevention

Mohammad Athar and Levy Kopelovich

Cancer Prev Res 2011;4:957-961.

Updated version	Access the most recent version of this article at: http://cancerpreventionresearch.aacrjournals.org/content/4/7/957
Supplementary Material	Access the most recent supplemental material at: http://cancerpreventionresearch.aacrjournals.org/content/suppl/2011/06/28/4.7.957.DC3

Cited articles	This article cites 57 articles, 27 of which you can access for free at: http://cancerpreventionresearch.aacrjournals.org/content/4/7/957.full#ref-list-1
Citing articles	This article has been cited by 4 HighWire-hosted articles. Access the articles at: http://cancerpreventionresearch.aacrjournals.org/content/4/7/957.full#related-urls

E-mail alerts	Sign up to receive free email-alerts related to this article or journal.
Reprints and Subscriptions	To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org .
Permissions	To request permission to re-use all or part of this article, use this link http://cancerpreventionresearch.aacrjournals.org/content/4/7/957 . Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.